Clinical Cancer Research

Phase II Study of Nilotinib in Melanoma Harboring KIT Alterations Following Progression to Prior KIT Inhibition

Richard D. Carvajal^{1,2}, Donald P. Lawrence³, Jeffrey S. Weber⁴, Thomas F. Gajewski⁵, Rene Gonzalez⁶, Jose Lutzky⁷, Steven J. O'Day⁸, Omid Hamid⁹, Jedd D. Wolchok^{1,2}, Paul B. Chapman^{1,2}, Ryan J. Sullivan³, Jerrold B. Teitcher¹, Nikhil Ramaiya¹⁰, Anita Giobbie-Hurder¹⁰, Cristina R. Antonescu¹, Michael C. Heinrich¹¹, Boris C. Bastian¹², Christopher L. Corless¹¹, Jonathan A. Fletcher¹⁰, and F. Stephen Hodi¹⁰

Abstract

Purpose: Although durable responses can be achieved with tyrosine kinase inhibitors such as imatinib in melanomas harboring KIT mutations, the efficacy of alternative inhibitors after progression to imatinib and the activity of these agents on brain metastases are unknown.

Experimental Design: We conducted a phase II study of nilotinib 400 mg twice a day in two cohorts of patients with melanomas harboring KIT mutations or amplification: (A) those refractory or intolerant to a prior KIT inhibitor; and (B) those with brain metastases. The primary endpoint was 4-month disease control rate. Secondary endpoints included response rate, time-to-progression (TTP), and overall survival (OS). A Simon two-stage and a single-stage design was planned to assess for the primary endpoint in cohorts A and B, respectively.

Results: Twenty patients were enrolled and 19 treated (11 in cohort A; 8 in cohort B). Three patients on cohort A [27%; 95% confidence interval (CI), 8%–56%] and 1 on cohort B (12.5%; 90% CI, 0.6%–47%) achieved the primary endpoint. Two partial responses were observed in cohort A (18.2%; 90% CI, 3%–47%); none were observed in cohort B. The median TTP and OS was 3.3 (90% CI, 2.1–3.9 months) and 9.1 months (90% CI, 4.3–14.2 months), respectively, in all treated patients.

Conclusions: Nilotinib may achieve disease control in patients with melanoma harboring KIT alterations and whose disease progressed after imatinib therapy. The efficacy of this agent in KIT-altered melanoma with brain metastasis is limited. *Clin Cancer Res;* 21(10); 2289–96. ©2015 AACR.

Introduction

Alterations in the *KIT* proto-oncogene define one unique molecular subset of melanoma. Mutations and amplification of *KIT* are observed in 3% of all melanomas, and are more common in disease arising from mucosal, acral, or chronically sun-damaged surfaces (1). The mutations identified are, in most cases, substitution mutations mutually exclusive of BRAF and NRAS

Corresponding Author: F. Stephen Hodi, Dana-Farber Cancer Institute, 450 Brookline Avenue, Boston, MA 02215. Phone: 617-632-5053; Fax: 617-582-7992; E-mail: Stephen hodi@dfci.harvard.edu

doi: 10.1158/1078-0432.CCR-14-1630

©2015 American Association for Cancer Research.

www.aacrjournals.org

mutations and often affect the juxtamembrane or kinase domains of KIT, leading to constitutive activation of KIT tyrosine kinase activity.

The clinical activity of KIT inhibition in those melanomas driven by KIT alterations has been reported in patients treated with agents such as imatinib (2–4), dasatinib (5), sorafenib (6), and sunitinib (7), with efficacy observed in prospective trials of imatinib (8–10) and sunitinib (11). Despite the clinical benefit achieved with KIT inhibition in select patients with melanoma harboring KIT mutations, most patients ultimately experience disease progression. Failure of these agents has been observed within the brain (12), which may be related to the frequent development of brain metastases in patients with advanced melanoma, as well as the limited central nervous system (CNS) penetration of many small-molecule kinase inhibitors.

Secondary resistance to KIT inhibition in patients with gastrointestinal stromal tumors (GIST), a disease characterized by activating deletions or insertions in *KIT*, is caused primarily by the development of secondary KIT mutations commonly affecting the tyrosine kinase domains (13). There can additionally be outgrowth of resistant subclones present at baseline that are selected during KIT inhibitor therapy. In GIST, the use of alternative KIT tyrosine kinase inhibitors after progression on imatinib, including sunitinib (14), sorafenib (15), and regorafenib (16), has proven beneficial; however, the efficacy of sequential KIT inhibitors in melanoma is unknown.



¹Memorial Sloan Kettering Cancer Center, New York, New York. ²Weill Medical College of Cornell University, New York, New York. ³Massachusetts General Hospital, Boston, Massachusetts. ⁴H. Lee Moffitt Cancer Center, Tampa, Florida. ⁵The University of Chicago, Chicago, Illinois. ⁶The University of Colorado Cancer Center, Aurora, Colorado. ⁷Mount Sinai Comprehensive Cancer Center, Miami Beach, Florida. ⁸Beverly Hills Cancer Center, Beverly Hills, California. ⁹Angeles Clinic and Research Institute, Los Angeles, California. ¹⁰Dana Farber Cancer Institute, Boston, Massachusetts. ¹¹Oregon Health and Science University, Portland, Oregon. ¹²The University of California San Francisco, San Francisco, California

Note: Supplementary data for this article are available at Clinical Cancer Research Online (http://clincancerres.aacrjournals.org/).

Translational Relevance

Although significant clinical benefit can be achieved with KIT inhibition in a subset of patients with melanoma driven by activating alterations in KIT, the development of secondary resistance is common. In this phase II study of nilotinib 400 mg twice a day, 3 of 11 patients with melanomas harboring KIT mutations or amplification who were refractory to a prior KIT inhibitor had disease control lasting 4 months or greater, with 2 achieving a partial response to therapy. One of 8 patients with melanomas metastatic to the brain harboring KIT mutations or amplification had disease control lasting 4 months or greater, with none achieving a radiographic response. We conclude that nilotinib can achieve disease control in a subset of patients with melanoma harboring KIT alterations after progression on a prior tyrosine kinase inhibitor; however, the efficacy of this agent in KIT-altered melanoma with brain metastasis is limited.

