Evaluation of raw pork as a commercially manufactured diet option for zoo-managed African wildcats (*Felis silvestris lybica*)

C. J. Iske,* C. L. Morris,^{†1} and K. L. Kappen[†]

*Department of Animal Sciences, Iowa State University, Ames, 50011; and †Omaha's Henry Doorly Zoo & Aquarium, Omaha, NE, 68107

ABSTRACT: Second to beef, pork is a major protein source produced in the US. Properly sourced and handled pork could be utilized as a protein option for zoo-managed carnivores. Concerns of high levels of microbial populations in raw meat diets are common. The objectives of this study were to determine apparent total tract macronutrient and energy digestibility and fecal scores from cats fed a commercially manufactured raw pork-based diet compared with commercially available raw carnivore diets formulated with either horse or beef and evaluate typical microbial population variation among the diets. Dietary treatments consisted of 4 raw meat-based diets: Horse, Beef, Pork, and beef/ horse Blend. All diets were highly digestible, especially fat digestibility (98.6 to 99.7%) in which there were no statistical differences among diets. Digestibility

of organic matter (OM) was greater (P = 0.05) when cats consumed the Blend diet (97.2%) compared to the Pork diet (93.1%). Fecal scores ranged from 1.6 to 2.6 (on a 5-point scale), with Beef (2.6) being greater than (P = 0.01) Horse (1.6) and (P = 0.02) Pork (1.9). E. coli counts ranged from 110 to 10,000 cfu/g; total coliforms: 150 to 28,000 cfu/g; yeast: 20 to 4,000 cfu/g; mold count: not detectable to 10 cfu/g; and aerobic plate count: 23,000 to 26,000,000 cfu/g. Staphylococcus aureus was not detected in any of the diets. Salmonella was presumptive positive in the Pork and Blend diet, and was negative in the other 2 diets. In conclusion, commercially manufactured diets have varying microbial counts. All diets, including the raw pork-based diet were well utilized by exotic small cats and can be included among dietary options for managed felids.

Key words: African wildcat (Felis silvestris lybica), digestibility, microbes, pork, raw meat diet

© 2017 American Society of Animal Science. This is an open access article distributed under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

Transl. Anim. Sci. 2017.1: doi:10.2527/tas2017.0047

INTRODUCTION

Managed small exotic cat species in the United States are typically fed combinations of extruded or canned commercial cat foods, commercial raw meat formulations, and whole prey. Raw diets are more digestible than canned or extruded cat foods mainly due to reduced nutrient quality caused by processing (Kendall et al., 1982; Crissey et al., 1997; Vester et al., 2010b; Kerr et al., 2012). Raw meat diets provide a more natural, bioavailable diet compared to processed cat foods, and avoid concerns facing exclusive use of whole prey

¹Corresponding author: cherylm@omahazoo.com

Received July 26, 2017.

Accepted August 19, 2017.

feeding such as public perception and meeting nutrient requirements (Kerr, 2012; Robinson, 1998). Beef and horse-meat comprise the majority of raw diet formulations commercially manufactured for exotic carnivores in the US. Previously, our research team reported a raw pork diet was well utilized by large exotic felids when palatability and digestibility were measured and compared with other common diets (Iske et al., 2016). Previous studies have seen differences in digestibility between large and small cat species, therefore, further evaluation in other species is warranted to provide data that may aid diet purchasing decisions at zoological institutions (Kerr et al., 2013b). Additionally, there are no published regulations or recommendations for allowable microbial loads in raw carnivore diets, but the ability of cats to remain asymptomatic after consuming raw

meat contaminated with Salmonella suggests they tolerate high numbers of microorganisms in their diets (Carter and Quinn, 2000; Finley et al., 2006). Evaluation and presentation of typical microbe ranges observed in raw carnivore diets would also benefit animal managers to understand expected ranges. The objectives of this study were to determine if a commercially available raw porkbased diet produced similar apparent total tract macronutrient digestibility and fecal scores as standard, commercially available zoological carnivore diets formulated with either horse or beef. Results may indicate the use of properly handled raw pork as a dietary option for small managed exotic cats. Additionally, general microbial population screens in dietary treatments were evaluated to determine typical variation among commonly used products. The aims of this study were not to formulate or compare diets varying strictly in protein source, but rather to evaluate and compare a raw pork diet with beef and horse-based diets commonly utilized to assist animal managers in making informed decisions regarding commercial products in the US.

MATERIALS AND METHODS

Animal procedures were approved by the Lee G. Simmons Conservation Park and Wildlife Safari Animal Care and Use Committee (IACUC) before animal experimentation.

Animals

Four nonbreeding African wildcats (*Felis silvestris lybica*; 2 males, 2 females, average age 9.5 yr, average weight 4.8 kg) were used. Cats received an extruded diet prior to study and were individually housed indoors in 9.2 m³ enclosures at the Lee G. Simmons Conservation Park and Wildlife Safari. Park staff provided care and monitoring of health. Water was provided ad libitum. Animals were weighed at the start and conclusion of the study to ensure body weight was maintained.

Diet Composition and Microbes

Four commercially produced raw meat diets formulated to meet or exceed nutrient requirements for domestic cats were used in the study (National Research Council, 2006). Diets were stored frozen at –18°C until 24 h prior to feeding, at which time they were moved to a cooler (2°C). Dietary treatments were sourced from single manufactured lots. Ingredient compositions of diets are listed in Table 1 and chemical compositions of diets are included in Table 2. Treatments included a horse-based [Nebraska Brand, North Platte, NE; Premium Feline Diet (Horse)], beef-based [Nebraska Brand; Special Beef Feline Diet (Beef)], beef and horse-based blend [Triple A Brand Meat Company, Burlington, CO; Complete Feline Diet (Blend)] and pork-based [Carnivore Essentials, Sheboygan Falls, WI; Carnivore Essentials (Pork)]. Pork

 Table 1. Ingredient composition of horse-, beef-, pork-, and horse/beef blend-based raw meat diets fed to managed

