## Synthesis, Characterization and Antihyperlipidemic of Rutin-calcium (II) Complex

Zhang Jing-liang<sup>a</sup>, Cui Chang<sup>b</sup>, Zhang Hui-li<sup>c</sup>, Wang Shi-guang<sup>a</sup>, Liu Jin-hai<sup>c</sup>, Zhu Wei<sup>d</sup>, Zhai Guang-yu<sup>c, e</sup>

<sup>a</sup> School of Medicine, Zhengzhou university of industrial technology, Zhengzhou,451100, PR China
<sup>b</sup> School of Medicine, huanghe science and technology college, Zhengzhou,450000, PR China
<sup>c</sup> School of Pharmacy, Zhengzhou university of industrial technology, Zhengzhou,451100, PR China
<sup>d</sup> Department of Biomedical Engineering & Instrument Science, Zhejiang University, Hangzhou 310027, PR China
<sup>e</sup> Department of Nursing, Zhengzhou University, Zhengzhou 450052, PR China
Corresponding author, Tel.: +86 371 66961670; E-mail address: zhaiguangyu@zzu.edu.cn (ZHAI Guang-yu)

**Abstract:** A rutin-ca complex was synthesised in methanol and was characterized using elemental analysis, UV-Vis spectroscopy, IR spectroscopy and differential thermal analysis. Spectroscopic data suggested that the metal/ligand ratio of the complex is 2:1. The antihyperlipidemic activity of the complex was evaluated by animal experiment of 30 wistar rats. It was shown that the complex was much more effective antihyperlipidemics than the free flavonoid.

[Zhang Jing-liang, Cui Chang, Zhang Hui-li, Wang Shi-guang, Liu Jin-hai, Zhu Wei, Zhai Guang-yu. **Synthesis, Characterization and Antihyperlipidemic of Rutin-calcium (II) Complex.** *Biomedicine and Nursing* 2016;2(2): 40-48]. ISSN 2379-8211 (print); ISSN 2379-8203 (online). http://www.nbmedicine.org. 8. doi:10.7537/marsbnj02021608

Keywords: flavonoid, rutin-calcium (II) complex, synthesis, antihyperlipidemic

#### 1. Introduction

Flavonoids is a group of natural polyphenolic compounds that widely distributed in plant foods, which shows a wide range of biological activities and pharmacological properties, such as cardiovascular protection, anti-cancer, anti-ulcer, anti-virus and anti-inflammatory [1]; it is an kind of important natural antioxidant, free radical scavenger as well as metal chelator [2]. Rutin (quercetin-3-O-rutinoside) is the most common flavonol widely distributed in higher plants, such as vegetables, fruits, especially citrus fruits (oranges, grapefruit, lemon), buckwheat and beverages. The most important properties of rutin is

scavenging oxygen free radicals, such as hydroxyl radicals, oxygen radicals and peroxyl radicals [3]. The ability of scavenging free radicals of rutin have been demonstrated by vitro experiments [4]. what's more, rutin has been used in the treatment of diseases [5]. The pharmacological activity of rutin including: antiallergic[6], anti-inflammatory and dilation of blood vessels[7], anti-cancer [8], anti-bacterial [9], anti-virus [10]. Clinically for the treatment of allergic purpura and a variety of increased capillary fragility caused by bleeding disorders [11], also used to treat high blood pressure and senile bronchitis[12].

Fig. 1 The chemical structure of rutin

Rutin, which has the appropriate spatial configuration and strong ligand oxygen atoms, is an excellent metal chelator [13], can form the stable complexes with many metal ions. Studies show that

the biological activities of rutin complexes is stronger than that of the ligand [14]. The study on rutin metal complexes has aroused widespread concern [15]. So far, many a rutin complex has been reported: Al (III), Zn (II) [16], Cu (II) [17], Co (II) [18], Ni [19], Fe (II) [20], Fe (III) [21], Mn (II) [22], V (II) [23], U (II) [24], Au (III) [25]. In daily life, regular supplementing of rutin-rich foods, such as ginkgo biloba, buckwheat, tomatoes, onions, tea,may be contributed to scavenging free radicals and reducing the content of heavy metal ions in our body [26].