Nilotinib (Tasigna, AMN107) is a tyrosine kinase inhibitor structurally derived from imatinib that is approved in the United States for the treatment of chronic and accelerated phase Philadelphia chromosome-positive chronic myelogenous leukemia in patients resistant or intolerant to prior therapy with imatinib. Nilotinib binds to and inhibits the kinase domain of ABL/BCR-ABL and of the DDR, KIT, PDGF, and several EPH receptor kinases with greater potency than imatinib (17, 18), and maintains activity against a range of exon 9, 11, and 13 KIT mutations (19). We conducted a phase II trial of nilotinib in patients with melanoma harboring KIT aberrations who experienced disease progression or intolerance to a prior KIT inhibitor. Given the frequent complication of brain metastases in patients with this disease and the potential for second-generation inhibitors of KIT to have activity within the CNS (20), a cohort of patients with brain metastases was included.

Materials and Methods

Study design and objectives

The primary objective was to assess the efficacy of nilotinib in patients with metastatic melanoma arising from acral, mucosal, or chronically sun-damaged surfaces characterized by mutations or amplification of KIT after demonstration of disease progression or intolerance to a prior KIT tyrosine kinase inhibitor. Secondary objectives included efficacy assessment of nilotinib in patients with advanced KIT-mutant melanoma and CNS metastases. Tumor samples from all patients were prospectively tested for KIT mutation or amplification by qPCR) or FISH as previously described (8, 10).

Patients who met eligibility criteria received nilotinib 400 mg by mouth twice daily. Safety evaluations, including clinical and laboratory assessments, were conducted at baseline, every week for 4 weeks, every 2 weeks for 4 weeks, every 4 weeks for 28 weeks, and then every 3 months subsequently. Adverse event severity was graded using the NCI Common Terminology Criteria for Adverse Events, v3.0. Tumor response was measured radiographically every 8 weeks for 32 weeks and every 12 weeks subsequently using RECIST 1.0 criteria, and included brain imaging for those with CNS involvement. Patients remained on study until the time of progression or the development of unacceptable toxicity not manageable with dose modification.

The primary endpoint was the proportion of patients who were alive and without progression of disease 4 months after beginning treatment with nilotinib. Secondary endpoints included best overall response rate (BORR), time-to-progression (TTP), overall survival (OS), and tolerability.

Patients

Patients were enrolled from eight academic medical centers between January 23, 2009 and June 14, 2011. Eligible patients had advanced melanoma harboring a KIT mutation or amplification and arising from acral, mucosal, or chronically sun-damaged surfaces, as documented by the presence of solar elastosis. Patients without CNS metastases were enrolled onto cohort A and must have experienced disease progression or intolerance to one or more KIT tyrosine kinase inhibitors. Intolerance was defined as drug discontinuation due to grade-2 events persisting for one month or longer, or any grade-3 or grade-4 rash, fluid retention, cardiopulmonary events, thrombocytopenia, liver function abnormalities, or diarrhea that persisted despite optimal supportive care measures. Patients with measureable CNS disease harboring a KIT mutation were enrolled onto cohort B and did not require prior therapy for eligibility. For those who received prior radiotherapy for CNS disease, progression was required in previously treated lesions or new lesions must have developed.

Other key inclusion criteria included age greater than 18 years, life expectancy greater than 3 months, Eastern Cooperative Oncology Group performance status of zero, one, or two, measurable disease by Response Evaluation Criteria in Solid Tumors (RECIST) v1.0, and adequate organ function. Exclusion criteria included prior therapy with nilotinib and clinically significant heart disease. All patients provided written-informed consent before initiating study procedures. The study was reviewed and approved by Institutional Review Boards at all participating centers.

Trial design

Cohort A used an optimal Simon, two-stage design with 87% power to compare a null disease control rate (DCR) of 5% with an alternative of 25%, with a one-sided type-I error of 7.5%. The target sample size was 28 patients, of whom 25 were expected to be evaluable for outcome. In the first stage, 13 evaluable patients would be assessed. If 2 or more patients achieved 4-month disease control, an additional 12 evaluable patients would be assessed in the second stage. If 3 or more of 25 achieved 4-month disease control, then nilotinib would be considered promising in this disease setting. A second feasibility cohort of 10 patients (cohort B) was added after the study began to estimate the 4-month DCR in patients with advanced, KIT-mutated melanoma and CNS metastases. Nilotinib would be of interest in this cohort if at least 2 of 10 patients achieved 4-month disease control.

Statistical analysis

Baseline patient characteristics and adverse events were summarized using descriptive methods. Adverse events were reported as the most severe manifestation of each event category during any cycle of treatment. Four-month DCR was defined as the proportion of treated patients with a complete or partial response (PR), or stable disease (SD) per RECIST 1.0 after 4 months of therapy. BORR was defined as the proportion of treated patients with either complete or PR (per RECIST) as best response to therapy. The number of treated patients in each cohort was the denominator for estimates of DCR and BORR. TTP was defined as the time from initiation of nilotinib to the date of progression or last follow-up. OS was defined as the time from initiation of nilotinib to the date of death or last follow-up. Four-month DCR and BORR are presented with 90% exact binomial confidence intervals. TTP and OS are presented using the method of Kaplan-Meier, with pointwise 90% confidence intervals (CI) estimated using log(-log(survival)) methodology.

Role of the funding source

Dr. Hodi developed the original study design and was responsible for the IND. Novartis provided investigational drug in addition to funding, and was involved in study design that was developed in conjunction with the authors. The study sponsor had no role in the data collection, the data analysis, data interpretation, writing of the report, or the decision to submit for publication.