 African wildcats¹

Treatment	Ingredient composition
Horse (Nebraska Brand; Premium Feline, Nebraska Packing Inc., North Platte, Nebraska)	Horsemeat, powdered cellulose, dicalcium phosphate, calcium carbonate, Vitamin Premix [Roughage Products, Vitamin E Supplement, Mineral Oil, Niacin Supplement, Biotin, Menadione Sodium Bisulfite Complex (source of Vitamin K Activity), Vitamin A Supplement, Riboflavin, Pyridoxine Hydrochloride, Folic Acid, Calcium Pantothenate, Thiamine Mononitrate, Vitamin D3 Supplement] Trace Mineral Premix (Copper Sulfate, Manganese Sulfate, Ethylenediamine dihydriodide, Sodium Selenite), Choline chloride, taurine, salt
Beef (Nebraska-Brand; Special Beef Feline, Nebraska Packing Inc., North Platte, Nebraska)	Beef, meat by-products, fish meal, soy bean meal, dried beet pulp, calcium carbonate, dicalcium phosphate, dried egg, brewers dried yeast, salt, Vitamin Premix (Choline chloride, vitamin E supplement, niacin, vitamin B-12 riboflavin, folic acid, vitamin A acetate, thiamine mononitrate, d-calcium pantothenate, mineral oil, biotin, pyridoxine hydrochloride, vitamin D-3 supplement), taurine, Trace Mineral Premix, (zinc oxide, manganous oxide, copper oxide, mineral oil, sodium selenite, calcium iodate)
Pork (Sustainable Swine Resources, LLC; Carnivore Essentials, Sheboygan Falls, Wisconsin)	 Pork, Pork By-products, Vitamins [Beet Pulp, Cellulose, Calcium Carbonate, Rice Hulls, Sodium Chloride, Mineral Oil, Vitamin E Supplement, d-alpha-Tocopheryl Acetate (Source of Natural Vitamin E), Biotin, Niacin Supplement, Thiamine Mononitrate, Vitamin B12 Supplement, Vitamin A Acetate, Vitamin D3 Supplement, Pyridoxine Hydrochloride, Riboflavin Supplement, d-Calcium Pantothenate, Folic Acid], Minerals (Beet Pulp, Cellulose, Calcium Carbonate, Rice Hulls, Mineral Oil, Choline Chloride, Calcium Phosphate, Magnesium Oxide, Potassium Chloride, Ferrous Sulfate, Zinc Sulfate, Copper Sulfate, Manganese Sulfate, Zinc Oxide, Sodium Selenite, Cobalt Carbonate, Calcium Iodate)
Blend (Triple A Brand Meat Company; Feline Complete Diet, Burlington, Colorado)	

¹Ingredient compositions are listed as given by manufacturers of each diet.

Translate basic science to industry innovation

Ŕ

 Table 2. Chemical composition of horse-, beef-, pork-, and horse/beef blend-based raw meat diets fed to managed African wildcats (DMB)

8	(/		
Item ¹	Horse	Beef	Pork	Blend
DM, % as-fed	36.9	34.6	33.0	28.7
OM, %	90.4	91.3	94.0	91.1
СР, %	50.3	52.4	51.0	65.7
Fat, %	31.6	33.1	39.9	25.4
NFE, %	1.1	0.3	0	0
CF, %	3.1	2.3	2.4	1.2
TDF, %	7.3	5.5	5.6	1.2
GE, kcal/g	6.1	6.3	6.7	6.1

 1 DM = dry matter; OM = organic matter; CP = crude protein; NFE = nitrogen free extract; CF = crude fiber; TDF = total dietary fiber; GE = gross energy.

was sourced from a bio-secure, integrated swine operation. Each dietary treatment was subsampled once, dried at 55°C, ground through a 2-mm screen (Wiley mill, model 3383-L10, Thomas Scientific, Swedesboro, NJ) and analyzed for chemical composition. Dietary treatments were analyzed for dry matter (DM; Method 934.01) and organic matter (OM; Method 942.05; AOAC, 2006). Crude protein (CP) was determined using a Leco Nitrogen/ Protein Determinator (Method 992.15; model FP-528, Leco Corporation, St. Joseph, MI). Fat concentrations were determined by hexane extraction (Method 991.36; AOAC, 1995). Gross energy (GE) was determined by bomb calorimetry (model AC 500, Leco Corporation, St. Joseph, MI). Crude fiber (CF) was analyzed by Midwest Laboratories (Omaha, NE; AOCS Ba6a-05; Thiex, 2008). Total dietary fiber (TDF) was adjusted for high protein samples (Prosky et al., 1994). One representative subsample per diet was collected, frozen at -18° C, and transported without thawing to Midwest Laboratories (for microbial evaluations; Method 991.14; Method 2003.07; AOAC, 1994; Maturin and Peeler, 2001; Tournas et al., 2001; AOAC, 2003). Salmonella and Escherichia coli (E. Coli) were analyzed for microbial presence of serotypes including non-pathogenic strains (presumptive positive or negative). All chemical analyses were conducted in the nutrition lab at Omaha's Henry Doorly Zoo and Aquarium (Omaha, NE) unless otherwise noted.

Experimental Design

The experimental design was a 4×4 Latin square with animals randomly assigned to 1 of 4 dietary treatments. Animals were fed treatment diets isocalorically for GE to maintain body weight, determined prior to start of study. Each treatment period consisted of a 10 d diet adaptation phase followed by a 4 d collection phase. Total food intake, fecal output and fecal scores were obtained daily during each collection phase. Feces were evaluated using a scale of 1 to 5 with: 1 = hard, dry pellets; 2 = dry, well-formed; 3 = soft, moist, formed; 4 = soft, unformed; 5 = watery liquid (Felid Taxon Advisory Group, 2014).

