**Soil DNA prep**

1. Put 0.5 g of each soil sample into 1.5 ml eppendorf tube with sterile spatula
2. Add 0.5 g 0.5 mm glass beads (sterile) to each
3. Add 600 μl Extraction buffer, vortex at high speed 2 min, spin at 15K, 2 min
4. Pour off supernatant (keep) in new eppendorf tube; dispose of pellet
5. Add equal volume of CHCl3 to supernatant, mix by hand, spin at 8K, 5 min
6. Pipette off supernatant into new eppendorf tube
7. Add equal volume CHCl3, mix by hand, spin at 15K, 5 min
8. Pipette off supernatant into new eppendorf tube
9. Add equal volume of -20C isopropanol, mix by hand
10. Store at -20C overnight or longer
11. Spin at 15K, 10 min, dispose of supernatant, wash pellet with 70% EtOH
12. Quick spin and pour off supernatant; allow pellet to dry
13. Add 1-200 μl TE, pH 8 and resuspend pellet by flicking occasionally
14. Run 15 μl on 0.8 – 1% agarose gel to check DNA quantity, quality

Extraction buffer S Extraction buffer T

50 mM Na-phosphate, pH 8 500 mM Tris-HCl, pH 8

50 mM NaCl 100 mM NaCl

500 mM Tris-HCl, pH 8 10% SDS

5% SDS

Equipment/reagants

1 mm glass beads

Extraction buffer

Spatula(s)

Microfuge

Eppendorf tubes 5 per sample

ClCH3; 91% isopropanol; 70% EtOH; TE pH 8