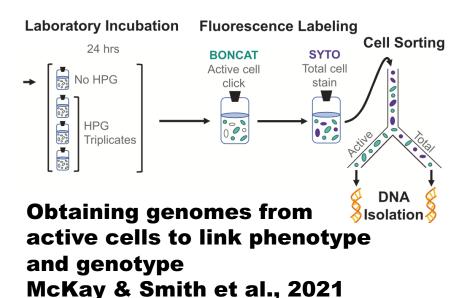
Bioorthogonal non-canonical amino acid tagging - BONCAT -

Research examples

100 4

50

25



Active lung microbiome of

cystic fibrosis patients

50

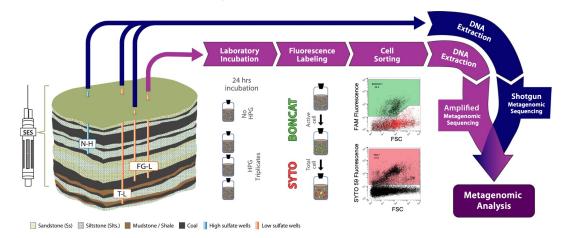
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25

Sort positive

Metagenomes of protein-synthesizing cells from the subsurface

Schweitzer et al., 2022



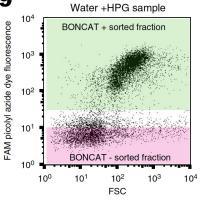
Sorting and identifying active fraction of cells in soil with BONCAT-FACS

50

Couradeau et al., 2019

16S rRNA gene sequencing

+ DADA2 analysis



BONCAT is a **Next-generation** physiology approach

<u>Definition:</u> ...any combination of techniques that analyze the phenotype of an individual cell in a microbiome in a non-destructive way, which enables the physical separation of this cell based solely on its phenotype for subsequent, downstream applications

Specifically, BONCAT is a type of substrate analog probing (SAP)

SAP uses molecules that carry either a fluorescence tag or a side group amenable to azide-alkyne click chemistry to obtain information on the overall biosynthetic activity or specific enzymatic function of the cell.

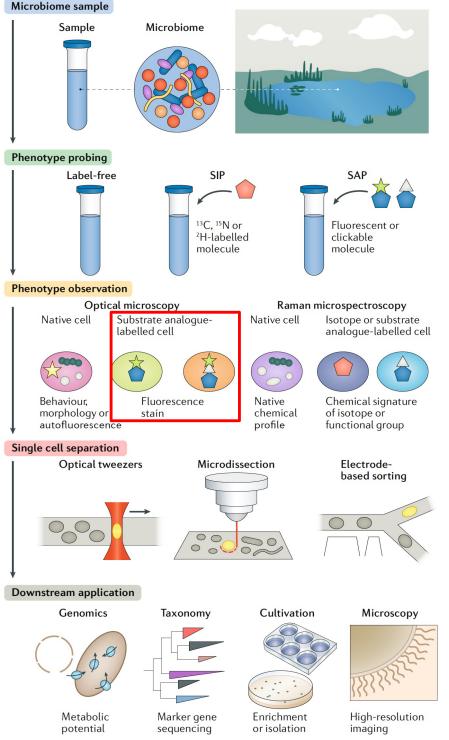


Fig. 2 | Next-generation physiology workflow to study microorganisms.

