

# **Bioorthogonal non-canonical amino acid tagging - BONCAT -**

# **First, some definitions**

**bioorthogonal**

**non-interacting with cellular functionalities**

**non-canonical**

**synthetic, not part of biological machinery**

**Click chemistry**

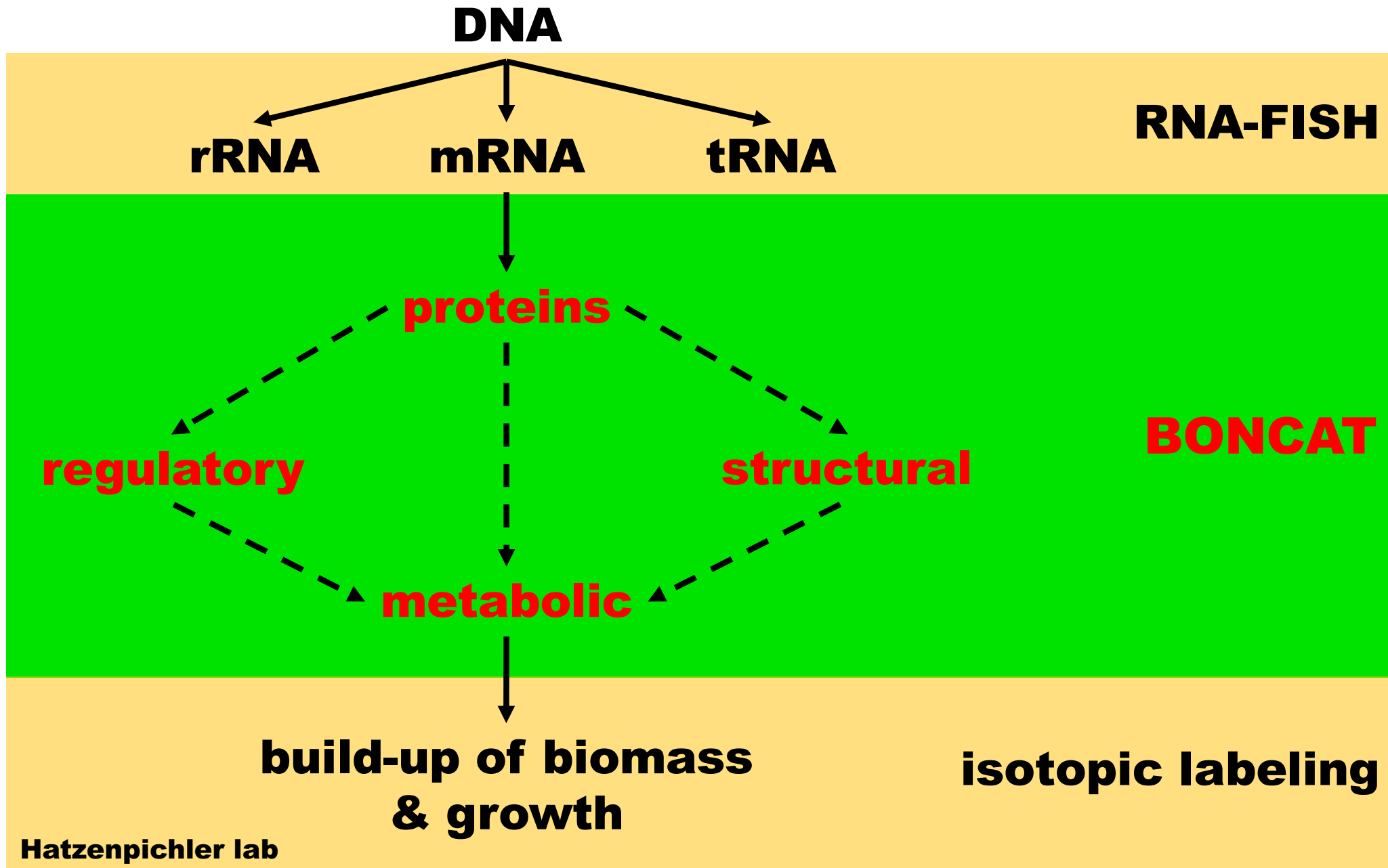
**complete conversion of reagents to single product**