Calcium is one of the most important elements in living organisms. It plays an important role in the metabolism of nitrogen in plants, and represents the structural component of animal bones [27]. In cell physiological level, calcium is used to regulate the permeability and electrical properties of biological membranes, which in turn function as a signal for many cellular processes [28]. Calcium ion also serves important functions in blood clotting, and by acting as a "second messenger" in signal transduction, in the triggering of muscle contractions, and in the transmissions of nerve impulses [29]. In addition, it can lower the levels of  $\alpha\text{-LP}$  and  $\beta\text{-LP}$ . The atherosclerosis and cardiac arrhythmia could be induced by the lack of calcium [30]. It is suggested that the biological activity of an organic ligand can be increased when coordinated or mixed with suitable metal ion: because of its ability to act as free radical acceptor [31].

Herein, we put forward the methodology of the complexation process between Ca (II) and rutin, characterization of the complex as well as the investigation of antihyperlipidemic properties of the rutin-calcium complex.

#### 2. Materials and methods

## 2.1 Reagents

Rutin was purchased from Sigma(Sigma ChemicalCo.,St.Louis.MO,USA). The other reagent and solvents were of analytical reagent grade. Cholesterol was purchased from shanghai chemical reagent company, sodium cholate from ASUS shanghai fine chemicals company, propylthiouracil from shanghai fuxing zhaohui pharmaceutical Co., Ltd., total cholesterol kit, triglyceride kit and high-density lipoprotein cholesterol kit from BIOSINO Bio-Technology Co., Ltd., and thirty healthy Wister rats weighing 200g±12g from the experimental animal center of Henan province.

#### 2.2 Instrumentation

UV Spectra was obtained by a UV-2550 ultraviolet-visible spectrophotometer using standard 1.00cm quartz cells (Shimadzu Corporation). Elemental analyses were performed using Perkin-400CHN elemental analyzer. The IR spectral was recorded by using KBr pellets in the spectral range 4000-400 on a FTS-3000 FTIR spectrophotometer. Melting points were determined on a X-5 melting point apparatus and are uncorrected. The differential

thermal analysis of the complex was performed by the Thermal analyzer STA 409 PC/PG (Netzsch,Germany), from room temperature to 784  $^{\circ}$ C at a heating rate of 10  $^{\circ}$ C/min.

#### 2.3 Synthesis of the complex

Rutin (0.61g, 1mmol) was added to a 100ml round-bottom flask containing 25ml MeOH, stir 15min until the solid rutin was completely dissolved, then the pH value was adjusted to 8.6 with CH<sub>3</sub>ONa. Anhydrous calcium chloride (222mg, 2mmol) dissolved in methanol (20ml) was added. The reaction mixture was refluxed for 4 h and monitored by TLC. The formed complex was collected by filtration and washed three times with a 1:3 methanol/water mixture and then several times with water and dried in a vacuum desiccator. A brown-yellow solid rutin-calcium complex was then obtained (m.p:301-305°C).

# 2.4 Antihyperlipidemia activity of the complex 2.4.1 Method

The preparation of fat emulsions: In a 500ml beaker, 80g of lard were heated to melt. 40g of cholesterol were added and mixed well. 4g of propylthiouracil and 8g of sodium cholate are mixed in a mortar. Then the powder, 80ml of tween 80, 80ml of 1, 2-propanediol was added into the mixture and was stirred until smooth. And double-distill water was added to 400ml. The emulsions were stored in the refrigerator. Before application, it needed to be heated. The concentration of the fat emulsions was: cholesterol 100g/L, lard 200g/L, sodium cholate 20g/L, propylthiouracil 10g/L.