First, some definitions

bioorthogonal non-interacting with cellular functionalities

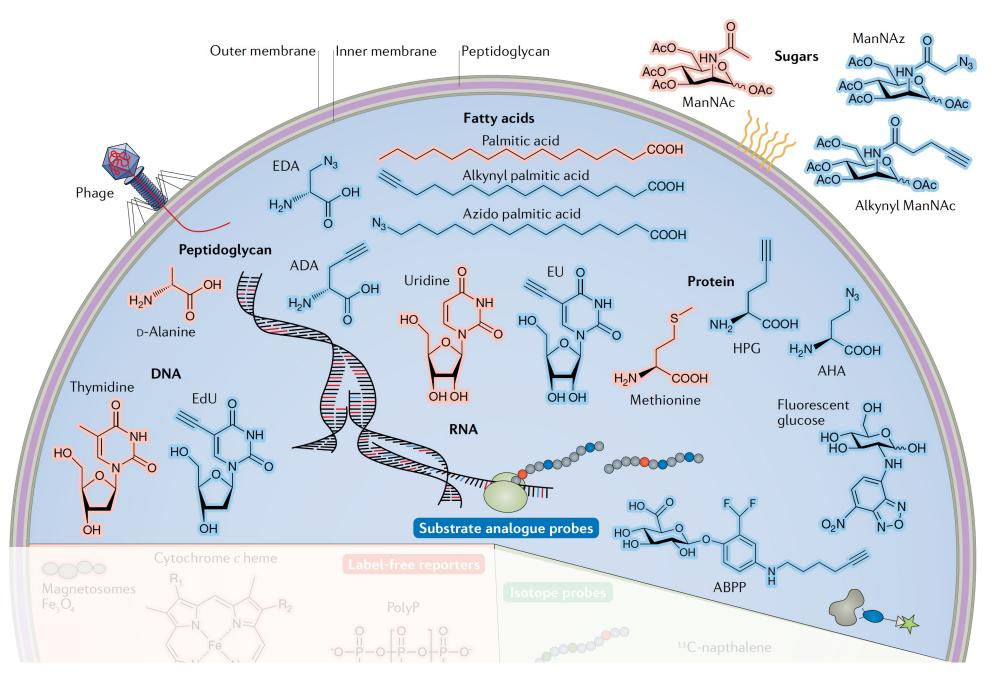
non-canonical synthetic, not part of biological machinery

Click chemistry

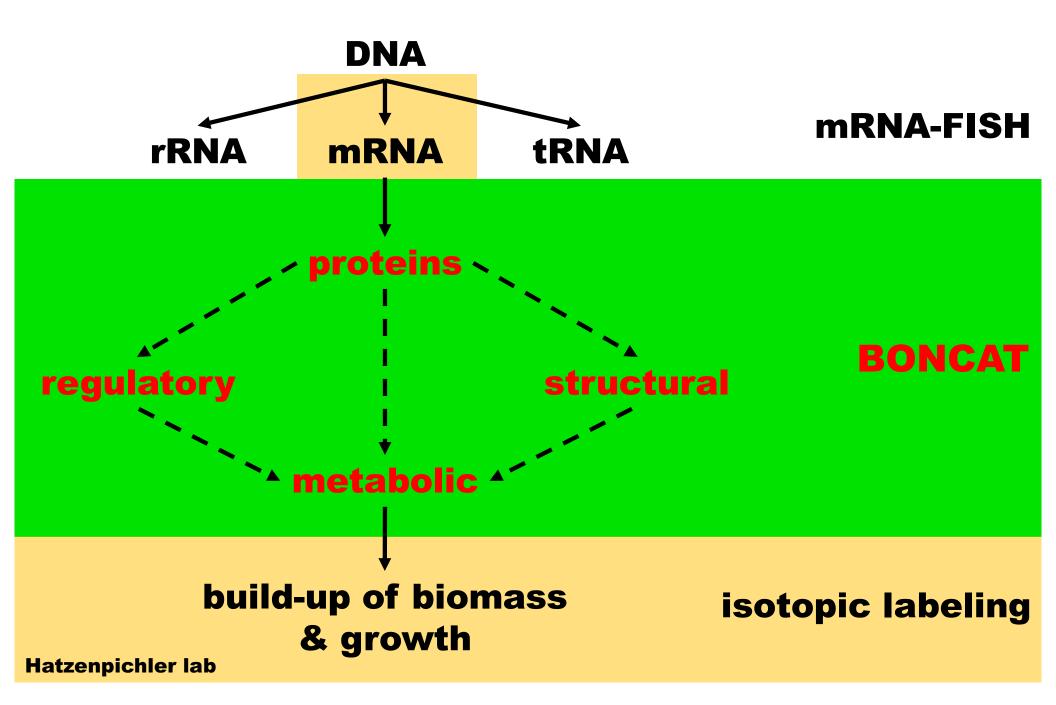
complete conversion of reagents to single product