Results

Patient characteristics

Twenty patients were enrolled (11 in cohort A and 9 in cohort B) and 19 treated on this study (11 in cohort A and 8 in cohort B). One patient who enrolled in cohort B withdrew consent before receiving study therapy. With the completion of a series of studies of imatinib and other agents targeting KIT in patients with melanoma harboring KIT alterations, enrollment to second-line trials became increasingly challenging and enrollment to this trial

was closed before completion of either the first stage of the two-stage trial or the CNS feasibility component. The design for cohort A was modified to a single-stage design with 11 patients, with 87% power to compare a null DCR of 5% with an alternative of 39.5%, using an exact binomial test and a one-sided type-I error of 7.5%.

Baseline patient characteristics of the 19 treated patients are shown in Table 1. Patients were predominantly female (74%), with a median age of 67 years (range, 38–85 years). Twelve (63%) patients had mucosal melanoma, 4 (21%) had acral melanoma, and 3 (16%) had melanoma arising from chronically sun-damaged skin (CSD). All patients had locoregionally advanced (5%) or distant disease (95%); most patients received one or more prior therapies. Sixteen patients received prior imatinib, one received both sorafenib and imatinib (patient 2), and 3 received prior ipilimumab therapy. All patients previously treated with a KIT inhibitor experienced progression on those agents and were not enrolled onto this study due to intolerance of prior therapy. Six of the 8 patients treated on cohort B received prior therapy with imatinib, and 2 patients were naïve to KIT inhibition. Patient demographic and disease characteristics were similar between cohorts A and B.

Tumor from the 19 treated patients was tested for the presence of KIT mutations, with 17 harboring such alterations (Tables 2 and 3). The specific mutations identified included exon 11 L576P (n = 4), exon 11 V560D (n = 1), exon 11 V560E (n = 1), exon 11 W557R (n = 1), exon 11 V559C (n = 1), exon 11 WKVVE 557-561 (n = 1), exon 13 K642E (n = 3), exon 13 Y646D (n = 1), exon 17 D820Y (n = 1), exon 17 N822K (n = 1), and exon 18 L831P (n = 1). One patient had tumor harboring two exon 13 mutations

| | Overall population (n = 19) | Cohort A (<i>n</i> = 11) | Cohort B (<i>n</i> = 8) |
|--|-----------------------------|---------------------------|--------------------------|
| Age in years, median (range) | 67.0 (38.0-85.0) | 68.0 (55.0-82.0) | 60.0 (38.0- 85.0 |
| Gender | | | |
| Male (%) | 5 (26.3%) | 2 (18.2%) | 3 (37.5%) |
| Female (%) | 14 (73.7%) | 9 (81.8%) | 5 (62.5%) |
| Race | | | |
| Caucasian (%) | 16 (84.2%) | 10 (90.9%) | 6 (75.0%) |
| Black/African American (%) | 1 (5.3%) | 1 (9.1%) | 0 (0%) |
| Other (%) | 2 (10.5%) | 0 (0%) | 2 (25.0%) |
| Ethnicity | | | |
| Hispanic or Latino (%) | 3 (15.8%) | 1 (9.1%) | 2 (25.0%) |
| Non-Hispanic (%) | 10 (52.6%) | 7 (63.6%) | 3 (37.5%) |
| Not reported (%) | 6 (31.6%) | 3 (27.3%) | 3 (37.5%) |
| Clinical melanoma subtype | | | |
| Acral | 4 (21.0%) | 2 (18.2%) | 2 (25.0%) |
| Mucosal | 12 (63.2%) | 7 (63.6%) | 5 (62.5%) |
| CSD | 3 (15.8%) | 2 (18.2%) | 1 (12.5%) |
| ECOG ^a performance status | | | |
| 0 (%) | 11 (57.9%) | 7 (63.6%) | 4 (50.0%) |
| 1 (%) | 8 (42.1%) | 4 (36.4%) | 4 (50.0%) |
| 2 (%) | 0 (0%) | 0 (0%) | 0 (0%) |
| Stage | | | |
| III (%) | 1 (5.3%) | 1 (9.1%) | 0 (0%) |
| IV (%) | 18 (94.7%) | 10 (90.9%) | 8 (100%) |
| Elevated lactate dehydrogenase ^b (%) | 8 (42.1%) | 5 (45.5) | 3 (37.5%) |
| Number of prior systemic therapies, median (range) | 2 (0-5) | 2 (1-4) | 2 (0-5) |
| Imatinib (%) | 17 (89.5%) | 11 (100%) | 6 (75.0%) |
| Sorafenib (%) | 1 (5.3%) ^c | 1 (9.1%) | 0 (0%) |
| Other kinase inhibitor (%) | 0 (0%) | 0 (0%) | 0 (0%) |
| Ipilimumab (%) | 3 (15.8%) | 1 (9.1%) | 2 (25.0%) |

^bValue exceeding 280 U/L

^cPatient 2 received both sorafenib and imatinib.

www.aacrjournals.org

Clin Cancer Res; 21(10) May 15, 2015 2291

Table 2. Clinical melanoma subtype, associated KIT alterations, clinical response to prior therapy with a KIT inhibitor, and clinical response to nilotinib in those without CNS involvement (cohort A)