**+ mild conditions**

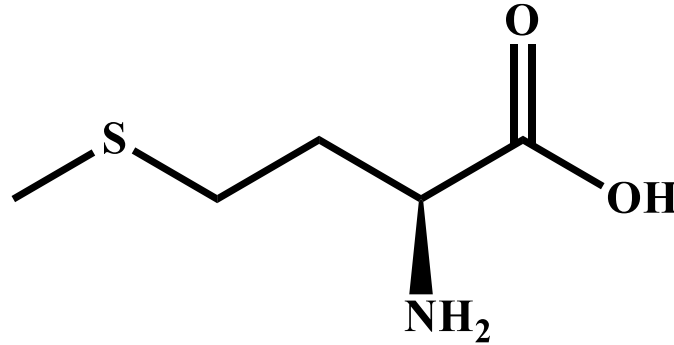
**+ very fast**

**+ in water**

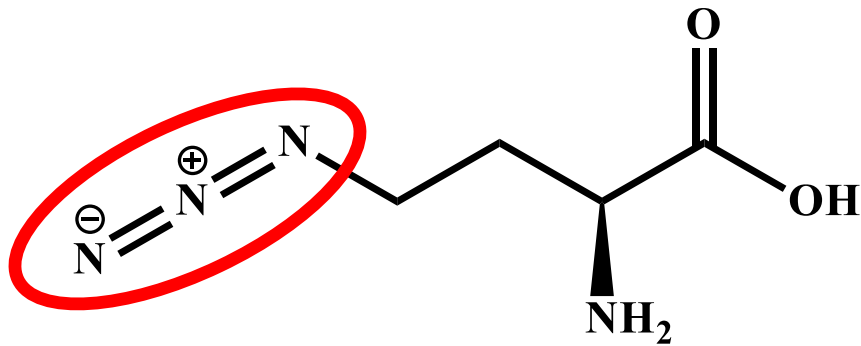
# Activity assays on individual cell level



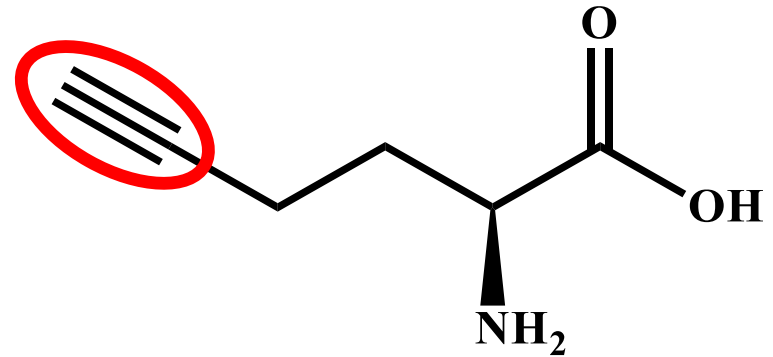
# Bioorthogonal non-canonical amino acids



**L-Methionine**  
**Met**



**L-Azidohomoalanine**  
**AHA**



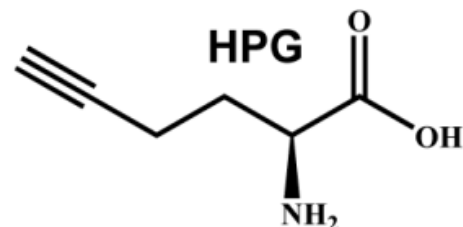
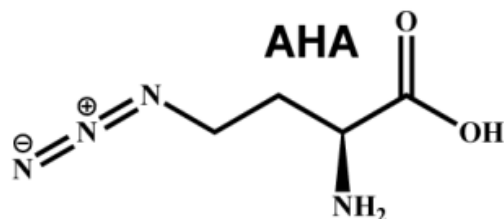
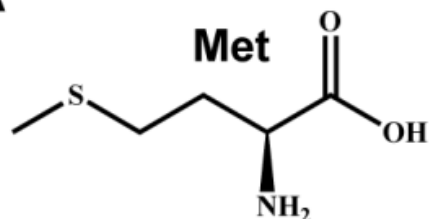
**L-Homopropargylglycine**  
**HPG**

