

**Bioorthogonal non-canonical
amino acid tagging
- BONCAT -**

First, some definitions

bioorthogonal

non-interacting with cellular functionalities

non-canonical

synthetic, not part of biological machinery

Click chemistry

complete conversion of reagents to single product

+ mild conditions

+ very fast

+ in water

Activity assays on individual cell level

DNA

rRNA **mRNA** **tRNA**

```
graph TD; DNA --> rRNA; DNA --> mRNA; DNA --> tRNA; mRNA -.-> proteins; proteins -.-> regulatory; proteins -.-> structural; proteins -.-> metabolic; regulatory -.-> metabolic; proteins -.-> biomass; metabolic -.-> biomass;
```

RNA-FISH

proteins

regulatory **structural**

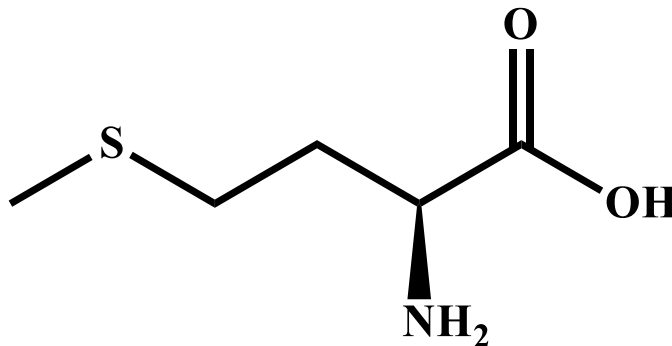
metabolic

BONCAT

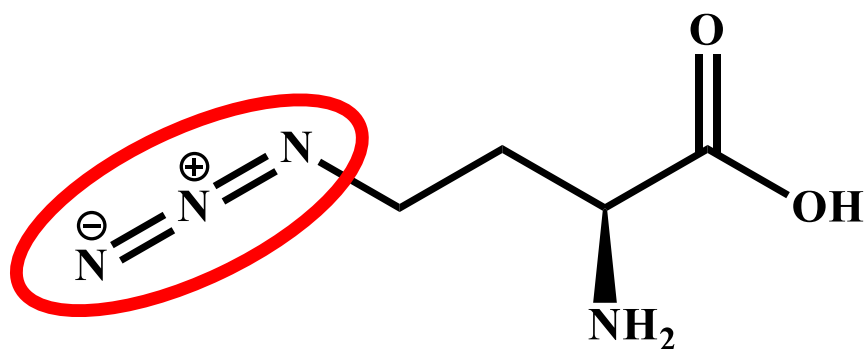
**build-up of biomass
& growth**

isotopic labeling

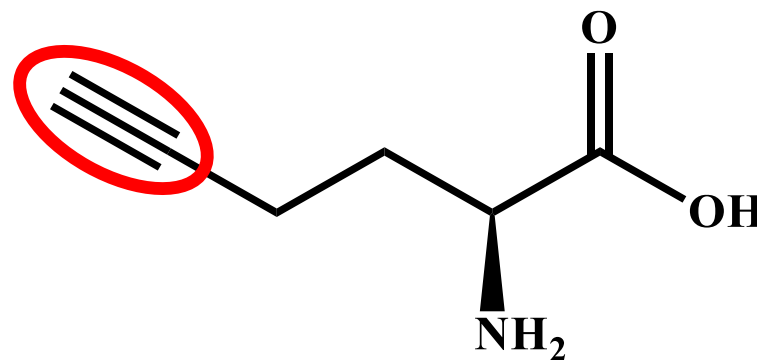
Bioorthogonal non-canonical amino acids



L-Methionine
Met



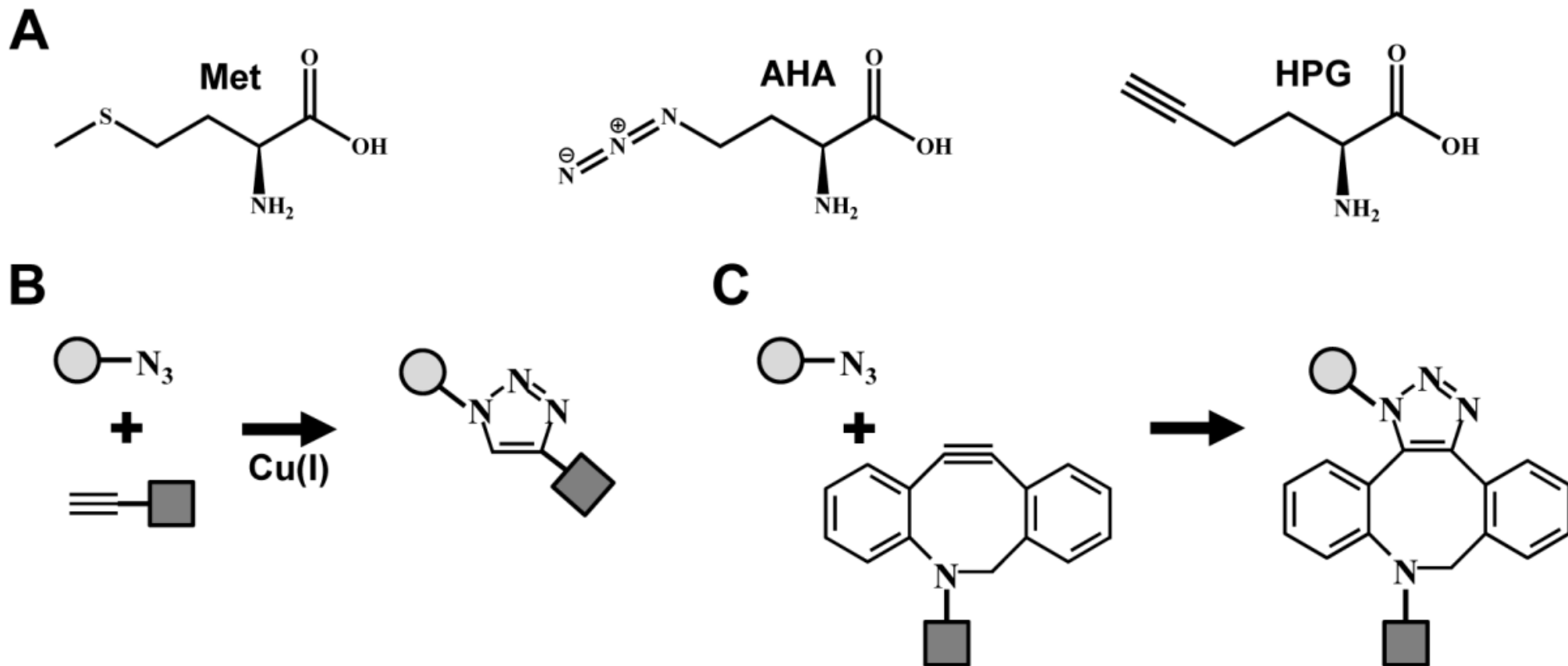
L-Azidohomoalanine
AHA



L-Homopropargylglycine
HPG

synthetic amino acids
incorporate into new proteins instead of Met

Azide-alkyne click reactions