30 Wistar rats were randomly divided into three groups: normal control group, model group and experimental group, and were fed with common feedstuff. The rats in the normal control group were given normal saline [10ml/(kg·d)] for 14 days. The rats of the model group were administered intragastrically with fat emulsion [10ml/(kg·d)] for 14 days while the rats of the experimental group were administered intragastrically with fat emulsion and rutin-calcium complex [300mg/(kg·d)].

After continuous intragastric infusion for 14 days, the serum TC, TG, HDL was measured respectively. All rats were sacrificed to weigh the liver, heart, spleen and kidney and to compute the organ coefficient.

#### 2.4.2 Statistical treatment

SPSS 10.0 software was used for homogeneity of variance and one-way ANOVA. Then LSD-t test was used for multiple comparison. The associations of the serum lipid levels and Viscera Index were analyzed by using Pearson correlation analysis.

### 3. Results and discussions

In this study, fifteen experiments that we designed were performed from pH 7.2 to pH 10 with

every 0.2~pH as an experimental unit. The results showed that the optimum pH is 8.6. In the same way, we found that the optimum reaction temperature was  $85^{\circ}\text{C}$  and the optimum reaction time was 4h.

#### 3.1 elemental analysis

Table 1. Elemental analysis of the rutin-ca complex

Element	The contents of elements (%)				
	Found values	Calculated values			
С	46.28	47.40			
Н	5.31	5.27			
Ca <sup>2+</sup>	2.93	3.02			

The chemical compositions were analyzed by the

combustion analysis spectrum for C, H and EDTA titration was applied to the determination of calcium ion. The results are listed in table 1.

We know that coordination molar ratio of rutin to calcium is 1:2.

#### 3.2 UV/Visible spectrum analysis

The UV/vis spectrum of the rutin-ca complex and rutin in MeOH are described in Fig.2 and 4. Rutin, like most flavones, exhibited two major absorption bands in the UV/vis region, namely 355nm (band I) representing B-ring absorption (cinnamoyl system), and 257nm (band II) representing the A- ring absorption (benzoyl system).

Fig.2. The structure of rutin and the division of band I and band II related to UV/vis absorption bands

The spectra were related to the  $\pi$ - $\pi$ \* transitions within the aromatic ring of the ligand molecules. In comparison with flavonoids absorption spectra, the

band of the complex was shifted to the long-wavelength region as shown in Fig. 3.

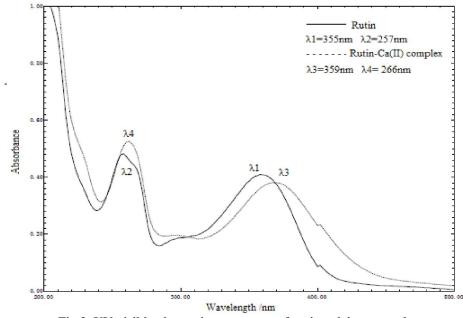


Fig.3. UV-visible absorption spectrum of rutin calcium complex

The isosbestic point which was characteristic of the formation of a complex was observed at 359nm.

Such bathochromic shift could be explained by the extension of the conjugated system with the

complexation. After chelating Ca (II) ion, each of the two bands had a shift (from 355 to 359nm for band I and from 257 to 266nm for band II)[32].

IR spectroscopies of the rutin-ca complex and rutin were analyzed and the main data are summarized in Table 2.

## 3.3 IR spectroscopy analysis

Table.2. IR spectra data (cm-1) of the rutin-ca complex and rutin

Compound	v (O-H)	v (C=O)	v (C=C)	v (C-OH)	v (C-O-C)
Rutin	3423.42	1655.03	1603.69	1361.52	1295.67
Ca-rutin	3377.50	1632.45	1551.29	1355.81	1288.76

The spectra presented evidence for the coordination between the metal ions and flavonoid molecules. Some features of the spectra are shown in Fig.4 and 5.