**synthetic amino acids**  
**incorporate into new proteins instead of Met**

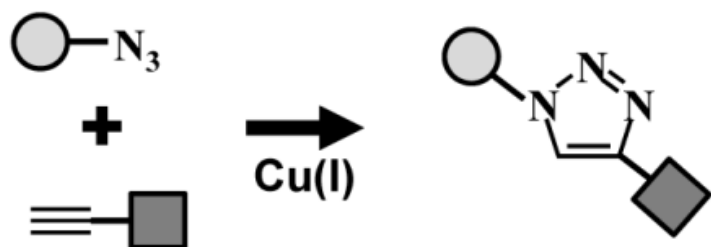


# Azide-alkyne click reactions

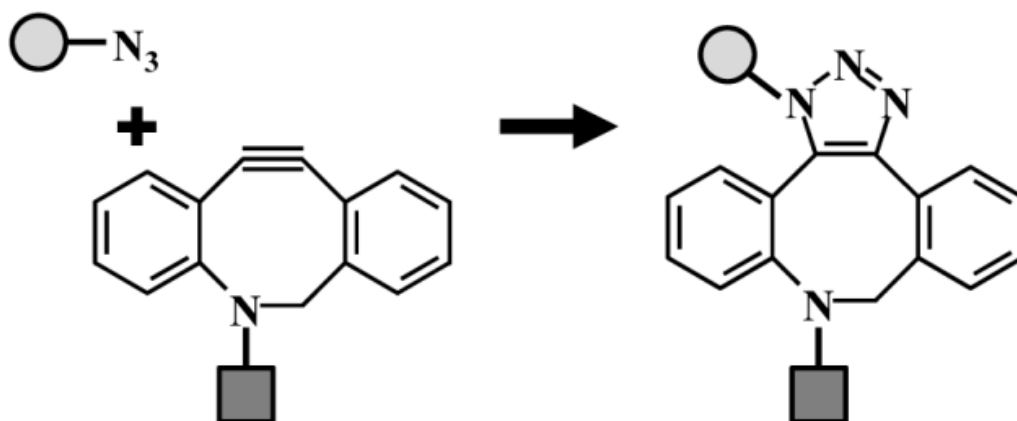
**A**



**B**



**C**

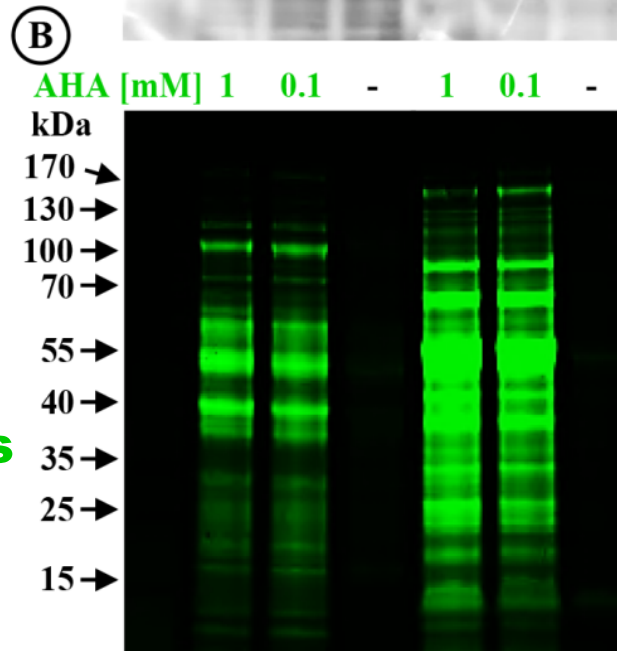
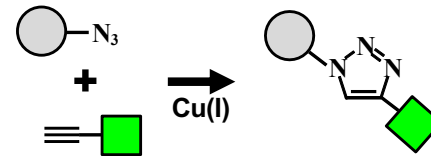
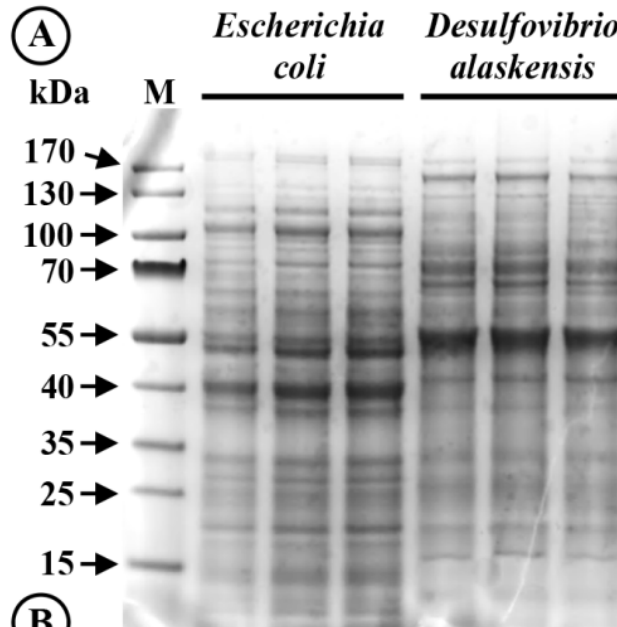