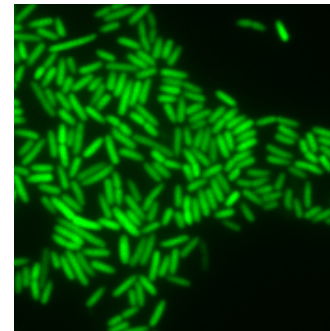
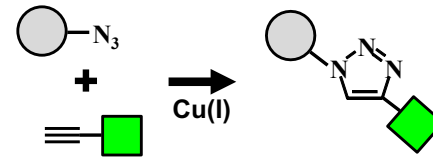
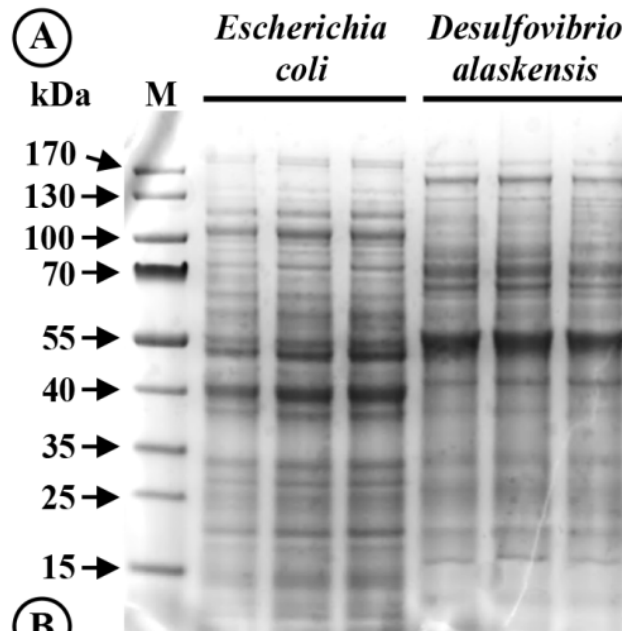


A. Structures of Met and its surrogates AHA and HPG, which compete with Met during translation.

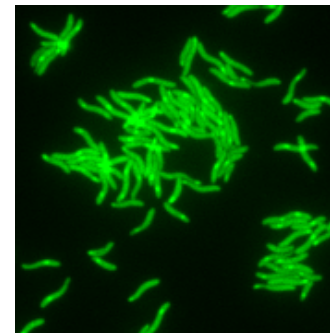
B. In Cu(I)-catalyzed click chemistry an azide group (N_3) is linked to a terminal alkyne residue, yielding a triazole conjugate.

C. Strain-promoted click chemistry allows the copper-less conjugation of an azide group (N_3) with a cyclo-octyne-carrying molecule, yielding a triazole conjugate.