- + mild conditions
 - + very fast
 - + in water

Examples for clickable substrate analogs



Activity assays on individual cell level



Limitations and advantages of BONCAT

uptake and incorporation differ between species

Methionine-rich samples are tough (need mM concentrations)

hard to quantitate amount of new proteins in uncultured cells

potential for cell inactivation or community shifts (can be controlled!)

links cellular identity and function (Hatzenpichler et al. 2014)

fluorescence-based in situ activity studies (Hatzenpichler et al. 2014)

metabolic screening (Reichart et al. 2020)

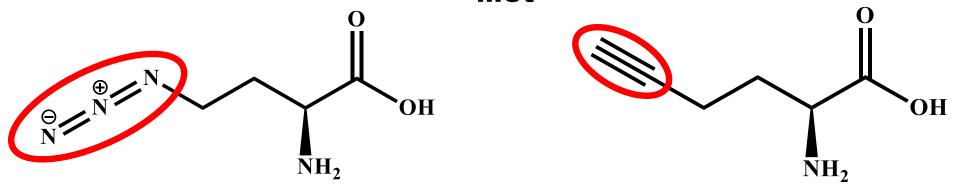
activity-based cell-sorting (Hatzenpichler et al. 2016)

fast + highly selective + cheap + easily available

1 h azide-alkyne ~\$500 epifluorescence-scope

Non-canonical amino acids

L-Methionine Met

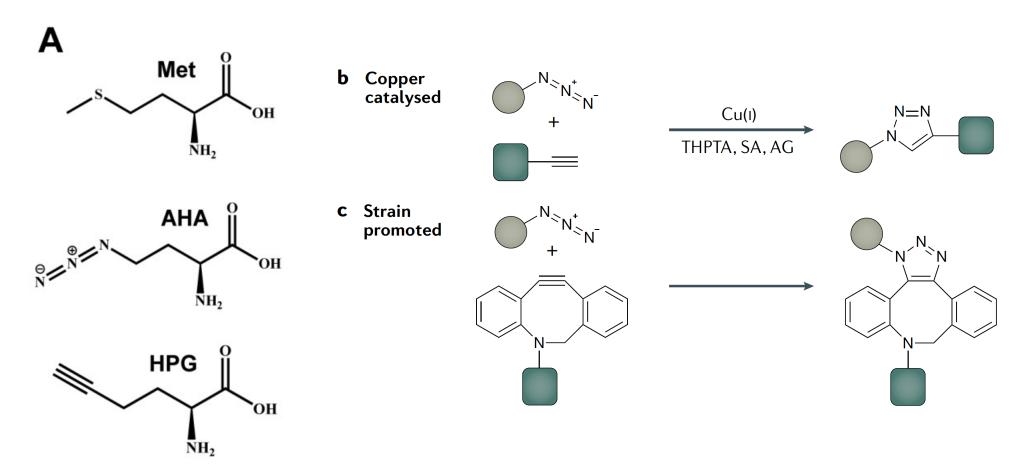


L-AzidohomoalanineAHA

L-Homopropargylglycine HPG

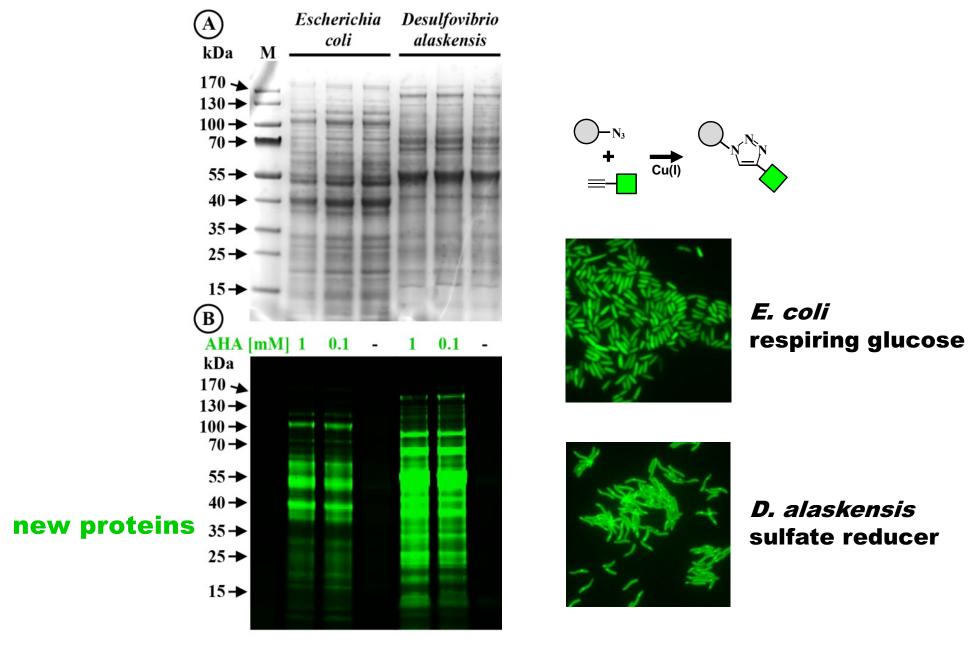
synthetic amino acids incorporate into new proteins instead of Met

Azide-alkyne click chemistry reactions



- A. Structures of Met and its surrogates AHA and HPG, which compete with Met during translation.
- B. In Cu(I)-catalyzed click chemistry an azide group (N_3) is linked to a terminal alkyne residue, yielding a triazole conjugate.
- C. Strain-promoted click chemistry allows the copper-less conjugation of an azide group (N₃) with a cyclo-octyne-carrying molecule, yielding a triazole conjugate.