**A. Structures of Met and its surrogates AHA and HPG, which compete with Met during translation.**

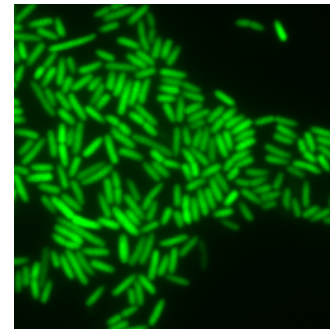
**B. In Cu(I)-catalyzed click chemistry an azide group (N<sub>3</sub>) is linked to a terminal alkyne residue, yielding a triazole conjugate.**

**C. Strain-promoted click chemistry allows the copper-less conjugation of an azide group (N<sub>3</sub>) with a cyclo-octyne-carrying molecule, yielding a triazole conjugate.**

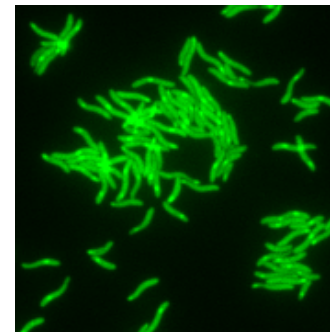
# Incorporation into newly made proteins



new proteins

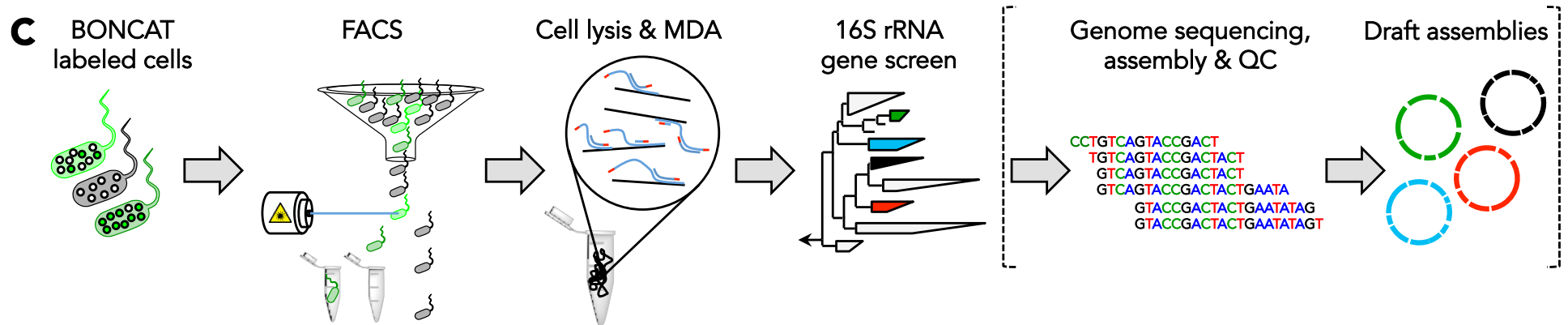
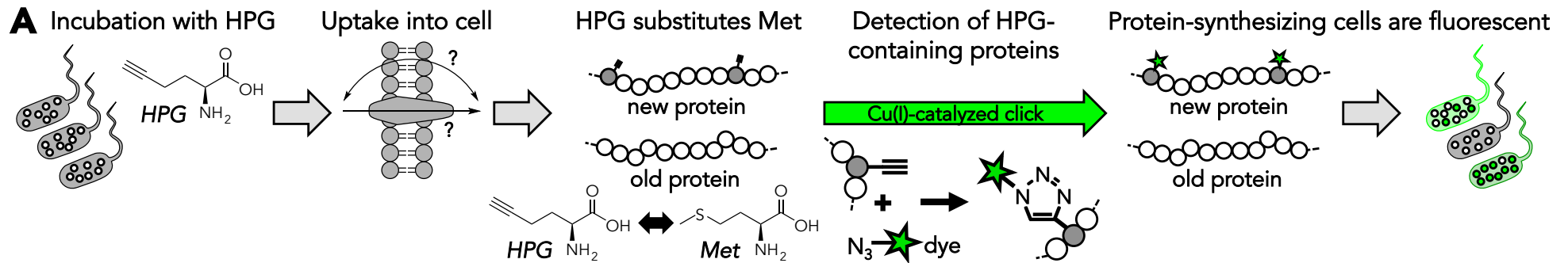