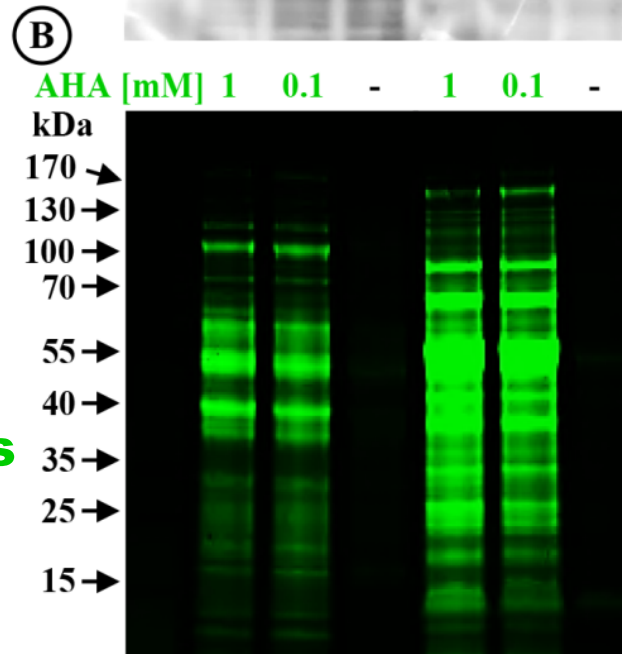
Incorporation into newly made proteins



E. coli
respiring glucose

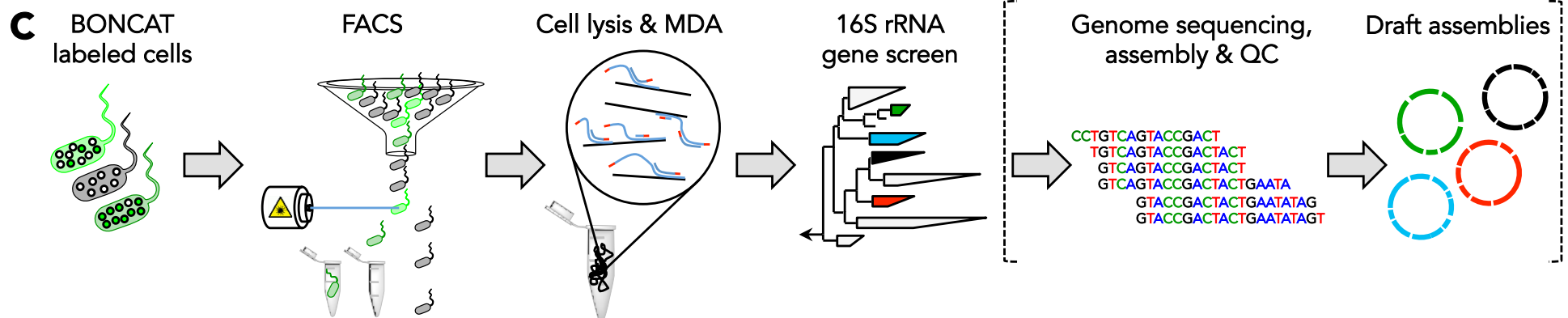
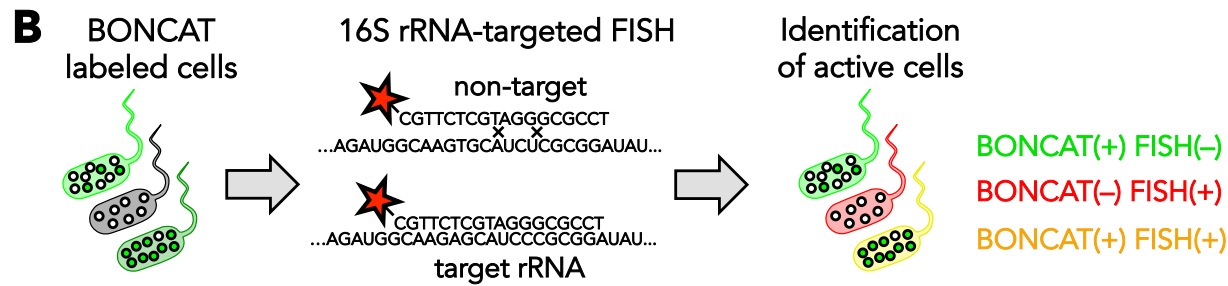
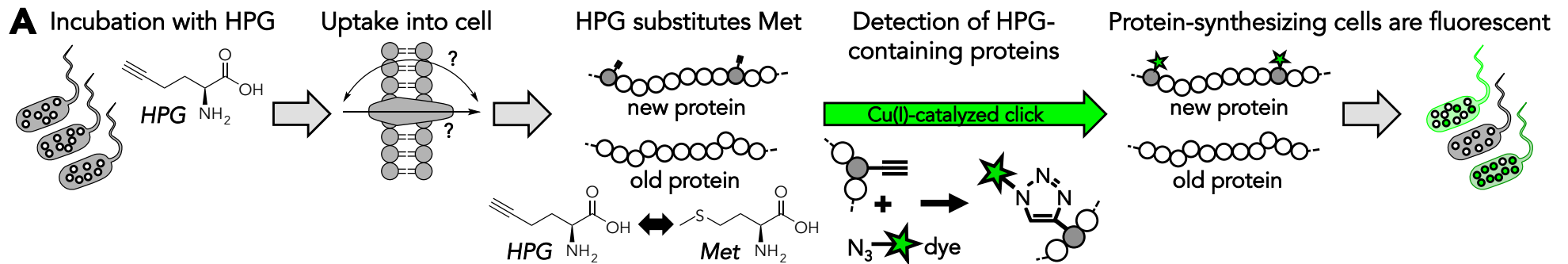


