A. Solutions and Reagents

NOTE: Prepare solutions with Milli-Q or equivalently purified water.

1. **Wash Buffer:** 1X Tris buffered saline, 0.1% Tween-20 (TBS/T)
2. **Stripping Buffer:** To prepare 100 ml, mix 0.76 g Tris base, 2 g SDS and 700 µl β-mercaptoethanol. Bring to 100 ml with deionized H_2O. Adjust pH to 6.8 with HCl.

B. Protocol

1. After film exposure, wash membrane four times for 5 minutes each in TBS/T. Best results are obtained if the membrane is not allowed to dry.
2. Incubate membrane for 30 minutes at 50°C in stripping buffer (with slight agitation).
3. Wash membrane six times for 5 minutes each in TBS/T.
4. (Optional) To assure that the original signal is removed, wash membrane twice for 5 minutes each with 10 ml of TBS/T. Incubate membrane with LumiGLO® with gentle agitation for 1 minute at room temperature. Drain membrane of excess developing solution. Do not let dry. Wrap in plastic wrap and expose to x-ray film.
5. Wash membrane again four times for 5 minutes each in TBS/T.
6. The membrane is now ready to reuse. Start detection at the “Membrane Blocking and Antibody Incubations” step in the Western Immunoblotting Protocol.

- See more at: http://www.cellsignal.com/contents/resources-protocols/western-blot-reprobing-protocol/western-reprobing?Ntt=western+blot+stripping&fromPage=search#sthash.biof4hHX.dpuf