*E. coli*  
respiring glucose

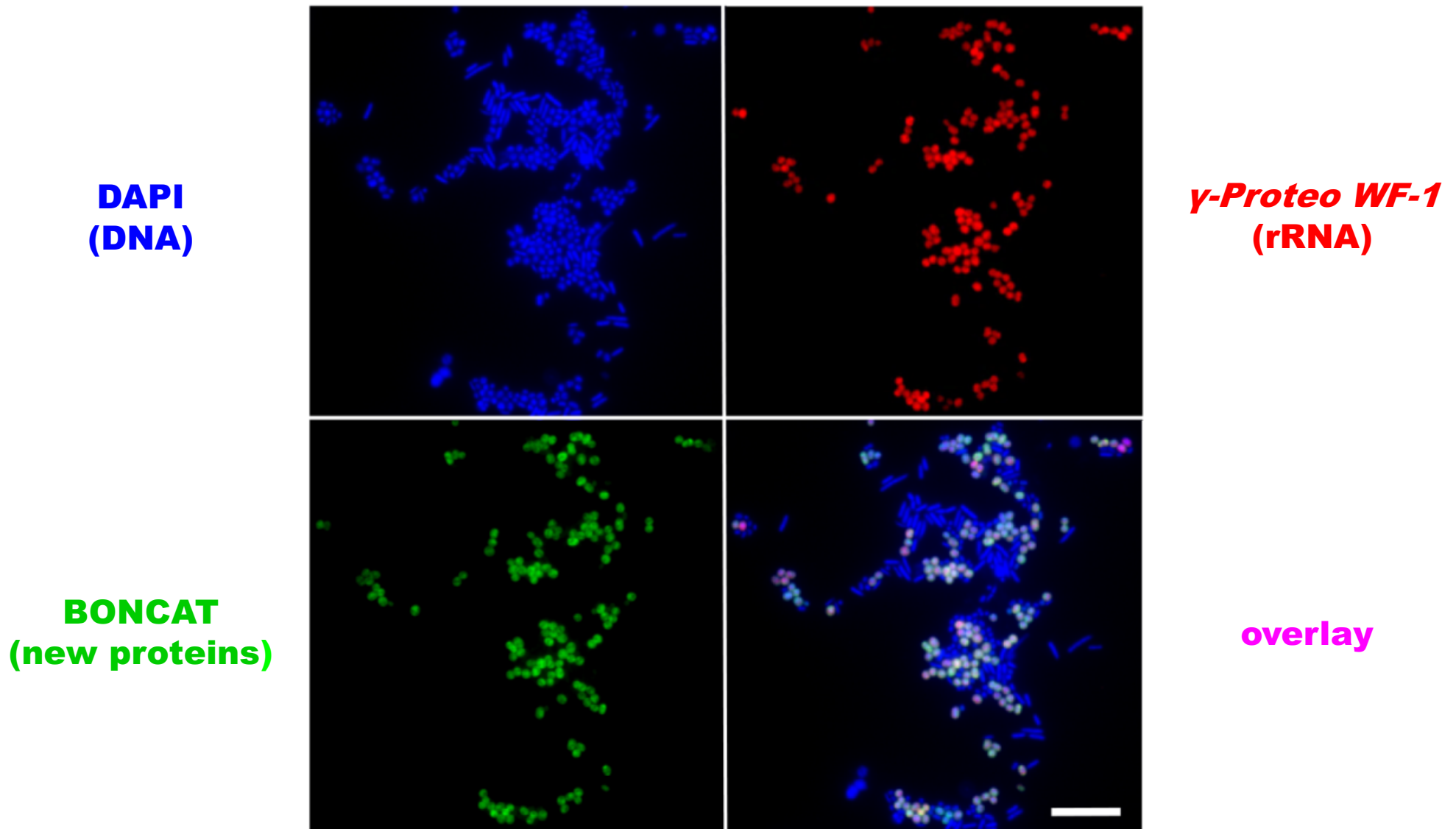


*D. alaskensis*  
sulfate reducer

# Visualizing, identifying, and sorting translationally active microbes



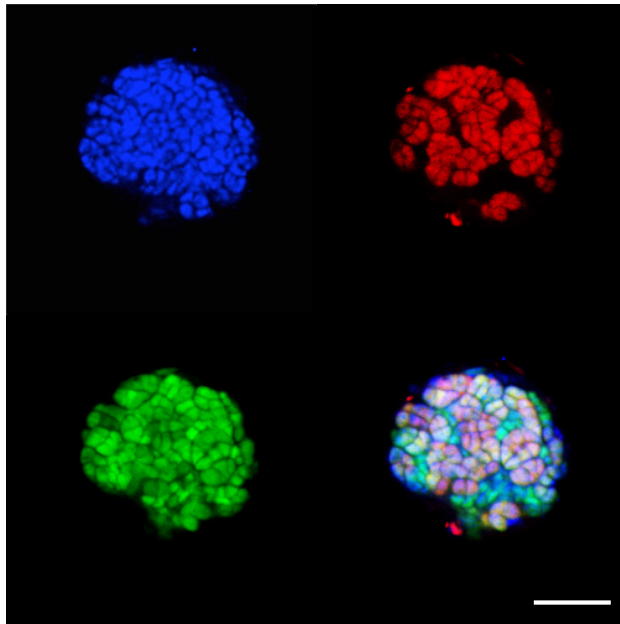
# Identification of translationally active cells