D. alaskensis
sulfate reducer



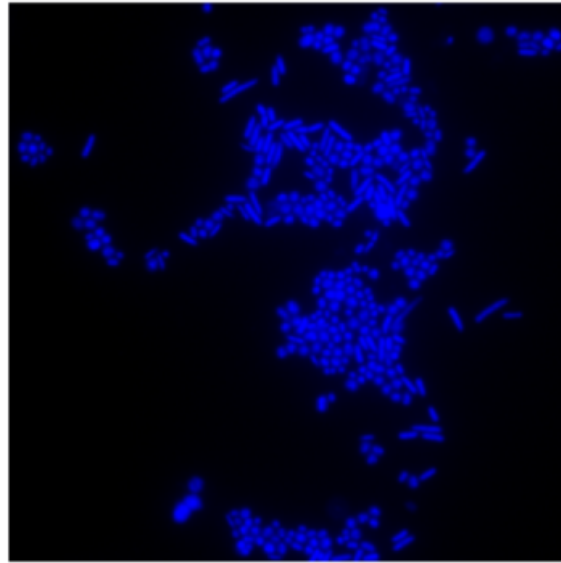
new proteins

Visualizing, identifying, and sorting translationally active microbes

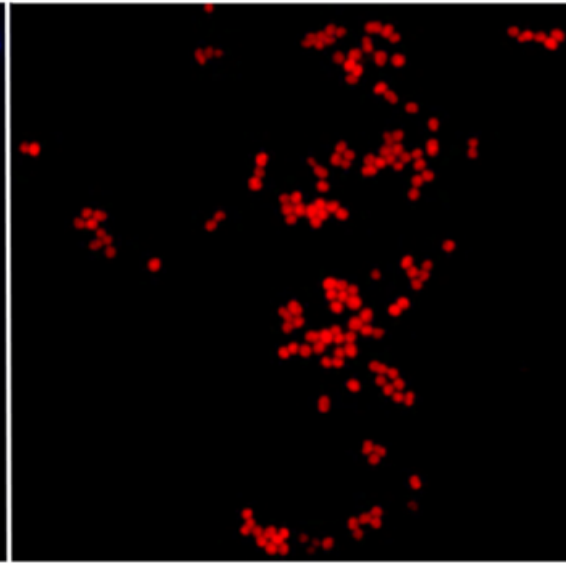


Identification of translationally active cells

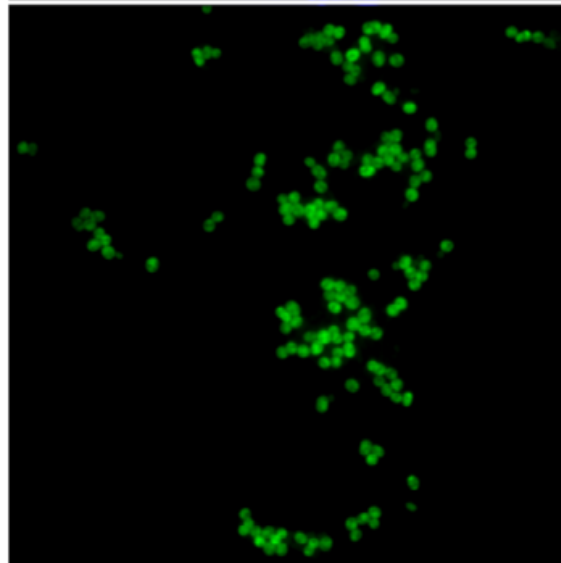
**DAPI
(DNA)**



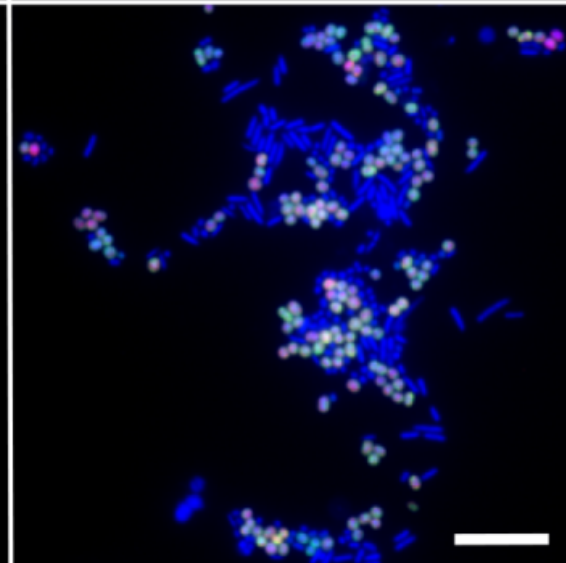
***γ-Proteo WF-1*
(rRNA)**



**BONCAT
(new proteins)**



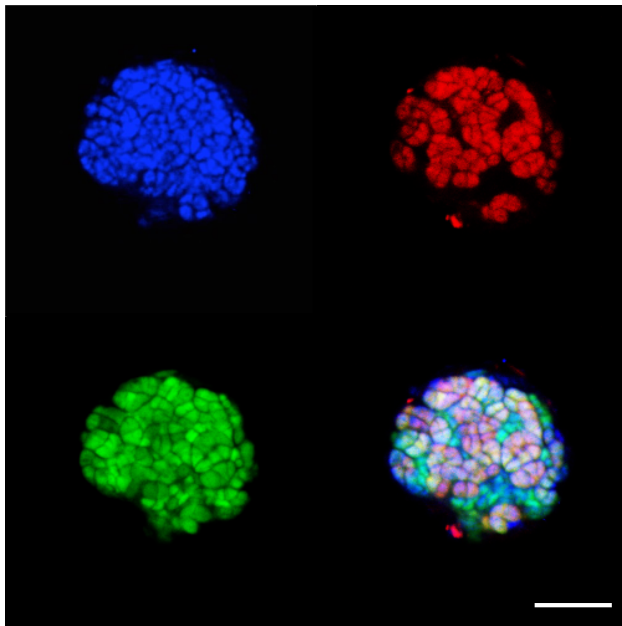
overlay



Bar = 10 μm

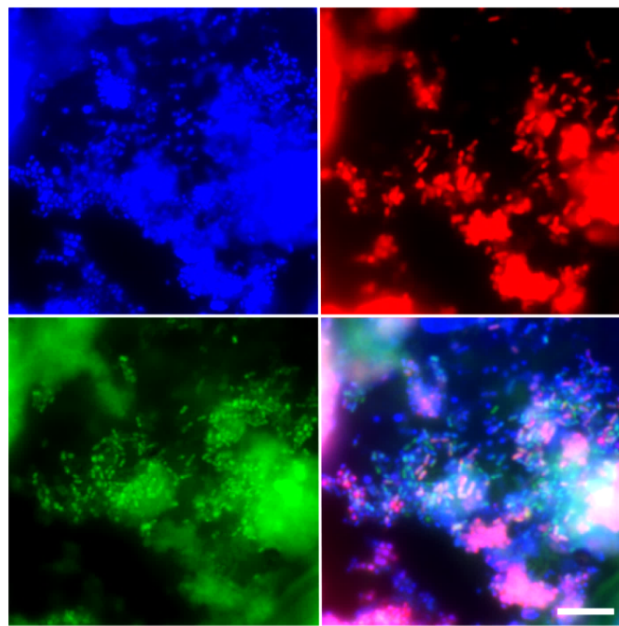
BONCAT-FISH of uncultured microbes

Arch915



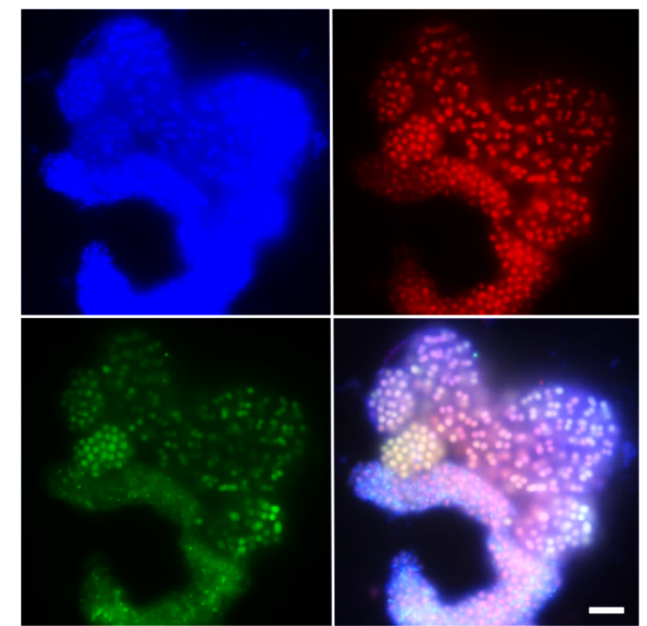
**Methane seep
ANME-SRB consortium**

EUB338 I-III



Tongue biofilm and saliva

Gam42a + competitor



**Freshwater from Lily
pond on Caltech campus**

**DAPI
(DNA)**

**BONCAT
(new proteins)**

**FISH
(rRNA)**

Overlay

Hatzenpichler *et al.*, 2014; Hatzenpichler *et al.*, 2015; Hatzenpichler *et al.*, 2016

Visualizing new proteins *in situ*

**generally applicable
(works for all taxonomies and
physiologies tested so far)**

**detectable after 2%
of generation time**

**FISH-BONCAT links function
and identity of a cell**