Energy and Macronutrient Digestibility

Fecal samples per animal were pooled at conclusion of each period, dried at 55°C, and ground through a 2-mm screen (Wiley mill, model 3383-L10, Thomas Scientific). Fecal samples were analyzed for DM, OM, CP, GE, and fat concentrations using methods previously described for diet analyses. Apparent total tract digestibility values were calculated using the following equation: [(nutrient intake – fecal output) / (nutrient intake)] × 100. Digestible energy (DE) values were calculated using the following equation: (kcal/g gross energy in diet \times energy digestibility of respective diet). For method comparison, metabolizable energy (ME) of diets were calculated using the published National Research Council (NRC) equation [ME = DE – $(0.77 \times \text{g protein in diet})$] as well as using modified Atwater values (8.5 kcal/g fat, 3.5 kcal/g protein, 3.5 kcal/g carbohydrate) and unmodified Atwater values (9.0 kcal/g fat, 4.0 kcal/g protein, 4.0 kcal/g carbohydrate) multiplied by fat, protein, and carbohydrate concentration of each diet (National Research Council, 2006). Carbohydrate concentrations of diets were calculated by difference as nitrogen free extract (NFE) using the following equation (DMB): (100 - (% ash + % CP +% fat + % TDF). Although crude fiber is typically used in calculating NFE, more accurate measures of dietary fiber result in more accurate estimations of NFE; therefore, TDF was used for calculating NFE (Cho et al., 2001; de-Oliveira et al., 2012). Due to very low fiber and NFE concentrations in diets and high relative error, NFE of some diets produced a slightly negative value; therefore, a value of 0 was used for NFE.

Statistical Analysis

All data were analyzed using the Mixed Models procedure of SAS (SAS Inst. Inc., Cary, NC). The fixed effect of diet was tested and cat was considered a random effect. All data were reported as least squared means and a probability of P < 0.05 was considered statistically significant. Reported standard error of the means (SEM) were determined according to the Mixed Models procedure of SAS. Data of 1 animal was not included for periods 2 (Blend) and 4 (Horse) because of lack of intake (less than 30% of daily offered diet for 3 consecutive days at any point during the study). This animal did consume adequate amounts (more than 30% of daily offered diet) for periods 1 (Beef) and 3 (Pork); therefore, data were included for those 2 periods with the exception of fat digestibility because of lack of adequate sample volume for fecal fat determination.

RESULTS

Diet Composition

Diets contained similar concentrations of OM (90.4 to 94.0%) and GE (6.1 to 6.7 kcal/g DM), but were variable in other nutrients (Table 2). The Blend diet had the lowest DM (28.7%) and fat (25.4%) concentrations and highest CP (65.7%) concentrations. The other diets (Horse, Beef, and Pork) ranged in DM from 33.0 to 36.9% and CP from 50.3 to 52.4%. The Pork diet was highest in fat concentration (39.9%) compared to other diets. Fiber concentrations (CF and TDF) were highly variable among diets ranging from a low in the Blend diet (1.2 and 1.2%, respectively) to a high in the Horse diet (3.1 and 7.3%, respectively; Table 2).

Diet Microbes

Microbial counts were variable among diets and can be found in Table 3. *E. coli* and total coliform counts were numerically highest in the Beef diet (10,000 and 28,000 cfu/g, respectively) and numerically lowest in the Blend diet (110 and 150 cfu/g, respectively). The Beef diet tested positive for mold (10 cfu/g) while all other diets did not contain detectable levels of mold. Yeast and aerobic plate counts were greatest in the Blend diet (4,000 and 26,000,000 cfu/g, respectively), while yeast was lowest in the Pork diet (20 cfu/g). *Staphylococcus aureus* was not detected in any of the diets (10 cfu/g reporting limit). *Salmonella* was presumptive positive in Pork and Blend diets, and was negative in the other two diets.

Energy and Macronutrient Digestibilities

Average diet intakes and fecal output and characteristics are found in Table 4. Dry matter intakes (g/d) and GE intake (kcal/d DMB) of all diets were not different (P > 0.05). With the exception of the single animal removed **Table 4**. Intake, fecal output, fecal characteristics, and apparent total tract macronutrient digestibility in managed African wildcats fed horse-, beef-, pork-, and horse/beef blend-based raw meat diets

Diet						
Item ¹	Horse	Beef	Pork	Blend	SEM	
Intake						
Food intake, g DM/d	35.3	46.4	40.9	50.7	11.2	
GE intake, kcal/d DMB	209.7	292.8	275.0	305.8	70.7	
Fecal output						
Fecal output, g as-is/d	8.8 ^{a,b}	15.3 ^a	8.8 ^{a,b}	4.1 ^b	4.0	
Fecal output, g DM/d	5.4	5.4	4.6	2.8	1.7	
Fecal scores ²	1.6 ^b	2.6 ^a	1.9 ^b	2.2 ^{a,b}	0.3	
Apparent nutrient digestibili	ity					
DM, %	87.6 ^a	88.9 ^{a,b}	90.1 ^{a,b}	94.1 ^b	2.5	
OM, %	91.1 ^a	92.2 ^a	93.1 ^a	97.2 ^b	1.8	
СР, %	96.8 ^b	93.4 ^a	96.0 ^{a,b}	98.1 ^b	1.3	
Fat, %	98.6	99.4	99.4	99.7	0.5	
GE, %	92.1 ^a	93.3 ^a	94.5 ^{a,b}	97.3 ^b	1.7	
DE ³ , kcal/g	5.6	5.9	6.4	6.0		
Calculated ME, kcal/g4	4.5	4.7	5.2	4.5		
Calculated ME, kcal/g ⁵	4.9	5.1	5.6	4.9		
Calculated ME, kcal/g ⁶	5.2	5.5	6.0	5.5		

^{a,b}Means within a row lacking a common superscript letter are different (P < 0.05).

 ${}^{1}GE =$ gross energy; DE = digestible energy; ME = metabolizable energy; DMB = dry matter basis.

²Fecal scores were evaluated using a scale of 1 to 5 with: 1 = hard, dry pellets; 2 = dry, well-formed; 3 = soft, moist, formed; 4 = soft, unformed; 5 = watery liquid.

 ^{3}DE = calculated using following equation: kcal/g gross energy × energy digestibility.

 ^{4}ME = Calculated using modified Atwater: 8.5 kcal of ME/g of fat + 3.5 kcal of ME/g of CP + 3.5 kcal of ME/g of N-free extract.

 ^{5}ME = Calculated using unmodified Atwater: 9 kcal of ME/g of fat + 4 kcal of ME/g of CP + 4 kcal of ME/g of N-free extract.