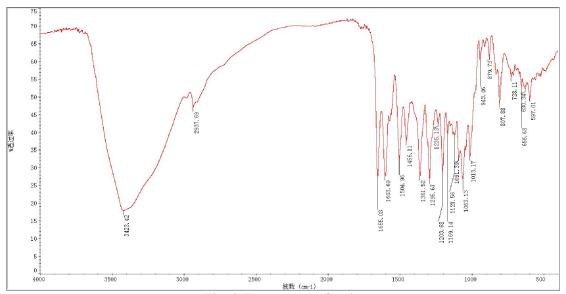


Fig. 4. IR spectrum of rutin

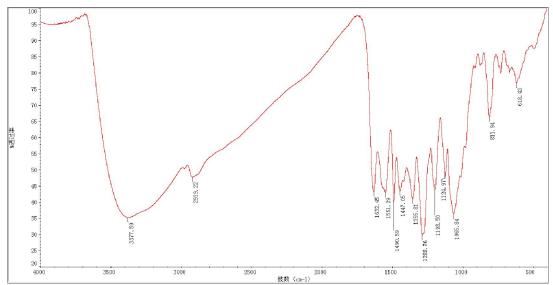


Fig. 5. IR spectrum of the rutin-ca complex

The characteristic stretching v(C=O) mode of the rutin occurred at 1655.05 cm<sup>-1</sup>, while due to the

formation of the rutin-ca complex, this band shifted to 1632.45cm<sup>-1</sup>. It suggested that the Ca (II) coordination

occurred through the carbonyl oxygen atom and the 5-OH group of the rutin. The shift of v (C-O-C) vibration frequency was much slighter (6.91cm<sup>-1</sup> from 1295.67cm<sup>-1</sup> of rutin to 1288.76cm<sup>-1</sup> of rutin-Ca(II) complexes) than that of v (C=O) vibration frequency (obviously declines by 22.58cm<sup>-1</sup>) indicating that the ring oxygen was not involved in the complexation. There was also a decline of 52.40cm<sup>-1</sup> for the v (C=C) vibration frequency because the conjugative effect was obviously enhanced after the chelation occurring [33]. The big bound of v (O-H) vibration frequency (from 3423.42 to 3377.50cm<sup>-1</sup>) indicated the existence of water in the complex [34].

## 3.4 H NMR spectroscopy analysis

The <sup>1</sup>H NMR(400 MHz, DMSO) data of rutin

and its complex with Ca(II) are given bellow separately:

Rutin: 12.630 (5-OH), 10.831 (7-OH), 9.646 (4'-OH), 9.186(3'-OH), 7.55 (2'-H, 5'-H, 6'-H), 6.836(8-H), 6.388(6-H) [35]

Rutin-Ca complex: 10.306(7-OH), 9.049 (3'-OH), 7.520(2'-H), 7.404(6'-H), 7.294 (5'-H), 6.470(8-H), 6.315(6-H).

The  $^{1}$ H NMR data shows that the  $\delta$  values of rutin-ca complex were shifed to higher field compared with the pure rutin. It maybe due to the increase of the conjuation caused by the coordination. The absece of 5-OH and 4'-OH reveals that they were involved in the coordination when the complex is formed  $^{[36]}$ .

Table 2 <sup>1</sup>H NMR (cm<sup>-1</sup>) of rutin and rutin-Ca (II) complex

	Chemical shift (ppm)								
	5-OH	7-OH	4'-OH	3′-OH	2′-H	6′-H	5′-H	8-H	6-H
Rutin	12.631	10.831	9.646	9.186	7.590	7.561	7.501	6.836	6.388
Rutin-Ca		10.306		9.049	7.520	7.404	7.294	6.470	6.315

### 3.5 The differential thermal analysis

Thermal methods of analysis provided interesting ways for the investigation of metal complexes. Heating of material caused occurrence of chemical and physical transformations, which were accompanied by the absorption or liberation of heat [37]. The stability and structure of the complex could be determined by the thermogravimetric analysis [38]. Table 3 showed the maximum temperature values for decomposition along with the corresponding weight loss values. Thermogravimetric analysis was carried out for rutin-Ca (II) complex under the flow of air. Fig.7 showed the characteristic thermal events those change with heating

rate of  $10^{\circ}$ C/min and the temperature range was from room temperature to  $784^{\circ}$ C.