| Study subject # | Melanoma subtype | KIT mutation | KIT amplification | Prior KIT inhibitor ^a | RECIST response to prior KIT inhibitor | PFS to prior KIT inhibitor (mo) | RECIST response to nilotinib (best percent response) | PFS to nilotinib (mo) |
|--------------------|---------------------|-------------------------|----------------------|--|--|---------------------------------------|--|-----------------------------|
| 1 | Mucosal | Exon 17 D820Y | Present (qPCR) | Imatinib | PR | 3.8 | SD (-26%) | 3.6 |
| 2 ^b | Acral | Exon 13 K642E | Present (FISH) | Imatinib | uPR | 4.1 | PD (6%) ^c | 0.8 |
| 3 | Mucosal | Exon 11 L576P | Not Present (FISH) | Imatinib | PR | 12.4 | PR (-59%) | 37.5+ |
| 5 | Mucosal | None | Present (qPCR) | Imatinib | SD | 11.5 | Clinical PD | 0.9 |
| 8 | CSD | Exon 17 N822K | Present (FISH) | Imatinib | SD | 8.3 | SD (0%) | 3.3 |
| 9 | Mucosal | Exon 13 R643Q and K642E | Not tested | Imatinib | Unk | Unk | SD (14%) | 3.9 |
| 11 | CSD | Exon 11 V559C | Present (FISH) | Imatinib | SD | 16 | Ineval | 0.1 |
| 12 | Mucosal | Exon 11 L576P | Present (qPCR) | Imatinib | PR | 7 | Clinical PD | 1.8 |
| 13 | Acral | Exon 11 WKVVE 557-561 | Not tested | Imatinib | SD | 8 | PD ^c (18%) | 2.1 |
| 14 | Mucosal | Exon 13 K642E | Not tested | Imatinib | SD | 3 | SD (-22%) | 5.5 |
| 20 | Mucosal | Exon 13 K642E | Present (qPCR) | Imatinib | CR | 20 | PR (-54%) | 11.5 |

^aAll patients previously treated with a KIT inhibitor experienced progression on those agents and were not enrolled onto this study due to intolerance of prior therapy. ^bPatient 2 also received sorafenib; however, his response to this therapy is not known.

^cSignifies the development of progression in nontarget lesions or the development of new lesions.

(R634Q and K642E). KIT amplification was tested in 12 cases, with 8 found to harbor such alteration. Two cases harbored amplification without a concurrent KIT mutation.

At the time of data analysis, 18 of the 19 treated patients were off-study, 14 of whom due to progressive disease. Median followup was 16.2 months in cohort A (90% CI, 6.9–37.5 months) and 11.7 months in cohort B (90% CI, 2.1 months– ∞). (n = 2). No patient experienced grade-4 related adverse events. Toxicity rates and patterns were comparable for cohorts A and B.

Clinical activity

Toxicity

Adverse events classified as possibly, probably, or definitely related to nilotinib are shown in Supplementary Table S1. Events that were recorded multiple times for any patient are reported only once according to the worst grade. Although nilotinib was generally well tolerated, 17 of the 19 patients treated reported adverse events, with fatigue (26%) and low-grade musculoskeletal and gastrointestinal discomfort (32%) most commonly observed. Grade-3 toxicities were observed in 4 patients, and included rash (n = 1), elevated pancreatic enzymes (n = 2), and transaminitis and hyponatremia (n = 1). Grade 3 toxicity was managed by dose reduction to 400 mg QD (n = 2) or dose delay followed by reinitiation of treatment at 400 mg twice a day

Four-month disease control rate. In cohort A, 3 of 11 patients were alive without disease progression at 4 months (27%; 90% CI, 8%–56%), a proportion significantly greater than the DCR of 5% (P = 0.03) assumed under the null hypothesis. On the basis of three observed responses, there is sufficient evidence to conclude that nilotinib would have been considered worthy of further study in cohort A based on the initial two-stage design. In cohort B, 1 of 8 treated patients achieved disease control at 4 months (12.5%; 90% CI, 0.6%–47%), with no evidence that 4-month DFR is greater than 5% in this population.

Response rate. Of the 19 patients treated, 4 were inevaluable for radiographic response to therapy in non-CNS lesions. In cohort A, patient 11 initiated therapy but subsequently underwent resection of abdominal disease due to tumor-associated gastrointestinal bleeding. In cohort B, patient 16 initiated therapy but developed rapid clinical decline due to progressive leptomeningeal disease and withdrew consent for further treatment and

Table 3. Clinical melanoma subtype, associated KIT alterations, clinical response to prior therapy with a KIT inhibitor, and clinical response to nilotinib in those with CNS involvement

| Study subject # | Melanoma subtype | KIT mutation | KIT amplification | Prior KIT inhibitor ^a | RECIST response to prior KIT inhibitor | PFS to prior KIT inhibitor (mo) | RECIST response to nilotinib in non-CNS lesions (best percent response) | RECIST response to nilotinib in CNS lesions (best percent response) | PFS to Nilotinib (Months) |
|--------------------|---------------------|-----------------|----------------------|--|---|--|---|---|---------------------------------|
| 4 | Mucosal | Exon 11 V560D | Not present (FISH) | None | n/a | n/a | SD (-20%) | PR (-36%) ^b | 3.9 |
| 6 | Acral | Exon 11 W577R | Not present (FISH) | Imatinib | Unk | Unk | SD (-23%) | SD ^c (11%) | 6.6 |
| 7 | Acral | None | Present (qPCR) | Imatinib | PD | 1.7 | SD (3%) | PD (37.5%) | 2.1 |
| 10 | Cutaneous | Exon 11 V560D | Not tested | Imatinib | PR | 4.6 | PD (44%) | PD (40%) | 2.4 |
| 15 | Mucosal | Exon 11 L576P | Not tested | None | n/a | n/a | n/a ^b | SD (-25%) | 2.9 |
| 16 | Mucosal | Exon 13 Y646D | Not tested | Imatinib | SD | ~ 4 | Uneval | Uneval | 0.2 |
| 17 | Mucosal | Exon 18 L831P | Not present (FISH) | Imatinib | SD | \sim 7 | n/a ^d | SD (9%) | 1.6 |
| 18 | Mucosal | Exon 11 L576P | Not tested | Imatinib | SD | 2.8 | SD (14%) | PD (0%) ^b | 1.8 |

Abbreviation: Unk, unknown

^aAll patients previously treated with a KIT inhibitor experienced progression on those agents and were not enrolled onto this study due to intolerance of prior therapy. ^bSignifies the development of progression in nontarget lesions or the development of new lesions.