 ^{6}ME = Calculated using NRC equation: DE – (0.77 × g protein of diet).

from the study for 2 periods, all animals maintained body weight within 5% throughout the study. Fecal dry matter outputs (g/d) were not different among animals but as-is

Table 3. Microbial population counts of commercially available horse-, beef-, pork-, and horse/beef blend-based raw meat diets fed to managed African wildcats¹

	Cfu ² /g					Org ³ /25g	
Diet	E. Coli (generic)	Total coliforms	Yeast	Mold count	Staphylococcus aureus	APC ⁴	Salmonella
Horse	400	1,000	480	n.d.	n.d.	2.3×10^4	Negative
Beef	10,000	28,000	840	10	n.d.	4.4×10^6	Negative
Pork	3,900	9,600	20	n.d.	n.d.	3.9×10^4	Presumptive Positive
Blend	110	150	4,000	n.d.	n.d.	2.6×10^7	Presumptive Positive

¹Reporting limits: E. Coli = 10 cfu/g, Total Coliforms = 10 cfu/g, Yeast = 10 cfu/g, Mold Count = 10 cfu/g, Staphylococcus aureus = 10 cfu/g, Salmonella = 1 org/25g.

 2 Cfu = colony-forming units.

 3 Org = organisms.

 $^{4}APC = Aerobic plate count.$

grams of fecal output (g/d) were greater (P = 0.01) when animals were fed Beef (15.3 g) compared to when fed the Blend diet (4.1 g). Fecal scores ranged from 1.6 to 2.6, with the Beef treatment producing higher fecal scores than Horse (P = 0.01) and Pork (P = 0.02).

All diets were highly digestible, especially regarding fat (98.6 to 99.7%), and were not different. Most statistical differences were seen between Horse, Beef, and Blend diets. The Blend diet was 7.4% more digestible (P = 0.04) in DM than Horse, 5.0% more digestible (P =0.01) in CP than Beef, 6.7% and 5.4% more digestible in OM compared to Horse (P = 0.01) and Beef (P = 0.02), respectively, and 5.6% and 4.3% more digestible in GE compared to Horse (P = 0.02) and Beef (P = 0.04), respectively. Digestibility of OM was the only difference detected between the Pork diet (93.1%) and the Blend diet (97.2%), with the Blend diet being 4.4% more digestible (P = 0.05). No other differences in nutrient digestibility were detected for Pork compared with the other diets. Using energy digestibility values from the current study, DE values for Horse, Beef, Pork, and Blend diets were 5.6, 5.9, 6.4, and 6.0 kcal/g DM, respectively. The NRC equation predicted ME of 5.2, 5.5, 6.0, and 5.5 kcal/g DM for the Horse, Beef, Pork, and Blend diets, respectively. Modified Atwater factors yielded ME predictions of 4.5, 4.7, 5.2, and 4.5 kcal/g DM, respectively, while unmodified Atwater factors yielded 4.9, 5.1, 5.6, and 4.9 kcal/g DM of ME for each respective diet (Table 4).

DISCUSSION

Reluctance to feed raw pork originated from concerns associated with Trichinae and pseudorabies (Catty, 1969; Nauwynck et al., 2007). In the US, herd biosecurity, improved farm practices, microbial interventions, inspection, vaccination programs, and freezing of raw pork products have drastically reduced these concerns if not eradicated them altogether (Hwang and Beuchat, 1995; Farkas, 1998; Rastogi et al., 2007; Greve, 2012; Lindsay et al., 2012). In the US, pseudorabies was considered eradicated in all 50 states by 2004; however, it still exists in some feral hogs and in other countries (Anderson et al., 2008). Between 2008 and 2012 only 90 cases of human Trichinae were confirmed by the Centers for Disease Control (CDC) of which only 22 were reported from pork and the remainder associated with wild game consumption (Wilson et al., 2015). Similar reductions are documented in other developed countries including Denmark and the Netherlands that consider themselves free of Trichinae (Gamble, 2001). Freezing pork at -23.3°C will kill Trichinae; therefore, pork sourced from integrated swine operations in the US, that are frozen prior to feeding, do not pose risk of these 2 concerns (Gamble, 2001). However, fresh pork not frozen prior to

feeding, or pork sourced from bio-insecure or feral sources, are of concern and should not be fed without proper testing. Additionally, the USDA has published proper handling and storage procedures for raw meat and prey fed to captive exotic animals to control microorganism growth and contamination (Crissey et al., 2001).

The objectives of this study were to determine if a commercially manufactured raw pork diet had similar apparent total tract macronutrient and energy digestibility and fecal scores as other common zoological formulations in small exotic cats, and to evaluate typical microbial population ranges in the diets. Cat ages ranged in this study, and though differences in digestibility may exist in animals of varying ages, separating age effects was not an intention of this study (Taylor et al., 1995; Teshima et al., 2010). Additionally, significant differences in nutrient digestibility between sexes are not typically observed (Wynne, 1989; Vester et al., 2008).

Prior to the study, cats received an extruded diet. Three of 4 cats transitioned from kibble to raw diets within 1 wk. One cat consumed less than 30% of daily offered diet when offered Blend and Horse treatments. While DM intake did not differ statistically, large standard error (11.2) indicated a wide range (35.3 to 50.7 g/d) across animals. Lack of consumption of Blend and Horse diets shown by one cat may have resulted from novelty of raw meat since an extruded diet was previously fed. This is characteristic of cats fed 1 diet or diet type long term who display an initial aversion to other apparently palatable foods, termed neophobia (Bradshaw, 2006). Another explanation could be preference of protein source because the cat consumed Beef and Pork diets without intake reduction. Microbe levels in the Horse diet were lower than the other diets (Table 3); therefore, bacterial contamination was an unlikely explanation.

Diet Composition

Dietary treatments were largely composed of raw meat or raw animal by-products, commercially formulated and manufactured to meet domestic cat NRC requirements and Association of American Feed Control Officials (AAFCO) recommendations for commerce in the US (National Research Council, 2006). All diets contained similar DM (28.7 to 36.9%), OM (90.4 to 94.0%), and CP (50.3 to 65.7%) concentrations as reported in previous studies analyzing raw diets (29.0 to 38.2% DM, 91.5 to 94.6% OM, and 44.9 to 64.5% CP; Crissey et al., 1997; Vester et al., 2008; Vester et al., 2010a; Vester et al., 2010b; Kerr et al., 2013a). From these previous studies, fat (22.2 to 36.9%) and GE (5.9 to 6.4 kcal/g DM) concentrations also were similar to the current study with the exception of the Pork diet that was slightly greater (39.9% and 6.7 kcal/g DM GE, respectively). When reported, TDF concentrations were similar in previous studies (range of 4.8 to 8.4%) compared to the current study with the exception of the Blend diet having the lowest TDF (1.2%) concentration which would be expected as it did not contain labeled added fiber ingredients (cellulose, beet pulp; Vester et al., 2008; Kerr et al., 2013a).