**BONCAT correlates with
 $^{15}\text{NH}_3$ incorporation (nanoSIMS)**

**no change in protein expression
(Bagert *et al.*, 2014)**

Hatzenpichler lab



Hatzenpichler *et al.*, 2014

Limitations and advantages of BONCAT-FISH

uptake and incorporation

Methionine-rich samples are tough

hard to quantitate amount of new proteins in uncultured cells

potential for cell inactivation or community shifts

links cellular identity and function

fluorescence-based *in situ* activity studies

metabolic screening

activity-based cell-sorting

fast + highly selective + cheap + easily available

1 h azide-alkyne ~\$500 epi-scope

BONCAT in environmental microbiology (as of Nov. 2017)

Pasulka AL et al.

Interrogating marine virus-host interactions and elemental transfer with BONCAT and nanoSIMS-based methods
Environ Microbiol, DOI: 10.1111/1462-2920.13996 (accepted) 2017

- *first application of BONCAT to viruses; estimate of marine viral production rates by BONCAT and nanoSIMS*

Hatzenpichler R et al.

Visualizing *in situ* translational activity for identifying and sorting slow-growing archaeal-bacterial consortia

Proc Natl Acad Sci USA, 113: E4069-E4078 (2016)

- application of BONCAT-FISH and BONCAT-FACS to ANME-SRB consortia from three methane seep sediments; development of activity-based cell-sorting via bioorthogonal labeling

Hatzenpichler R and Orphan VJ

Detection of protein-synthesizing microorganisms in the environment via bioorthogonal non-canonical amino acid tagging (BONCAT)

Book chapter for Hydrocarbon and Lipid Microbiology Protocols, Vol. 7: Single-cell and single-molecule methods
Springer Protocols Handbooks, doi 10.1007/8623_2015_61 (2015)

- *description of how to design and perform BONCAT-experiments using AHA and HPG*

Samo TJ et al.

Broad distribution and high proportion of protein synthesis active marine bacteria revealed by click chemistry at the single cell level

Front Mar Sci, doi: 10.3389/fmars.2014.00048 (2014)

- *application of BONCAT to seawater; correlation of BONCAT with MAR*

Hatzenpichler R et al.

***In situ* visualization of newly synthesized proteins in environmental microbes using amino acid tagging and click chemistry**

Environ Microbiol, 16: 2568-2590 (2014)

- *first application of BONCAT to uncultured microbes in the environment; development of BONCAT-FISH; correlation of BONCAT with nanoSIMS*