Bar = 10  $\mu$ m

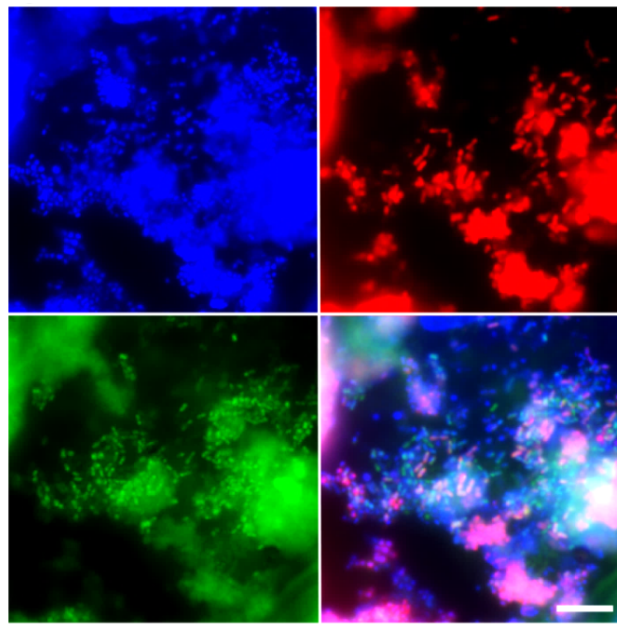
# BONCAT-FISH of uncultured microbes

**Arch915**



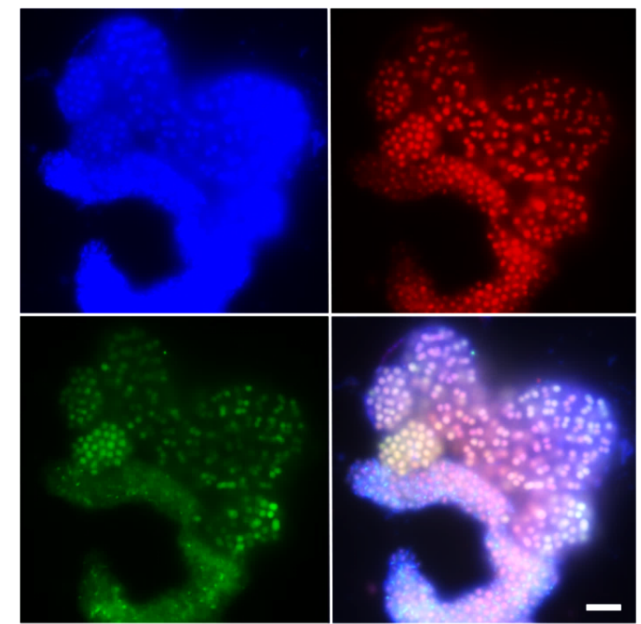
**Methane seep  
ANME-SRB consortium**

**EUB338 I-III**



**Tongue biofilm and saliva**

**Gam42a + competitor**



**Freshwater from Lily  
pond on Caltech campus**

**DAPI  
(DNA)**

**BONCAT  
(new proteins)**

**FISH  
(rRNA)**

**Overlay**

**Hatzenpichler *et al.*, 2014; Hatzenpichler *et al.*, 2015; Hatzenpichler *et al.*, 2016**

**Hatzenpichler lab**

# Visualizing new proteins *in situ*

**generally applicable  
(works for all taxonomies and  
physiologies tested so far)**

**detectable after 2%  
of generation time**

**FISH-BONCAT links function  
and identity of a cell**

**BONCAT correlates with  
 $^{15}\text{NH}_3$  incorporation (nanoSIMS)**

**no change in protein expression  
(Bagert *et al.*, 2014)**

**Hatzenpichler lab**



**Hatzenpichler *et al.*, 2014**

# **Limitations and advantages of BONCAT-FISH**

**uptake and incorporation**

**Methionine-rich samples are tough**

**hard to quantitate amount of new proteins in uncultured cells**

**potential for cell inactivation or community shifts**

**links cellular identity and function**

**fluorescence-based *in situ* activity studies**

**metabolic screening**

**activity-based cell-sorting**

**fast + highly selective + cheap + easily available**

**1 h      azide-alkyne      ~\$500      epi-scope**



# BONCAT in environmental microbiology (as of Jul 2019)

**Hatzenpichler R et al.**

***In situ* visualization of newly synthesized proteins in environmental microbes using amino acid tagging and click chemistry**

Environ Microbiol, 16: 2568-2590 (2014)