Diet Microbes

Pathogenic Salmonella and E. coli contamination are common concerns associated with raw meat diets. Although USDA published a document outlining proper handling and storage of raw meat to control microorganism growth and contamination, there are no publications outlining allowable microbial loads of raw carnivore diets for zoos (Crissey et al., 2001). Results from the current study indicated "presumptive positive" (Pork and Blend) or "negative" (Beef and Horse) for evaluation of Salmonella species. Aerobic colony counts and E. coli counts in diets were 5 and 20 times greater, respectively, than allowable levels set by the European Commission issued for human grade raw meat products (European Commission, 2005). A lack of consumption was seen with the Horse and Blend diets in one cat; as discussed previously. No other clinical symptoms (vomiting, diarrhea, lethargy, lameness) or diseases related to foodborne pathogens were observed or reported during the experiment. Fecal shedding of Salmonella was documented in up to 95% of exotic cats with no clinical symptoms or signs of illness (Clyde et al., 1997). The ability of cats to remain asymptomatic after consuming raw meat containing Salmonella suggests they tolerate high numbers of foodborne microorganisms, compared to humans (Carter and Quinn, 2000; Finley et al., 2006). Therefore, it is likely healthy carnivores are not clinically affected by presence of pathogens such as Salmonella and standards set for human grade meat products may not be appropriate.

Microbial levels in carnivore diets may pose potential risks to humans through preparation and handling of raw meat diets. Animal managers should adhere to USDA published standard operating procedures for handling raw meat to reduce risks (Crissey et al., 2001). Additionally, the CDC advises the immediate washing of hands after contacting animal fecal material to reduce the risk and contraction of Salmonellosis (Center for Disease Control and Prevention, 2015).

In the current study, only general microbes typically evaluated in quality control programs in zoos were considered to demonstrate the expected range of properly handled raw meat diets. It is critical to understand that "presumptive positive" results for *Salmonella* indicates presence of any of 2,400 serotypes of *Salmonella* species, only a handful of which are pathogenic (Carlson et al., 2012). Similarly, *E. coli* counts include all species, totaling more than 50,000 serotypes, of which few are pathogenic (Ørskov and Ørskov, 1992). For this reason, high levels do not inherently infer a diet is dangerous to a carnivore. Serotyping samples would be necessary to evaluate specific strains of concern. Bacteria such as *Listeria monocytogenes* and *Campylobacter jejuni* have not been critically evaluated in zoo raw diets and likely warrant research (Eisel et al., 1997). If clinical signs are observed in an animal consuming a raw meat diet, it may be necessary to evaluate the diet for microbes beyond general screens typically used.

Energy and Macronutrient Digestibilities

Digestibility of nutrients is simply a measure of how efficiently a diet is digested by the animal. In animal production systems, higher digestibility typically results in less feed and reduced costs of production. In pets or exotic animals managed in zoos, higher diet digestibility may not be necessary but could reduce feeding costs. Goals of animal feeding programs vary across institutions; therefore, digestibility measures can be considered in tandem with feeding goals. All diets evaluated were highly digestible across nutrients, regardless of protein source. Values reported in the current study were similar to those previously reported in studies analyzing raw meat diets (horse or beef). Previous studies reported similar ranges of digestibility for OM (87.4 to 96.5%), CP (91.7 to 97.2%), fat (90.5 to 99.0%), and GE (89.6 to 97.3%; Crissey et al., 1997; Vester et al., 2008; Vester et al., 2010a; Vester et al., 2010b; Kerr et al., 2013a; Iske et al., 2016). However, DM digestibility values (83.6 to 90.0%) reported in the previous studies were slightly lower than those observed in the present study (87.6 to 94.1%). These ranges include data from our previous work feeding similar diets in large exotic cats (Iske et al., 2016). Macronutrient digestibility values of Pork were similar in large cats with 88.0, 90.8, 95.7, 98.5, and 92.4% digestibility for DM, OM, CP, fat, and GE, respectively (Iske et al., 2016). These values are within 3% of those observed in the current study.

Crude protein digestibility of Beef (93.4%) was lower than Horse (96.8%) and Blend (98.1%) diets. Protein digestibility could have been affected by protein content and dietary fiber (Clauss et al., 2010). Kienzle et al. (1991) demonstrated an 18.9% reduction in protein digestibility of a raw meat diet fed to domestic cats when 15% horn meal was added as a fiber source. Protein digestibility differences may be more profound when beet pulp is included compared with cellulose. In a study evaluating raw meat diets containing 9.5% TDF from various fiber sources, fed to 30 domestic cats, protein digestibility was approximately 5.8% lower when beet pulp was used compared with cellulose (Sunvold et al., 1995). High fermentative characteristics of beet pulp (compared to cellulose) in the large intestine consequentially increase bacterial protein, leading to more protein excretion and underestimation of apparent total tract protein digestibility (Kerr et al., 2013b). This is supported in the current study as the Beef diet contained beet pulp and may have contributed to lower observed protein digestibility.

Beyond fiber source, differences in protein digestibility could be attributed to protein ingredients and processing. The Beef diet also contained fish meal and soybean meal as protein sources whereas other treatment diets contained only raw muscle meat or raw animal meat by-products. Plant-based protein sources may contain as much as 18.8% NFE which cats are less efficient at digesting compared to protein (McDonald, 2002; Karr-Lilienthal et al., 2004). Heat processing to produce meal ingredients may cause Maillard reactions that could reduce protein digestion (Camire et al., 1990). Maillard reactions and cross-linkages of amino acids, may reduce amino acid retention and availability as well as protein digestibility and quality (Björck et al., 1983; Björck et al., 1984).