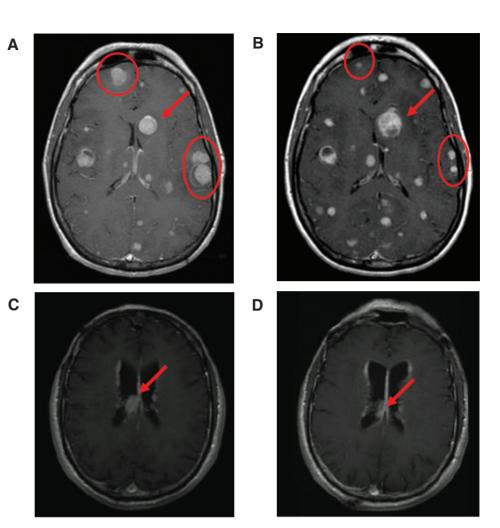
^cPatient 6 underwent resection of one symptomatic brain target lesion 1.6 months after initiation of therapy; a second CNS target lesion remained stable for 6.6 months after initiation of therapy; however, new CNS lesions were noted at that time and the patient was taken off for POD.

^dNo non-CNS lesions present.

Nilotinib in Previously Treated KIT-Altered Melanoma

Figure 1.

Representative images from two patients achieving radiographic responses in brain metastases with nilotinib. Magnetic resonance images of brain metastases present at baseline (A) and after 4 months of therapy (B) in a patient who achieved a minor response in extracranial metastases and a PR in target brain metastases as demonstrated by the circled lesions are presented. The baseline brain MRI was performed 5 months after receiving stereotactic radiosurgery to left temporal, left parietal, right frontal, and right midcerebellar lesions and demonstrates the development of progression in the previously treated lesions and the development of numerous new infra and supratentorial hemorrhagic brain metastases (A). Despite durable stability in the extracranial disease after 4 months of therapy and further reduction in the size of several of the brain metastases, there was progressing in nontarget brain metastases (arrow). Magnetic resonance images of a brain metastasis present at baseline (C) and after 2 months of therapy (D) in a patient who achieved a PR in a solitary brain metastases are presented. No prior radiotherapy or surgery was performed in this patient before initiation of study therapy.



evaluation. Patients 15 and 17 presented with CNS-only disease, without measurable lesions in extracranial sites.

In cohort A, two PRs were observed (18.2%, 90% CI, 3%– 47%). One PR was observed in an 81-year-old female with advanced vulvar melanoma harboring an exon 11 L576P mutation without concurrent amplification (Patient 3). She previously achieved a durable PR to therapy with imatinib lasting 12.4 months and has an ongoing response to nilotinib at 37.5 months. Additional patients achieved minor responses to therapy (Tables 2 and 3).

Of the 8 patients treated on cohort B, 7 were evaluable for response in CNS metastases which either were not previously treated with radiotherapy or which demonstrated progression following treatment (Fig. 1). Assessing CNS lesions only, we observed one PR (12.5%, 90% CI, 0.6%–47%) lasting 3.9 months (Patient 4) and one minor response (Patient 15), each in patients not previously treated with a KIT inhibitor. The PR was observed in a 48-year-old female with mucosal melanoma arising from the anorectal region harboring an exon 11 V560D mutation without concurrent amplification. A brain MRI performed 5 months after receiving stereotactic radiosurgery to left temporal, left parietal, right frontal, and right mid-cerebellar lesions demonstrated the development of numerous infra and supratentorial hemorrahgic brain metastases (Fig. 1A). She achieved a minor response

in her extracranial metastases (20% tumor regression by RECIST criteria) and a PR in her target brain metastases (36% regression by RECIST criteria) as demonstrated by the circled lesions in Fig. 1A and 1B. Despite durable stability in her extracranial disease after 4 months of therapy and further reduction in the size of several of the brain metastases, there was progression in nontarget brain metastases and she was taken off study.

Time-to-progression. The TTP achieved with nilotinib as well as to a prior KIT inhibitor, if applicable, is shown by patient in Fig. 2. The median TTP was 3.4 months (90% CI, 0.9–5.5 months) and 2.6 months (90% CI, 1.8–3.9 months; Fig. 3A) in cohorts A and B, respectively.

Overall survival. Eleven patients (57.9%) were deceased at the time of data analysis, with one patient lost to follow-up. The median OS in cohort A was 14.2 months (90% CI, 7.1 months– ∞) and was longer than observed in cohort B (4.3 months; 90% CI, 3.5–11.9 months; P = 0.05; Fig. 3B).

Discussion

These results demonstrate that a subset of patients with melanomas harboring genetic alterations of KIT may benefit from nilotinib after experiencing disease progression to a prior KIT inhibitor. Three of 11 patients without brain metastasis achieved disease control at 4 months with nilotinib, with observed