Thermal analysis of the rutin-Ca (II) complex showed two distinct breaks i.e. dehydration and decomposition. It was found the weight loss was 4.83% below 77°C which might be attributed to the liberation of two moles of hydrated water molecules. When the complex was heated to 182°C, the weight loss was 9.12% which might be due to loss of the four coordinated water molecules. This fact suggested that the rutin-Ca complex contains four coordinated water molecules and two crystal water molecules.

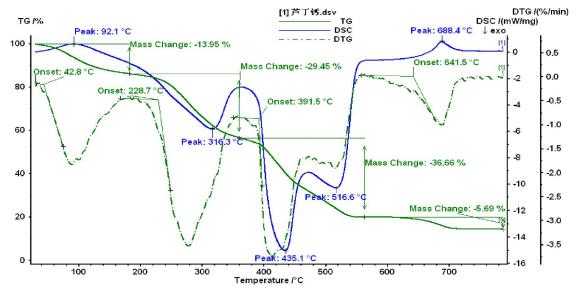


Fig.6 Thermogravimetric analysis of rutin-Ca complex

rutin-Ca compplex									
Compound	Mass(g/mol)	Decomposition Eliminated		%mass loss					
		$T_{max}(^{\circ}C)$	species	Found	calculated				
	First step	82	2H <sub>2</sub> O	4.83	4.52				
[C <sub>2</sub> (L)(H <sub>1</sub> O) 1 2H <sub>2</sub> O	Second step	182	4H <sub>2</sub> O	9.12	9.05				
[Ca <sub>2</sub> (L)(H <sub>2</sub> O) <sub>4</sub> ]·2H <sub>2</sub> O (795.66g/mol)	Third step	350	$C_{12}H_{10}O_4$	29.45	27.40				
	Fourth step	570	$C_{12}H_{21}O_{9}$	36.66	38.83				
	Fifth sten	720	C <sub>2</sub> O	5 69	5.03				

Table 3 The maximum temperature,  $T_{max}(^{\circ}C)$ , and weight loss values of the decomposition stages for the rutin-Ca compplex

The DSC for the complex showed endothermic and exothermic peaks that agreed with the mass losses observed in the TG. The endothermic peaks about 92.1  $^{\circ}$ C are due to dehydration. The exothermic peak about 316.3  $^{\circ}$ C is due to the elimination of pyrocatechol (Fig.7. Part a) and the weight loss at this step is 29.45%.

Residue

The second exothermic peak around 435.1°C may be associated with the oxidative degradation of the remaining organic component (Fig.7. Part b), corresponding to 36.66% weight loss. The final products are the metal oxides.

14.25

14.11

CaO

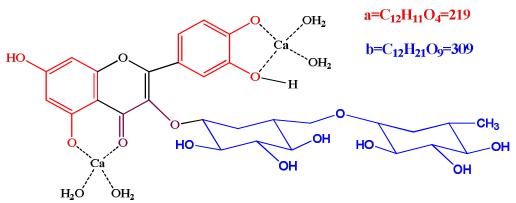


Fig.7. Schematic of Rutin-Ca complex thermal cracking

From this data, we could infer the structure of rutin calcium complex (Fig.8):

Fig.8. Prosposed structure of the Ca (II)-rutin complex

## 3.6 The level of blood-lipid in rats

The serum total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C) and triglyceride (TG) levels of the model group were significantly different

from those of the normal control group and the experimental group (P<0.05). But the serum high-density lipoprotein cholesterol (HDL-C) level of the model group was not significantly different from those

of the normal control group. The serum TC, LDL-C and TG levels of the experimental group were higher than those of the normal control group but lower than those of the model group, and the serum HDL-C level was significantly higher than those of the normal control group and the model group (P<0.05). The main data are summarized in table 3.