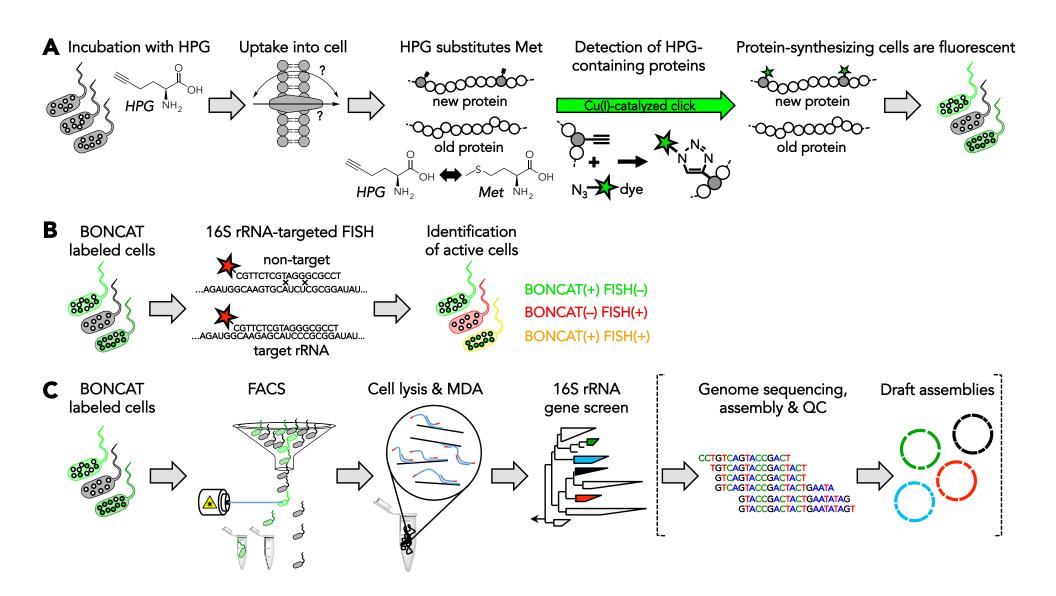
Incorporation into newly made proteins



Hatzenpichler lab

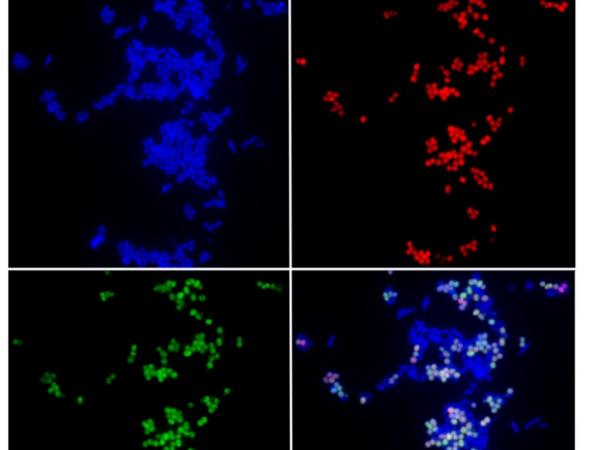
Hatzenpichler et al., 2014

Visualizing, identifying, and sorting translationally active microbes



Identification of translationally active cells

DAPI (DNA)



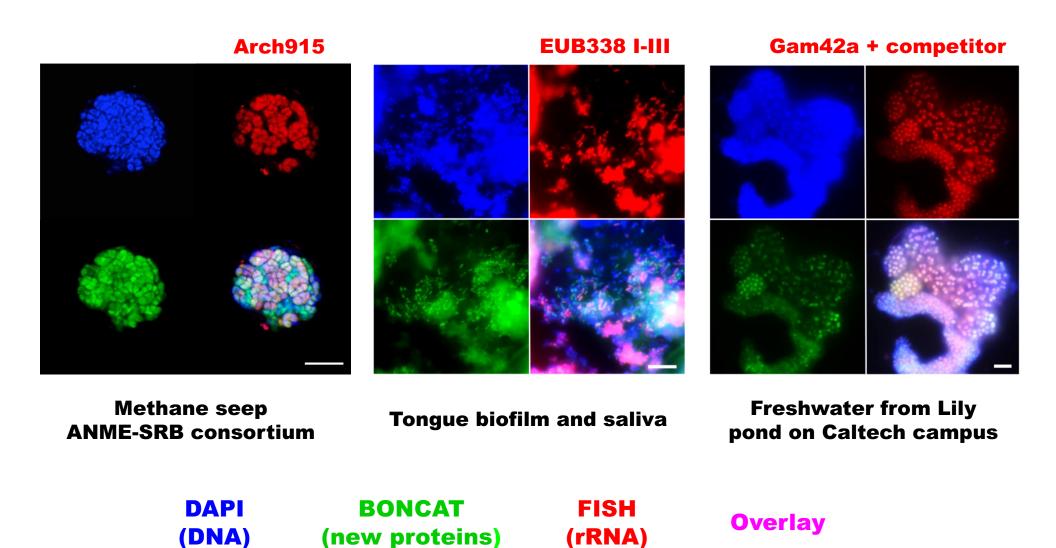
y-proteobacterium WF-1 (16S rRNA)

BONCAT (new proteins)

overlay

Bar = $10 \mu m$

BONCAT-FISH of uncultured microbes



Hatzenpichler et al., 2014; Hatzenpichler et al., 2015; Hatzenpichler et al., 2016