| Study subject # | KIT mutation | KIT amplification | 1 | | | | 1 | | | | | 1 | | | | Imat | inib | |
|-----------------|--|----------------------|---|---|---|----|----|---|----|------------|------------|---|---------|----|----|------|------|----|
| 1 | Exon 17 D820Y | Present | | | | | | | | | | | | | | Nilo | inib | |
| 2 | Exon 13 K642E | Present | | | | | | | | | | | | l | | | | |
| 3 | Exon 11 L576P | Not present | | | | | | | | | | | | | | | | |
| 4 | Exon 11 V560E | Not present | | | | | | | | | | | | | | | | |
| 5 | None | Present | | | | | | | | | | | | | | | | |
| 6 | Exon 11 W577R | Not present | * | | | | | | | | | | | | | | | |
| 7 | None | Present | | | | | | | | | | | | | | | | |
| 8 | Exon 17 N822K | Present | | | | | | | | | | | | | | | | |
| 9 | Exon 13 R643Q and K642E | Not tested | * | | | | | | | | | | | | | | | |
| 10 | Exon11 V560D | Not tested | | | | | | | | | | | | | | | | |
| 11 | Exon 11 V559C | Present | | | | | | | | | | | | | | | | |
| 12 | Exon 11 L576P | Present | | | | | | | | | | | | | | | | |
| 13 | Exon 11 WKVVE 557-561 | Not tested | | | | | | | | | | | | | | | | |
| 14 | Exon 13 K642E | Not tested | | | | | | | | | | | | | | | | |
| 15 | Exon 11 L576P | Not tested | | | | | | | | | | | | | | | | |
| 16 | Exon 13 Y646D | Not tested | | | | | | | | | | | | | | | | |
| 17 | Exon 18 L831P | Not present | | | | | | | | | | | | | | | | |
| 18 | Exon 11 L576P | Not tested | | | | | | | | | | | | | | | | |
| 20 | Exon13 K642E | Present | | | | | | | | | | | | | | | | |
| | atients 6 and 9 received prior of imatinib therapy is not kno | | 0 | 4 | 8 | 12 | 16 | 2 | 20 | 24 Time | 28 e (m | | 36) | 40 | 44 | 48 | 52 | 56 |

Figure 2.

Treatment response over time to imatinib and nilotinib by genetic alteration of KIT.

progression-free survival times of 5.5, 11.5, and 37.5+ months. Notably, patients 3 and 20 achieved a durable PR and CR, respectively, to imatinib lasting 12.4 and 20 months, respectively, before achieving durable PRs to nilotinib, demonstrating that nilotinib can overcome the development of secondary resistance to imatinib. On the basis of the original study design for cohort A which required 3 or more patients to achieve disease control at 4 months, the primary endpoint of 4-month DCR was achieved.

Given the high incidence of brain metastases in melanoma and the potential efficacy of second-generation KIT inhibitors in CNS metastases (20), we included an exploratory cohort of patients with brain metastases from melanoma harboring KIT alterations. Although available data suggest the limited penetration of nilotinib within the CNS, clinical activity has been observed in the brain in BCR-ABL–positive leukemia (21). Such efficacy may be explained by the high protein-binding affinity of nilotinib coupled with the low protein concentration within the cerebrospinal fluid, thus resulting in relatively higher amounts of free nilotinib within the CNS. Indeed, of 7 patients in our trial evaluable for response in brain lesions, one achieved a 36% reduction and another achieved a 25% reduction in the CNS tumor burden with therapy. A mixed response in the brain lesions was observed in some cases, with clear reduction in the size of several brain metastases and unambiguous progression in others. Although anecdotal, these variable responses may suggest more prominent intra-tumoral molecular heterogeneity in CNS lesions when compared with disease in other organs or variable pharmacologic penetration into the brain metastases. Of note, both patients who achieved radiographic responses within the brain were not previously treated with a KIT inhibitor such as imatinib. Despite the radiographic changes observed, the progression-free and OS in this cohort of patients were short.

The greater potency of nilotinib over imatinib against the mutant KIT oncoprotein provides pharmacologic rationale for using nilotinib (18, 22). Furthermore, the sensitivity of specific *KIT* mutations to clinically available inhibitors can differ, with some mutations affecting the binding affinity of specific inhibitors of KIT as previously demonstrated in *in vitro* and clinical studies of GIST (13, 19, 23). Although preliminary evidence of activity with nilotinib in patients with melanoma harboring KIT alterations not previously treated with a KIT inhibitor has been observed, with two PRs lasting 8.4 and 10+ months reported in nine patients with melanoma harboring a KIT alteration not previously treated with a KIT inhibitor (24), whether nilotinib is superior to imatinib in KIT-inhibitor naïve patients with melanoma is unknown. In advanced GIST, nilotinib was not superior

Clinical Cancer Research

Nilotinib in Previously Treated KIT-Altered Melanoma

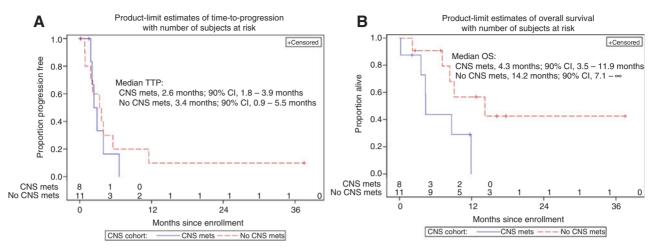


Figure 3.

TTP and OS. Kaplan-Meier estimates of TTP (A) and OS (B) in those enrolled on cohorts A (dotted red lines) and B (solid blue line) are shown. The vertical lines indicate that patients' data were censored. The median TTP was 3.4 months (90% CI, 0.9-5.5) in cohort A and 2.6 months (90% CI, 1.8-3.9) in cohort B. OS was 14.2 months (90% CI, $7.1 -\infty$) in cohort A and 4.3 months (90% CI, 3.5-11.9) in cohort B.

to imatinib as first-line therapy and did not improve outcomes when compared with best-supportive care in the third-line setting (25, 26). Importantly, mechanisms of secondary resistance in GIST, which commonly involve the development of secondary *KIT* mutations affecting the tyrosine kinase domains in exons 13 and 17 (27,28), appear to differ from those observed in melanoma driven by KIT alterations. Thus far, no such secondary mutations have been observed in KIT melanoma. Rather, the limited data available suggest that, in melanoma, the development of secondary NRAS mutations (11) and activation of the mTOR pathway by alternative mechanisms may result in secondary resistance (29).

In conclusion, the use of nilotinib in a subset of patients with melanoma harboring KIT alterations previously treated with an inhibitor of KIT can result in clinical benefit, although efficacy of this agent in brain metastasis is limited. Although this trial is underpowered to conclude clinical benefit, the data suggest further studies of sequential KIT inhibitor therapy for this molecular subset of patients is warranted.