#### 3.7 The viscera index of the rats

The main data are summarized in table 4. The liver and spleen index of the model group were significantly higher than those of the normal group and the experimental group (P<0.05), while no significant difference was shown between the normal group and the experimental group. There was no significant difference between cardiac index and renal index.

The liver index of the rats was positively correlated with triglyceride (r=0.453, P<0.05), total cholesterol (r=0.491, P<0.01) and low-density lipoprotein cholesterol (r=0.533, P<0.01), but not obviously correlated with high-density lipoprotein cholesterol (r=0.388, P<0.05). The spleen index of the rats was correlated with TG (r=0.388, P<0.05), but not obviously correlated with TC, HDL-C and LDL-C. The cardiac index and renal index were not obviously correlated with the serum lipid level.

It indicated that the hyperlipidemia induced by lipid emulsions might cause the abnormal enlargement of the liver and spleen, which was similar to the clinical symptom of hyperlipidemia. However, the rutin calcium complex could improve the various indexes of serum lipid and decrease the spleen index and liver index.

#### **4 Conclusion**

There were two ways to prepare the experimental hyperlipemia model. One was to feed the animals with high lipid diet. The other was that the animals were administrated by gastric perfusion of fat emulsion. Of the two, the latter was better than the former, because the latter could assure a coherent condition, and we could regulate the content of cholesterol, lard according to the actual condition. Also the time of model preparation was short. In this experiment, the hyperlipemia model was established successfully by gastric perfusion of fat emulsion.

In this study, disorder of lipometabolism was detected in the model group rats. Compared with those of the normal control group, the serum TC, TG levels of the model group were much higher, and it caused hyperlipidemia rats. But the serum TC, TG and HDL levels of the experimental group indicated that rutincalcium complex was quite effective in reducing serum TC and TG and adjusting blood lipid.

As a part of the research project, a new complex of Ca (II) with rutin was prepared and characterized.

This study examined the interaction of metal ion with rutin in methanolic solution. The spectroscopic data showed the importance of 5-OH group as a coordination site. The complex had been characterized on the basis of analytical and spectral data. The results showed that rutin-calcium complex had the effect of anti-hyperlipemia and protecting pathological changes of organs in rats with experimental hyperlipemia.

## Acknowledgments

The work was financially supported by the innovative experiment project for the undergraduate Students of Zhengzhou University.

#### **Corresponding author:**

Tel.: +86 371 66961670;

E-mail *address*: zhaiguangyu@zzu.edu.cn ( ZHAI Guang-yu)

#### References

- Bushra Sultana, Farooq Anwar. Flavones (kaempeferol, quercetin, myricetin) contents of selected fruits, vegetables and medicinal plants[J]. Food Chemistry, 2008, 108: 879– 884.
- 2. Bent H. Havsteen, The biochemistry and medical significance of the flavonoids, Pharmacology & Therapeutics, 2002, 96: 67
- 3. Jianxiong Yang, Juan Guo, Jiangfeng Yuan. In vitro antioxidant properties of rutin [J]. Food Science and Technology, 2008, 41(6): 1060-1066.
- 4. Mina Ghiasi, Majid M. Herav. Quantum mechanical study of antioxidative ability and antioxidative mechanism of rutin (vitamin P) in solution[J]. Carbohydrate Research, 2011, 346(6): 739-744.
- 5. Rachmat Mauludin, Rainer H. Müller, Cornelia M. Keck. Development of an oral rutin nanocrystal formulation [J]. Int J Pharm., 2009, 370(1-2): 202-209.
- 6. Chen, S., Gong, J., Liu, F., Mohammed, U., Naturally occurring polyphenolic antioxidants modulate IgE-mediated mast cell activation. Immunology, 2000, 100, 471–480.
- 7. Igor B. Afanas'ev, Elena A. Ostrakhovitch, Elena V. Mikhal'chik, Enhancement of antioxidant and anti-inflammatory activities of bioflavonoid rutin by complexation with transition metals, Biochemical Pharmacology, 2001, 61: 677–684.
- 8. Xue Tian, Fengju Li, Lu Zhu, et al., Study on the electrochemical behavior of anticancer herbal drug rutin and its interaction with DNA, Journal