In the US, commercially manufactured companion and exotic animal feeds for interstate commerce are regulated at state levels by the AAFCO, which suggests modified Atwater values for product labeling of ME (Association of American Feed Control Officials, 2014). Other methods of predicting ME include unmodified Atwater values and the NRC equation that uses digestible energy and protein content (National Research Council, 2006). Modified Atwater values reflect 85.0, 95.0, and 80.0% digestibility for carbohydrate, fat, and protein, respectively, based on standard kibble diets (Kienzle, 2002). In contrast, unmodified Atwater factors reflect 98.0, 98.0, and 90.0% digestibility for carbohydrate, fat, and protein, respectively which is more representative of digestibility values obtained from raw diets (Kienzle, 2002).

Differences between DE and ME of diets were 25.1, 17.5, and 8.5% using modified and unmodified Atwater factors, and the NRC equation, respectively. Urine energy losses in domestic cats were 6.5% of energy intake when fed a high protein (61.5%) canned diet (mixed with beef heart) and 4.6% when fed a high fat diet (49.1% fat, 39.9% protein; Riond et al., 2003). This, along with negligible gas loss because of limited fermentation in cats, suggests differences between DE and ME should be around 6.5%. The current study is supported by previous studies demonstrating modified Atwater values underestimate ME for raw meat diets. Animal managers should evaluate the ME content on labels of diets produced in the US to manage body weight and condition more accurately (Clauss et al., 2010). Although AAFCO suggests modified Atwater

values to predict ME for commercial labeling, unmodified Atwater values or NRC methodology better predicts ME of raw meat diets and should be considered for labeling of raw meat diets.

Fecal scores of cats consuming Beef(2.6) were greater than scores from cats consuming Horse (1.6) and Pork (1.9). Fiber sources varied among the treatment diets and likely contributed to differences observed in fecal scores. Differences may exist among exotic cat size as smaller species including cheetah appear to tolerate fermentable beet pulp (4% of diet) while larger species, such as tigers appear to have more ideal fecal scores when fed cellulose (4% of the diet; Kerr et al., 2013b). Domestic cats also show more ideal fecal scores with beet pulp inclusion, opposed to cellulose (Sunvold et al., 1995). The Beef diet in the current study contained beet pulp and had a TDF concentration of 5.5%, resulting in fecal scores closest to what is considered ideal (3.0). Cellulose, a non-fermentable fiber, provides the bowel with tactile stimulation inducing colonic motility and weight while fermentable fibers such as beet pulp induce chemical changes and production of short chain fatty acids (SCFA), that can be absorbed by colonocytes for energy and aid it colon health (Bueno et al., 2000a; Bueno et al., 2000b). Too much fermentable fiber, likely above 4% of diet for cats, may result in excess production of SCFA, increasing passive transport absorption and possibly resulting in soft stools (Kerr et al., 2013b). It has been suggested that raw diets for zoo carnivores should include a combination of beet pulp and cellulose (2% each) to achieve optimal intestinal health (Kerr et al., 2013b).

Conclusions

Similar macronutrient and energy digestion was observed in African wildcats fed a pork-based raw meat diet compared with other common diets varying in protein and fiber source. Additionally, cats tolerated high and variable ranges of microbial loads in raw meat diets without impact on fecal scores. Beyond lack of consumption, clinical signs of infection and disease were absent in cats fed diets and nutrient digestibilities fell within expected ranges despite the microbe variation, indicating guidelines for appropriate and expected levels of microbes in carnivore diets need further evaluation and clarification to aid animal managers and diet manufacturers alike. Additionally, because raw meat diets are digested more efficiently than processed foods, predictions of ME for labeling of commercial products should utilize either the NRC equation or unmodified Atwater values as AAFCO suggested modified Atwater values will underestimate ME and could contribute to animal obesity. In conclusion, if properly sourced and handled, pork can serve as a protein option in raw meat diets for zoo-managed carnivores.

LITERATURE CITED

- Anderson, L. A., N. Black, T. J. Hagerty, J. P. Kluge, L. Paul and P. L. Sundberg. 2008. Pseudorabies (Aujeszky's disease) and its eradication: A review of the U.S. experience. Technical Bulletin No. 1923. United Stated Department of Agriculture, Washington, D.C.
- AOAC. 1994. Coliforms and *E. coli* counts in foods. Dry rehydratable film method, method 991.14. Off. methods Anal. AOAC Int., 17th ed. Arlington, VA.
- AOAC. 1995. Official Method 991.36, Fat (crude) in meat, solvent extraction (Submersion) method. Off. Methods Anal. Arlington, VA.
- AOAC. 2003. Staphylococcus aureus in selected types of processed and prepared foods, method 2003.07. Off. Methods Anal. Arlington, VA.
- AOAC. 2006. Official methods of analysis. 17th ed. Assoc. Off. Anal. Chem, Arlington, VA.
- Association of American Feed Control Officials. 2014. Model regulations for pet food and specialty pet food under the model bill. Association of American Feed Control Officials, Oxford, UK. p. 136–164.
- Björck, I., N. G. Asp, and A. Dahlqvist. 1984. Protein nutritional value of extrusion-cooked wheat flours. Food Chem. 15:203– 214. doi:10.1016/0308-8146(84)90004-9
- Björck, I., A. Noguchi, N. G. Asp, J. Cheftel, and A. Dahlqvist. 1983. Protein nutritional value of a biscuit processed by extrusion cooking: Effects on available lysine. Food Chem. 31:488–492. doi:10.1021/jf00117a006
- Bradshaw, J. W. S. 2006. The evolutionary basis for the feeding behavior of domestic dogs (*Canis familiaris*) and cats (*Felis catus*). J. Nutr. 136:1–3.
- Bueno, A. R., T. G. Cappel, G. D. Sunvold, R. A. Moxley, G. A. Reinhart, and E. T. Clemens. 2000a. Feline colonic microbes and fatty acid transport: Effects of feeding cellulose, beet pulp and pectin/gum arabic fibers. Nutr. Res. 20:1319–1328. doi:10.1016/S0271-5317(00)00211-6
- Bueno, A. R., T.G. Cappel, G. D. Sunvold, G. A. Reinhart, and E. T. Clemens. 2000b. Feline colonic morphology and mucosal tissue energetics as influenced via the source of dietary fiber. Nutr. Res. 20:985–993. doi:10.1016/S0271-5317(00)00189-5
- Camire, M. E., A. Camire, and K. Krumhar. 1990. Chemical and nutritional changes in foods during extrusion. Crit. Rev. Food Sci. Nutr. 29:35–57. doi:10.1080/10408399009527513
- Carlson, S., A. Barnhill, and R. Griffith. 2012. Salmonellosis. In: J. Zimmerman, L. Karriker, A. Ramirez, K. Schwartz, and G. Stevenson, editors, Diseases of swine. 10th ed. John Wiley & Sons, West Sussex, UK. p. 821–831.
- Carter, M., and P. Quinn. 2000. Salmonella infections in dogs and cats. In: C. Wray and A. Wray, editors, Salmonella in domestic animals. CABI, Oxon, UK. p. 231–244. doi:10.1079/9780851992617.0231
- Catty, D. 1969. The immunology of nematode infections. Trichinosis in guinea-pigs as a model. Monogr. Allergy 5:1–40.
- Center for Disease Control and Prevention. 2015. Salmonella: More about prevention. https://www.cdc.gov/salmonella/ general/prevention/html. (Accessed 8 December 2016.)
- Cho, S., C. Clark, and S. Rickard. 2001. Dietary fiber and breast cancer risk. In: S. Cho and M. Dreher, editors, Handbook of dietary fiber. Marcel Dekker, Inc., New York. p. 47–88. doi:10.1201/9780203904220
- Clauss, M., H. Kleffner, and E. Kienzle. 2010. Carnivorous mammals: Nutrient digestibility and energy evaluation. Zoo Biol. 29:687–704. doi:10.1002/zoo.20302