Disclosure of Potential Conflicts of Interest

R.D. Carvajal is a consultant/advisory board member for AstraZeneca, Aura Biosciences, and Novartis. J. Weber is a consultant/advisory board member for Bristol-Myers Squibb, GlaxoSmithKline, Merck, and Novartis. P.B. Chapman is a consultant/advisory board member for Bristol-Myers Squibb, GlaxoSmithKline, and Roche/Genentech. M.C. Heinrich is a consultant/advisory board member for and reports receiving commercial research grants and speakers bureau honoraria from Novartis, has ownership interest (including patents) in MolecularMD; and has provided expert testimony in patent litigation proceedings related to a Korean patent concerning treating GI stromal tumors with imatinib for Novartis. B.C. Bastian is a consultant/advisory board member for Novartis. C.L. Corless reports receiving speakers bureau honoraria from and is a consultant/advisory board member for Novartis. No potential conflicts of interest were disclosed by the other authors.

Authors' Contributions

Conception and design: S.J. O'Day, J.D. Wolchok, A. Giobbie-Hurder, M.C. Heinrich, F.S. Hodi

Development of methodology: D.P. Lawrence, S.J. O'Day, A. Giobbie-Hurder, M.C. Heinrich, J.A. Fletcher, F.S. Hodi

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): R.D. Carvajal, D.P. Lawrence, J. Weber, T.F. Gajewski, R. Gonzalez, J. Lutzky, S.J. O'Day, O. Hamid, J.D. Wolchok, P.B. Chapman, R.J. Sullivan, J.B. Teitcher, C.R. Antonescu, M.C. Heinrich, C.L. Corless, J.A. Fletcher, F.S. Hodi

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): R.D. Carvajal, D.P. Lawrence, J. Weber, T.F. Gajewski, S.J. O'Day, O. Hamid, N.H. Ramaiya, A. Giobbie-Hurder, C.R. Antonescu, M.C. Heinrich, C.L. Corless, J.A. Fletcher, F.S. Hodi

Writing, review, and/or revision of the manuscript: R.D. Carvajal, D.P. Lawrence, J. Weber, T.F. Gajewski, R. Gonzalez, J. Lutzky, S.J. O'Day, O. Hamid, J.D. Wolchok, P.B. Chapman, R.J. Sullivan, J.B. Teitcher, N.H. Ramaiya, A. Giobbie-Hurder, C.R. Antonescu, M.C. Heinrich, B.C. Bastian, J.A. Fletcher, F.S. Hodi

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): R.D. Carvajal, D.P. Lawrence, C.L. Corless, F.S. Hodi

Study supervision: R.D. Carvajal, D.P. Lawrence, R. Gonzalez, O. Hamid, J.D. Wolchok, F.S. Hodi

Acknowledgments

The authors acknowledge the assistance of Carla Becerra, Maria Gargano, and Janice Russell from Dana-Farber Cancer Center and Shahnaz Singh-Kandah from Memorial Sloan Kettering Cancer Center for their contributions to this study, which included data acquisition, data analysis, and administrative support.

Grant Support

Funding for this trial was provided by Novartis. R.D. Carvajal and B.C. Bastian received funding from NIH/NCI grant P01 CA025874.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received June 24, 2014; revised January 9, 2015; accepted January 31, 2015; published OnlineFirst February 18, 2015.

www.aacrjournals.org

References

- 1. Curtin JA, Busam K, Pinkel D, Bastian BC. Somatic activation of KIT in distinct subtypes of melanoma. J Clin Oncol 2006;24:4340–6.
- Hodi FS, Friedlander P, Corless CL, Heinrich MC, Mac Rae S, Kruse A, et al. Major response to imatinib mesylate in KIT-mutated melanoma. J Clin Oncol 2008;26:2046–51.
- Satzger I, Kuttler U, Volker B, Schenck F, Kapp A, Gutzmer R. Anal mucosal melanoma with KIT-activating mutation and response to imatinib therapy-case report and review of the literature. Dermatology 2010;220:77–81.
- Lutzky J, Bauer J, Bastian BC. Dose-dependent, complete response to imatinib of a metastatic mucosal melanoma with a K642E KIT mutation. Pigment Cell Melanoma Res 2008;21:492–3.
- Woodman SE, Trent JC, Stemke-Hale K, Lazar AJ, Pricl S, Pavan GM, et al. Activity of dasatinib against L576P KIT mutant melanoma: molecular, cellular, and clinical correlates. Mol Cancer Ther 2009; 8:2079–85.
- Quintas-Cardama A, Lazar AJ, Woodman SE, Kim K, Ross M, Hwu P. Complete response of stage IV anal mucosal melanoma expressing KIT Val560Asp to the multikinase inhibitor sorafenib. Nat Clin Pract Oncol 2008;5:737–40.
- 7. Zhu Y, Si L, Kong Y, Chi Z, Yuan X, Cui C, et al. Response to sunitinib in Chinese KIT-mutated metastatic mucosal melanoma. J Clin Oncol 2009;27:15s (suppl; abstr e20017).
- Carvajal RD, Antonescu CR, Wolchok JD, Chapman PB, Roman RA, Teitcher J, et al. KIT as a therapeutic target in metastatic melanoma. JAMA 2011;305:2327–34.
- Guo J, Si L, Kong Y, Flaherty KT, Xu X, Zhu Y, et al. Phase II, open-label, single-arm trial of imatinib mesylate in patients with metastatic melanoma harboring c-Kit mutation or amplification. J Clin Oncol 2011;29:2904–9.
- Hodi FS, Corless CL, Giobbie-Hurder A, Fletcher JA, Zhu M, Marino-Enriquez A, et al. Imatinib for melanomas harboring mutationally activated or amplified KIT arising on mucosal, acral, and chronically sun-damaged skin. J Clin Oncol 2013;31:3182–90.
- Minor DR, Kashani-Sabet M, Garrido M, O'Day SJ, Hamid O, Bastian BC. Sunitinib therapy for melanoma patients with KIT mutations. Clin Cancer Res 2012;18:1457–63.
- Handolias D, Hamilton AL, Salemi R, Tan A, Moodie K, Kerr L, et al. Clinical responses observed with imatinib or sorafenib in melanoma patients expressing mutations in KIT. Br J Cancer 2010; 102:1219–23.
- Heinrich MC, Corless CL, Blanke CD, Demetri GD, Joensuu H, Roberts PJ, et al. Molecular correlates of imatinib resistance in gastrointestinal stromal tumors. J Clin Oncol 2006;24:4764–74.
- Demetri GD, van Oosterom AT, Garrett CR, Blackstein ME, Shah MH, Verweij J, et al. Efficacy and safety of sunitinib in patients with advanced gastrointestinal stromal tumour after failure of imatinib: a randomised controlled trial. Lancet 2006;368:1329–38.
- Park SH, Ryu MH, Ryoo BY, Im SA, Kwon HC, Lee SS, et al. Sorafenib in patients with metastatic gastrointestinal stromal tumors who failed two or more prior tyrosine kinase inhibitors: a phase II study of Korean gastrointestinal stromal tumors study group. Invest New Drugs 2012; 30:2377–83.