- of Electroanalytical Chemistry, 2008, 621: 1–6
- Panasiak, W., Wleklik, M., Oraczewska, A., et al, Influence of flavonoids on combined experimental infections with EMC virus and Staphylococcus aureus in mice. Acta Microbiol. Pol.1989, 38: 185–188.
- 10. Yu Han, Yan Ding, Dandan Xie,et al.Design, synthesis, and antiviral activity of novel rutin derivatives containing 1, 4-pentadien-3-one moiety, *European Journal of Medicinal Chemistry*, 2015,92: 732-737.
- 11. Anna Baldisserotto, Silvia Vertuani, Alessia Bino, et al. Design, synthesis and biological activity of a novel Rutin analogue with improved lipid soluble properties, *Bioorganic & Medicinal Chemistry*, 2015,23(1): 264-271.
- 12. Sylvain Tranchimand, Pierre Brouant, Gilles Iacazio, The rutin catabolic pathway with special emphasis on quercetinase, Biodegradation, 2010, 21: 833–859.
- 13. Qadeer K. Panhwar, Shahabuddin Memon, Synthesis, characterization and antioxidant study of Tin(II)–rutin complex: Exploration of tin packaging hazards, *Inorganica Chimica Acta*, 2013,407: 252-260.
- 14. Kostyuk, V. A.; Potapovich, A. I.; Kostyuk, T. V. et al. Metal complexes of dietary flavonoids: evaluation of radical scavenger properties and protective activity against oxidative stress in vivo. Cellular and Molecular Biology, 2007, 53(1), 62-69.
- 15. Rachmat Mauludin, Rainer H. Müller, Cornelia M. Keck, Kinetic solubility and dissolution velocity of rutin nanocrystals, european journal of pharmaceutical sciences, 2009, 36: 502–510.
- 16. Rubens F.V. de Souza, Wagner F. De Giovani, Synthesis, spectral and electrochemical properties of Al(III) and Zn(II) complexes with flavonoids, Spectrochimica Acta Part A, 2005, 61: 1985–1990.
- 17. Medvidovic-Kosanovic Martina, Samardzic Mirela, Malatesti Nela, et al. Electroanalytical Characterization of a Copper (II)-Rutin Complex [J]. International journal of electrochemical science, 2011, 6(4): 1075-1084.
- 18. Lu-Ming Lin, Han-Ying Wu, Wei-Shuen Li, et al., Kinetic studies of the oxidation of quercetin, rutin and taxifolin in the basic medium by (ethylenediaminetetraacetato) cobalt(III) complex , Inorganic Chemistry Communications, 2010, 13: 633 635.