- Clyde, V. L., E. C. Ramsay, and D. A. Bemis. 1997. Fecal shedding of salmonella in exotic felids. J. Zoo Wildl. Med. 28:148–152.
- Crissey, S. D., K. A. Slifka, P. Shumway, and S. B. Spencer. 2001. Handling frozen/thawed meat and prey items fed to captive exotic animals: A manual of standard operating procedures. United States Dep. Agric. Beltsville, MD.
- Crissey, S. D., J. A. Swanson, B. A. Lintzenich, B. A. Brewer, and K. A. Slifka. 1997. Use of a raw meat-based diet or a dry kibble diet for sand cats (*Felis margarita*). J. Anim. Sci. 75:2154–2160. doi:10.2527/1997.7582154x
- de-Oliveira, L. D., F. S. Takakura, E. Kienzle, M. A. Brunetto, E. Teshima, G. T. Pereira, R. S. Vasconcellos, and A. C. Carciofi. 2012. Fibre analysis and fibre digestibility in pet foods- a comparison of total dietary fibre, neutral and acid detergent fibre and crude fibre. J. Anim. Physiol. Anim. Nutr. 96:895–906. doi:10.1111/j.1439-0396.2011.01203.x
- Eisel, W., R. Linton, and P. Muriana. 1997. A survey of microbial levels for incoming raw beef, environmental sources, and ground beef in a red meat processing plant. Food Microbiol. 14:273–282. doi:10.1006/fmic.1996.0094
- European Commission. 2005. Commission Regulation (EC) No 2073/2005 of 15 November 2005 on microbiological criteria for foodstuffs. Off. J. Eur. Union. L 338:1–26.
- Farkas, J. 1998. Irradiation as a method for decontaminating food: A review. Food Rev. Int. 4:189–204. doi:10.1080/87559128809540822
- Felid Taxon Advisory Group. 2014. Basic fecal scale- felids.Felid Taxon Advisory Group, AZA, Silver Spring, MD.
- Finley, R., R. Reid-Smith, J. S. Weese, and F. J. Angulo. 2006. Human health implications of *salmonella*-contaminated natural pet treats and raw pet food. Clin. Infect. Dis. 42:686–691. doi:10.1086/500211
- Gamble, H.R. 2001. Trichinae: Pork facts—food quality and safety. Beltsville, MD: US Department of Agriculture. Agric. Res. Serv., Parasite Biol. Epidemiol. Lab. https://www.aphis.usda.gov/vs/trichinae/docs/fact_sheet.htm (Accessed 8 December 2016.)
- Greve, J. 2012. Internal Parasites: Helminths. In: J. Zimmerman, L. Karriker, A. Ramirez, K. Schwartz, and G. Stevenson, editors, Diseases of swine. 10th ed. John Wiley & Sons, West Sussex, UK. p. 912–913.
- Hwang, C. A., and L. R. Beuchat. 1995. Efficacy of a lactic-acid sodium benzoate wash solution in reducing bacterial-contamination of raw chicken. Int. J. Food Microbiol. 27:91–98. doi:10.1016/0168-1605(94)00150-5
- Iske, C. J., C. L. Morris, and K. L. Kappen. 2016. Influence of pork and pork by-products on macronutrient and energy digestibility and palatability in large exotic felids. J. Anim. Sci. 94:3738–3745. doi:10.2527/jas.2016-0414
- Karr-Lilienthal, L. K., C. M. Grieshop, N. R. Merchen, D. C. Mahan, and G. C. Fahey. 2004. Chemical composition and protein quality comparisons of soybeans and soybean meals from five leading soybean-producing countries. J. Agric. Food Chem. 52:6193–6199. doi:10.1021/jf049795+
- Kendall, P. T., D. W. Holme, and P. M. Smith. 1982. Comparative evaluation of net digestive and absorptive efficiency in dogs and cats fed a variety of contrasting diet types. J. Small Anim. Pract. 23:577–587. doi:10.1111/j.1748-5827.1982.tb02518.x
- Kerr, K. R., A. N. Beloshapka, C. L. Morris, C. Parsons, S. L. Burke, P. Utterback, and K. S. Swanson. 2013a. Evaluation of four raw meat diets using domestic cats, captive exotic felids, and cecectomized roosters. J. Anim. Sci. 91:225–237. doi:10.2527/jas.2011-4835