- Demetri GD, Reichardt P, Kang YK, Blay JY, Rutkowski P, Gelderblom H, et al. Efficacy and safety of regorafenib for advanced gastrointestinal stromal tumours after failure of imatinib and sunitinib (GRID): an international, multicentre, randomised, placebo-controlled, phase 3 trial. Lancet 2013;381:295–302.
- 17. Weisberg E, Manley PW, Breitenstein W, Bruggen J, Cowan-Jacob SW, Ray A, et al. Characterization of AMN107, a selective inhibitor of native and mutant Bcr-Abl. Cancer Cell 2005;7:129–41.
- Cullinane C, Natoli A, Hui Y, Conus N, Jackson S, Bruggen J, et al. Preclinical evaluation of nilotinib efficacy in an imatinib-resistant KIT-driven tumor model. Mol Cancer Ther 2010;9:1461–8.
- Guo T, Hajdu M, Agaram NP, Shinoda H, Veach D, Clarkson BD, et al. Mechanisms of sunitinib resistance in gastrointestinal stromal tumors harboring KITAY502–3ins mutation: an *in vitro* mutagenesis screen for drug resistance. Clin Cancer Res 2009;15:6862–70.
- Porkka K, Koskenvesa P, Lundan T, Rimpilainen J, Mustjoki S, Smykla R, et al. Dasatinib crosses the blood-brain barrier and is an efficient therapy for central nervous system Philadelphia chromosome-positive leukemia. Blood 2008;112:1005–12.
- Reinwald M, Schleyer E, Kiewe P, Blau IW, Burmeister T, Pursche S, et al. Efficacy and pharmacologic data of second-generation tyrosine kinase inhibitor nilotinib in BCR-ABL-positive leukemia patients with central nervous system relapse after allogeneic stem cell transplantation. BioMed Res Int 2014;2014:637059.
- 22. Sawaki A, Nishida T, Doi T, Yamada Y, Komatsu Y, Kanda T, et al. Phase 2 study of nilotinib as third-line therapy for patients with gastrointestinal stromal tumor. Cancer 2011;117:4633–41.
- Heinrich MC, Corless CL, Demetri GD, Blanke CD, von Mehren M, Joensuu H, et al. Kinase mutations and imatinib response in patients with metastatic gastrointestinal stromal tumor. J Clin Oncol 2003;21: 4342–9.
- Cho JH, Kim KM, Kwon M, Kim JH, Lee J. Nilotinib in patients with metastatic melanoma harboring KIT gene aberration. Invest New Drugs 2012;30:2008–14.
- Blay J, Shen L, Kang Y, Rutkowski P, Qin S, Nosov D, et al. Phase III trial of nilotinib versus imatinib as first-line targeted therapy of advanced gastrointestinal stromal tumors (GIST). J Clin Oncol 31, 2013 (suppl; abstr 10501).
- Reichardt P, Blay JY, Gelderblom H, Schlemmer M, Demetri GD, Bui-Nguyen B, et al. Phase III study of nilotinib versus best supportive care with or without a TKI in patients with gastrointestinal stromal tumors resistant to or intolerant of imatinib and sunitinib. Ann Oncol 2012; 23:1680–7.
- Lasota J, Corless CL, Heinrich MC, Debiec-Rychter M, Sciot R, Wardelmann E, et al. Clinicopathologic profile of gastrointestinal stromal tumors (GISTs) with primary KIT exon 13 or exon 17 mutations: a multicenter study on 54 cases. Mod Pathol 2008;21:476–84.
- Fletcher JA, Rubin BP. KIT mutations in GIST. Curr Opin Genet Dev 2007;17:3–7.
- Si L, Xu X, Kong Y, Flaherty KT, Chi Z, Cui C, et al. Major response to everolimus in melanoma with acquired imatinib resistance. J Clin Oncol 2012;30:e37–40.



Clinical Cancer Research

Phase II Study of Nilotinib in Melanoma Harboring KIT Alterations Following Progression to Prior KIT Inhibition

Richard D. Carvajal, Donald P. Lawrence, Jeffrey S. Weber, et al.

Г

Clin Cancer Res 2015;21:2289-2296. Published OnlineFirst February 18, 2015.



| Cited articles | This article cites 27 articles, 13 of which you can access for free at: http://clincancerres.aacrjournals.org/content/21/10/2289.full.html#ref-list-1 |
|-----------------|---|
| Citing articles | This article has been cited by 1 HighWire-hosted articles. Access the articles at: http://clincancerres.aacrjournals.org/content/21/10/2289.full.html#related-urls |

| E-mail alerts | Sign up to receive free email-alerts related to this article or journal. | |
|-------------------------------|--|--|
| Reprints and Subscriptions | To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org. | |
| Permissions | To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org. | |