- 19. Kang, Jingwan; Su, Biquan; Lu, Xiaoquan. Synthesis and characterization of coordination compounds of Cd(II), Co(II), Ni(II), Cu(II) and Zn(II) with rutin. Indian Journal of Chemistry, Section A: Inorganic, Bio-inorganic, Physical, Theoretical & Analytical Chemistry 2005, 44A(10): 2010-2014.
- Selvaraj Stalin; Krishnaswamy Sridharan; Devashya Venkappayya; et al.Synthesis, characterization and DNA binding properties of rutin-iron complex , RSC the Royal Society of Chemistry Advances, 2012, 2 (7): 2797-2802.
- 21. Nowak Dorota; Kuzniar Anna; Kopacz Maria, Solid complexes of iron(II) and iron(III) with rutin, Structural Chemistry, 2010, 21 (2): 323-330.
- 22. Franzoi Ana Cristina, Peralta Rosely A, Neves Ademir, et al. Biomimetic sensor based on Mn(III)Mn(II) complex as manganese peroxidase mimetic for determination of rutin [J]. Talanta, 2009, 78(1): 221-226.
- 23. Uivarosi Valentina; Barbuceanu Stefania Felicia; Aldea Victoria; et al. Synthesis, Spectral and Thermal Studies of New Rutin Vanadyl Complexes, Molecules, 2010,15 (3): 1578-1589.
- Vesna S. Kuntic', Dus'an L. Males 'ev, Zorica V. Radovic', et al., Spectrophotometric Investigation of Uranil(II)-Rutin Complex in 70% Ethanol, Agric. Food Chem. 1998, 46:5139 –5142.
- 25. A, S. A. Golovanova, N. V. Lariontseva, et al. Protective properties of gold in the composition of nanoparticles and Au—rutin complexes, Russian Chemical Bulletin, 2011, 60 (4): 667-670.
- Michael H. Gordon, Significance of Dietary Antioxidants for Health, International Journal of Molecular Sciences, 2012, 13: 173-179;
- 27. Stavros K. Dimitriou, Spyros P. Perlepes, Aris Terzis, Synthesis, crystal structures and spectroscopic studies of calcium(II) succinamate(-1) complexes, Polyhedron, 2010, 29 (10): 2213-2219.
- 28. Yoko Sakanashi, Keisuke Oyama, Hiroko Matsui, et al. Possible use of quercetin, an antioxidant, for protection of cells suffering from overload of intracellular Ca2+: A model experiment,Life Sciences, 2008,83 (5-6): 164-169.
- 29. Kristoffer Heindorff, Wolfgang Blenau, Bernd Walz, et al. Characterization of a Ca2+/calmodulin-dependent AC1 adenylyl cyclase in a non-neuronal tissue, the blowfly salivary gland, Cell Calcium, 2012,52 (2):

- 103-112.
- 30. Sheng-Nan Wu, Hung-Ting Chiang, Ai-Yu Shen, et al. Differential effects of quercetin, a natural polyphenolic flavonoid, on L-Type calcium current in pituitary tumor (GH3) cells and neuronal NG108-15 cells, Journal of Cellular Physiology, 2003,195 (2): 298–308.
- 31. Yury S. Tarahovsky, Elena A. Yagolnik, Eugeny N. Muzafarov, et al. Calcium-dependent aggregation and fusion of phosphatidylcholine liposomes induced by complexes of flavonoids with divalent iron, Biochimica et Biophysica Acta (BBA) -Biomembranes, 2012, 1818 (3): 695–702.
- 32. Paolo Matteini, Andrea Goti, Giovanni Agat, Theoretical conformational analysis of rutin, Monatsh Chem, 2010, 141: 793–800.
- Sergio A Payán-Gómez, Norma Flores-Holguín, Antonino Pérez-Hernández, et al. Computational molecular characterization of the flavonoid rutin, Chemistry Central Journal 2010, 4:12-17.
- 34. Alexander V. Pastukhov, Lidiya A. Levchenko, Anatoli P. Sadkov, Spectroscopic study on binding of rutin to human serum albumin,

- Journal of Molecular Structure, 2007, 842: 60–66
- Agnieszka Kicel, Aleksandra Owczarek, Piotr Michel, et al. Application of HPCCC, UHPLC-PDA-ESI-MS and HPLC-PDA methods for rapid, one-step preparative separation and quantification of rutin in *Forsythia* flowers, *Industrial Crops and Products*, 2015,76: 86-94.
- M. Samsonowicz, I. Kamińska, M. Kalinowska, et al. Alkali metal salts of rutin Synthesis, spectroscopic (FT-IR, FT-Raman, UV–VIS), antioxidant and antimicrobial studies, Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 2015,151: 926-938.
- 37. Ticiano Gomes do Nascimento, Everaldo Moreira da Costa, Jose' Maria Barbosa Filho, Thermal characterization of the quercetin and rutin flavonoids, Thermochimica Acta, 2002, 392–393: 79–84.
- 38. Buchner, Angelika Krumbein, Sascha Rohn, et al. Effect of thermal processing on the flavonols rutin and quercetin, Rapid Communications in Mass Spectrometry, 2006, 20 (21): 3229–3235.

6/18/2016