Hatzenpichler lab

Visualizing new proteins in situ

generally applicable (works for all taxonomies and physiologies tested so far)

in *E. coli*, detectable after 2% of generation time

In environmental samples, cells are detectable after 3.3-7% of generation time

FISH-BONCAT links function and identity of a cell

BONCAT correlates with other growth Proxies (e.g., SIP-nanoSIMS, MAR)

Hatzenpichler et al., 2014 and 2016



Hatzenpichler et al., 2014

Hatzenpichler R et al. *In situ* visualization of newly synthesized proteins in environmental microbes using amino acid tagging and click chemistry

Environ Microbiol, 16: 2568-2590 (2014)

first application of BONCAT to uncultured microbes; development of BONCAT-FISH; correlation of BONCAT with nanoSIMS

Samo TJ et al. Broad distribution and high proportion of protein synthesis active marine bacteria revealed by click chemistry at the single cell level

Front Microbiol, 1: 48 (2014)

application of BONCAT to seawater; correlation of BONCAT with microautoradiography

Hatzenpichler R and Orphan VJ Detection of protein-synthesizing microorganisms in the environment via bioorthogonal non-canonical amino acid tagging (BONCAT)

Book chapter for Hydrocarbon and Lipid Microbiology Protocols, Springer Protocols Handbooks, doi 10.1007/8623_2015_61 (2015)

description of how to design and protocols for how to perform BONCAT-experiments using AHA and HPG

Hatzenpichler R et al. Visualizing *in situ* translational activity for identifying and sorting slow-growing archaeal-bacterial consortia Proc Natl Acad Sci USA, 113: E4069-E4078 (2016)

development of activity-based cell-sorting via bioorthogonal labeling (BONCAT-FACS); applied BONCAT-(CARD)FISH and BONCAT-FACS to deep-sea sediment consortia catalyzing the anaerobic oxidation of methane with sulfate

Leizeaga et al. Using Click-Chemistry for Visualizing *in Situ* Changes of Translational Activity in Planktonic Marine Bacteria Front Microbiol, 8: 2360 (2017)

Pasulka AL et al. Interrogating marine virus-host interactions and elemental transfer with BONCAT and nanoSIMS-based methods Environ Microbiol, 20: 671-692 (2018)

first application of BONCAT to environmental phages and viruses; estimate of marine viral production rates by BONCAT and nanoSIMS

Couradeau et al. Probing the active fraction of soil microbiomes using BONCAT-FACS

Nat Comm, 10: 2770 (2019)

> first application of BONCAT to soil samples; reports that a surprisingly high proportion of soil microbes is translationally active

Sebastian et al. High Growth Potential of Long-Term Starved Deep Ocean Opportunistic Heterotrophic Bacteria Front Microbiol, 10: 760 (2019)

Kjeldsen et al., On the evolution and physiology of cable bacteria

Proc Natl Acad Sci USA, 116 (38) 19116-19125 (2019)

➤ Uses BONCAT to study the activity of individual cells along cable bacteria filaments

Steward et al. Metabolic Implications of Using BioOrthogonal Non-Canonical Amino Acid Tagging (BONCAT) for Tracking Protein Synthesis

Front Microbiol, 11:197 (2020)

> shows that growing E. coli with AHA or HPG alters ~15 % of global mass features (LC-MS). 7 % change if cells are grown on methionine.