- *first application of BONCAT to uncultured microbes in the environment; development of BONCAT-FISH; correlation of BONCAT with nanoSIMS*

**Samo TJ et al.**

**Broad distribution and high proportion of protein synthesis active marine bacteria revealed by click chemistry at the single cell level**

Front Mar Sci, doi: 10.3389/fmars.2014.00048 (2014)

- *application of BONCAT to seawater; correlation of BONCAT with MAR*

**Hatzenpichler R and Orphan VJ**

**Detection of protein-synthesizing microorganisms in the environment via bioorthogonal non-canonical amino acid tagging (BONCAT)**

Book chapter for Hydrocarbon and Lipid Microbiology Protocols, Vol. 7: Single-cell and single-molecule methods

Springer Protocols Handbooks, doi 10.1007/8623\_2015\_61 (2015)

- *description of how to design and perform BONCAT-experiments using AHA and HPG*

**Hatzenpichler R et al.**

**Visualizing *in situ* translational activity for identifying and sorting slow-growing archaeal-bacterial consortia**

Proc Natl Acad Sci USA, 113: E4069-E4078 (2016)

- *development of activity-based cell-sorting via bioorthogonal labeling; applied BONCAT-FISH and BONCAT-FACS to deep-sea sediment consortia*

**Pasulka AL et al.**

**Interrogating marine virus-host interactions and elemental transfer with BONCAT and nanoSIMS-based methods**

Environ Microbiol, DOI: 10.1111/1462-2920.13996 (accepted) 2017

- *first application of BONCAT to viruses; estimate of marine viral production rates by BONCAT and nanoSIMS*

**Leizeaga et al.**

**Using Click-Chemistry for Visualizing *in Situ* Changes of Translational Activity in Planktonic Marine Bacteria**

Front Aquat Microbiol, doi: 10.3389/fmicb.2017.02360 (2017)

**Couradeau et al.**

**Probing the active fraction of soil microbiomes using BONCAT-FACS**

Nat Comm, 10: 2770 (2019)

**Hatzenpichler lab**