- Kerr, K. R., C. L. Morris, S. L. Burke, and K. S. Swanson. 2013b. Influence of dietary fiber type and amount on energy and nutrient digestibility, fecal characteristics, and fecal fermentative end-product concentrations in captive exotic felids fed a raw beef-based diet. J. Anim. Sci. 91:2199–2210. doi:10.2527/jas.2012-5702
- Kerr, K. R. 2012. Nutritional evaluation of raw meat and whole prey diets for domestic and exotic cats. PhD Diss. University of Illinois at Urbana-Champaign, Champaign, IL.
- Kerr, K. R., B. M. Vester Boler, C. L. Morris, K. J. Liu, and K. S. Swanson. 2012. Apparent total tract energy and macronutrient digestibility and fecal fermentative end-product concentrations of domestic cats fed extruded, raw beef-based, and cooked beefbased diets. J. Anim. Sci. 90:515–522. doi:10.2527/jas.2010-3266
- Kienzle, E. 2002. Further developments in the prediction of metabolizable energy (ME) in pet food. J. Nutr. 132:1796S–1798S.
- Kienzle, E., H. Meyer, and R. Schneider. 1991. Investigations on palatability, digestibility and tolerance of low digestible food components in cats. J. Nutr. 121:S56–S57.
- Lindsay, D., J. Dubey, M. Santin-Duran, and R. Fayer. 2012. Coccidia and other protozoa. In: J. Zimmerman, L. Karriker, A. Ramirez, K. Schwartz, and G. Stevenson, editors, Diseases of swine. 10th ed. John Wiley & Sons, West Sussex, UK. p. 899–901.
- Maturin, L., and Peeler, J. T. 2001. Aerobic plate count. In: Bacteriological Analytical Manual. Food and Drug Administration, Silver Spring, MD.
- McDonald, P. 2002. Digestion in the dog and cat: Digestion. In: R. Edwards, J. Greenhalgh, C. Morgan, L. Sinclair, and R. Wilkinson, editors, Animal nutrition. Pearson Education, Harlow, UK. p. 166–190.
- National Research Council. 2006. Nutrient requirements of dogs and cats. National Academy Press, Washington, DC.
- Nauwynck, H., S. Glorieux, H. Favoreel, and M. Pensaert. 2007. Cell biological and molecular characteristics of pseudorabies virus infections in cell cultures and in pigs with emphasis on the respiratory tract. Vet. Res. 38:229–241. doi:10.1051/vetres:200661
- Ørskov, F., and I. Ørskov. 1992. *Escherichia coli* serotyping and disease in man and animals. Can. J. Microbiol. 38:699–704. doi:10.1139/m92-115
- Prosky, L., N. Asp, T. Schweizer, J. DeVries, I. Furda, and S. Lee. 1994. Determination of soluble dietary fiber in foods and food products: Collaborative study. J. AOAC Int. 77:690–694.
- Rastogi, N. K., K. S. M. S. Raghavarao, V. M. Balasubramaniam, K. Niranjan, and D. Knorr. 2007. Opportunities and challenges in high pressure processing of foods. Crit. Rev. Food Sci. Nutr. 47:69–112. doi:10.1080/10408390600626420

- Riond, J.-L., M. Stiefel, C. Wenk, and M. Wanner. 2003. Nutrition studies on protein and energy in domestic cats. J. Anim. Physiol. Anim. Nutr. 87:221–228. doi:10.1046/j.1439-0396.2003.00431.x
- Robinson, M. H. 1998. Enriching the lives of zoo animals, and their welfare: Where research can be fundamental. Anim. Welf. 7:151–175.
- Sunvold, G. D., G. C. Fahey, N. R. Merchen, L. D. Bourquin, E. C. Titgemeyer, L. L. Bauer, and G. A. Reinhart. 1995. Dietary fiber for cats: In vitro fermentation of selected fiber sources by cat fecal inoculum and in vivo utilization of diets containing selected fiber sources and their blends. J. Anim. Sci. 73:2329–2339. doi:10.2527/1995.7382329x
- Taylor, E. J., C. Adams, and R. Neville. 1995. Some nutritional aspects of ageing in dogs and cats. Proc. Nutr. Soc. 54:645–656. doi:10.1079/PNS19950064
- Teshima, E., M. A. Brunetto, R. S. Vasconcellos, K. N. V. Gonçalves, L. D. De-Oliveira, A. G. Valério, and A. C. Carciofi. 2010. Nutrient digestibility, but not mineral absorption, is age-dependent in cats. J. Anim. Physiol. Anim. Nutr. 94. doi:10.1111/j.1439-0396.2009.00964.x
- Thiex, N. 2008. Evaluation of analytical methods for the determination of moisture, crude protein, crude fat, and crude fiber in distillers dried grains with solubles. J. AOAC Int. 92:61–73.
- Tournas, V., M. E. Stack, P. B. Mislivec, H. A. Koch, and R. Bandler. 2001. Yeasts, molds and mycotoxins. In: Bacteriological Analytical Manual. Food and Drug Administration, Silver Spring, MD.
- Vester, B. M., A. N. Beloshapka, I. S. Middelbos, S. L. Burke, C. L. Dikeman, L. G. Simmons, and K. S. Swanson. 2010a. Evaluation of nutrient digestibility and fecal characteristics of exotic felids fed horse-or beef-based diets: Use of the domestic cat as a model for exotic felids. Zoo Biol. 29:432–448. doi:10.1002/zoo.20275
- Vester, B. M., S. L. Burke, C. L. Dikeman, L. G. Simmons, and K. S. Swanson. 2008. Nutrient digestibility and fecal characteristics are different among captive exotic felids fed a beef-based raw diet. Zoo Biol. 27:126–136. doi:10.1002/zoo.20172
- Vester, B. M., S. L. Burke, K. J. Liu, C. L. Dikeman, L. G. Simmons, and K. S. Swanson. 2010b. Influence of feeding raw or extruded feline diets on nutrient digestibility and nitrogen metabolism of African wildcats (*Felis lybica*). Zoo Biol. 29:676–686. doi:10.1002/zoo.20305
- Wilson, N. O., R. L. Hall, S. P. Montgomery, and J. L. Jones. 2015. Trichinellosis Surveill.— United States, 2008–2012. MMWR Surviellance Summ. 64:1–8.
- Wynne, J. E. 1989. Comparative digestibility values in four species of felidae. ZJ. Zoo Wildlife Med. 20:53–56.