Valentini et al. Bioorthogonal non-canonical amino acid tagging reveals translationally active subpopulations of the cystic fibrosis lung microbiota

Nature Comm, 11: 2287 (2020)

Case study that applies BONCAT and BONCAT-FACS to cystic fibrosis patients' lung microbiomes

Lindivat et al. Bioorthogonal Non-canonical Amino Acid Tagging Combined With Flow Cytometry for Determination of Activity in Aquatic Microorganisms

Front Microbiol, 11: 1929 (2000)

Reichart et al. Activity-based cell sorting reveals responses of uncultured archaea and bacteria to substrate amendment The ISME J, 14: 2851-2861 (2020)

> Uses BONCAT-FACS to detect changes in single cell activity of a hot spring microbial community incubated in the presence of various growth substrates or under changing physicochemical conditions

Riva et al. Conversion of Rutin, a Prevalent Dietary Flavonol, by the Human Gut Microbiota Front Microbiol, 11: 585428 (2020)

Taguer et al. Translational activity is uncoupled from nucleic acid content in bacterial cells of the human gut microbiota Gut Microbes, 13: e1903289 (2021)

First application of BONCAT to human gut microbes (stool samples).

Chen et al. Isolating and characterizing translationally active fraction of anammox microbiota using bioorthogonal non-canonical amino acid tagging

Chem Eng J, 418: 129411 (2021)

Michels et al. Amino acid analog induces stress response in marine Synechococcus

Appl Environm Microbiol, DOI: 10.1128/AEM.00200-21 (2021)

Bergkessel and Delavaine. Diversity in Starvation Survival Strategies and Outcomes among Heterotrophic Proteobacteria Microb Physiol, DOI: 10.1159/000516215 (2021)

Marlow et al. Spatially resolved correlative microscopy and microbial identification reveal dynamic depth- and mineral- dependent anabolic activity in salt marsh sediment

Environ Microbiol, doi:10.1111/1462-2920.15667 (2021)

➤ BONCAT, resin embedding, fluorescence microscopy, electron microscopy, and energy dispersive X-ray spectroscopy are combined to reveal the spatial patterning of translationally active cells around specific mineral particles in heterogenous salt marsh sediment

Martinez-Varela et al. Bacterial responses to background organic pollutants in the northeast subarctic Pacific Ocean Environ Microbiol, doi:10.1111/1462-2920.15646 (2021)

Du and Behrens. Tracking de novo protein synthesis in the activated sludge microbiome using BONCAT-FACS Water Research, 205: 117696 (2021)

McKay, Smith et al. Activity-based, genome-resolved metagenomics uncovers key populations and pathways involved in subsurface conversions of coal to methane

The ISME J, https://doi.org/10.1038/s41396-021-01139-x (2021)

> First study to combine BONCAT-FACS with metagenomics of sorted cells, thus directly linking phenotype and genotype of uncultured cells.

Madill et al., Activity-Based Cell Sorting Reveals Resistance of Functionally Degenerate *Nitrospira* during a Press Disturbance in Nitrifying Activated Sludge

mSystems, https://doi.org/10.1128/mSystems.00712-21 (2021)

Taguer et al., Changes in Gut Bacterial Translation Occur before Symptom Onset and Dysbiosis in Dextran Sodium Sulfate-Induced Murine Colitis

mSystems, 6: e00507-21 (2021)

Schweitzer et al., Subsurface hydrocarbon degradation strategies in low- and high-sulfate coal seam communities identified with activity-based metagenomics

Biofilms and Microbiomes, 8:7 https://doi.org/10.1038/s41522-022-00267-2 (2022)

Berjón-Otero et al., Visualization of giant virus particles using BONCAT labeling and STED microscopy bioRxiv, doi: https://doi.org/10.1101/2020.07.14.202192 (2022)

Birnstiel et al., Structure and activity of marine bacterial communities responding to plastic leachates Science of the Total Environment, https://doi.org/10.1016/j.scitotenv.2022.155264

Volland et al. A centimeter-long bacterium with DNA contained in metabolically active, membrane-bound organelles Science, Vol 376, Issue 6600: 1453-1458 (2022)

Wang et al. Detection and Quantification of Antimicrobial-Resistant Cells in Aquatic Environments by Bioorthogonal Noncanonical Amino Acid Tagging

Environ Sci Tech, DOI pending (2022)