Bioorthogonal non-canonical amino acid tagging - BONCAT -

Hatzenpichler lab www.environmental-microbiology.com doi: 10.13140/RG.2.1.3698.7040/1

BONCAT in environmental microbiology (as of February 2020)

Hatzenpichler R et al. *In situ* visualization of newly synthesized proteins in environmental microbes using amino acid tagging and click chemistry

Environ Microbiol, 16: 2568-2590 (2014)

First application of BONCAT to uncultured microbes; development of BONCAT-FISH; correlation of BONCAT with nanoSIMS

Samo TJ et al. Broad distribution and high proportion of protein synthesis active marine bacteria revealed by click chemistry at the single cell level

Front Aquat Microbiol, 1: 48 (2014)

application of BONCAT to seawater; correlation of BONCAT with microautoradiography

Hatzenpichler R and Orphan VJ Detection of protein-synthesizing microorganisms in the environment via bioorthogonal noncanonical amino acid tagging (BONCAT)

Book chapter for Hydrocarbon and Lipid Microbiology Protocols, Springer Protocols Handbooks, doi 10.1007/8623_2015_61 (2015)

description of how to design and protocols for how to perform BONCAT-experiments using AHA and HPG

Hatzenpichler R et al. Visualizing *in situ* translational activity for identifying and sorting slow-growing archaeal-bacterial consortia Proc Natl Acad Sci USA, 113: E4069-E4078 (2016)

development of activity-based cell-sorting via bioorthogonal labeling (BONCAT-FACS); applied BONCAT-(CARD)FISH and BONCAT-FACS to deep-sea sediment consortia catalyzing the anaerobic oxidation of methane with sulfate

Leizeaga et al. Using Click-Chemistry for Visualizing *in Situ* Changes of Translational Activity in Planktonic Marine Bacteria Front Aquat Microbiol, 8: 2360 (2017)

Pasulka AL et al. Interrogating marine virus-host interactions and elemental transfer with BONCAT and nanoSIMS-based methods Environ Microbiol, 20: 671-692 (2018)

First application of BONCAT to environmental phages and viruses; estimate of marine viral production rates by BONCAT and nanoSIMS

Couradeau et al. Probing the active fraction of soil microbiomes using BONCAT-FACS

Nat Comm, 10: 2770 (2019)

> first application of BONCAT to soil samples; reports that a surprisingly high proportion of soil microbes is translationally active

Sebastian et al. High Growth Potential of Long-Term Starved Deep Ocean Opportunistic Heterotrophic Bacteria

Front Aquat Microbiol, 10: 760 (2019)

Steward et al. Metabolic Implications of Using BioOrthogonal Non-Canonical Amino Acid Tagging (BONCAT) for Tracking Protein

Synthesis

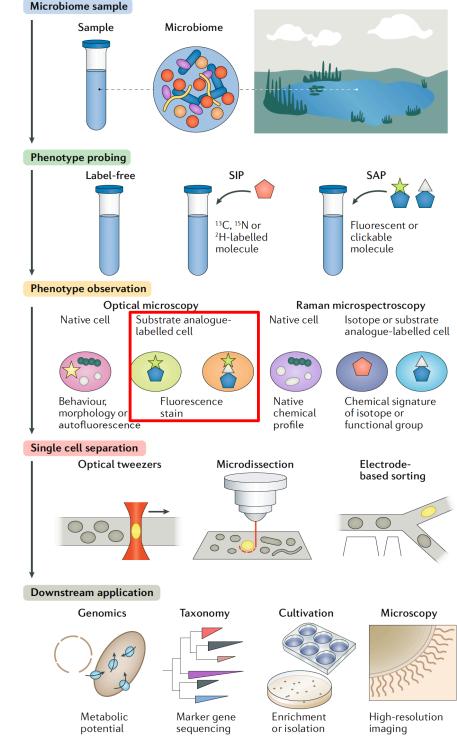
Front Micriol, 11:197 (2020)

BONCAT is a <u>Next-generation</u> physiology approach

Definition: ...any combination of techniques that analyze the phenotype of an individual cell in a microbiome in a non-destructive way, which enables the physical separation of this cell based solely on its phenotype for subsequent, downstream applications

Specifically, BONCAT is a type of substrate analog probing (SAP)

SAP uses molecules that carry either a fluorescence tag or a side group amenable to azide-alkyne click chemistry to obtain information on the overall biosynthetic activity or specific enzymatic function of the cell.



Hatzenpichler et al., 2020

Fig. 2 | Next-generation physiology workflow to study microorganisms.

First, some definitions

bioorthogonal

non-interacting with cellular functionalities

non-canonical

synthetic, not part of biological machinery

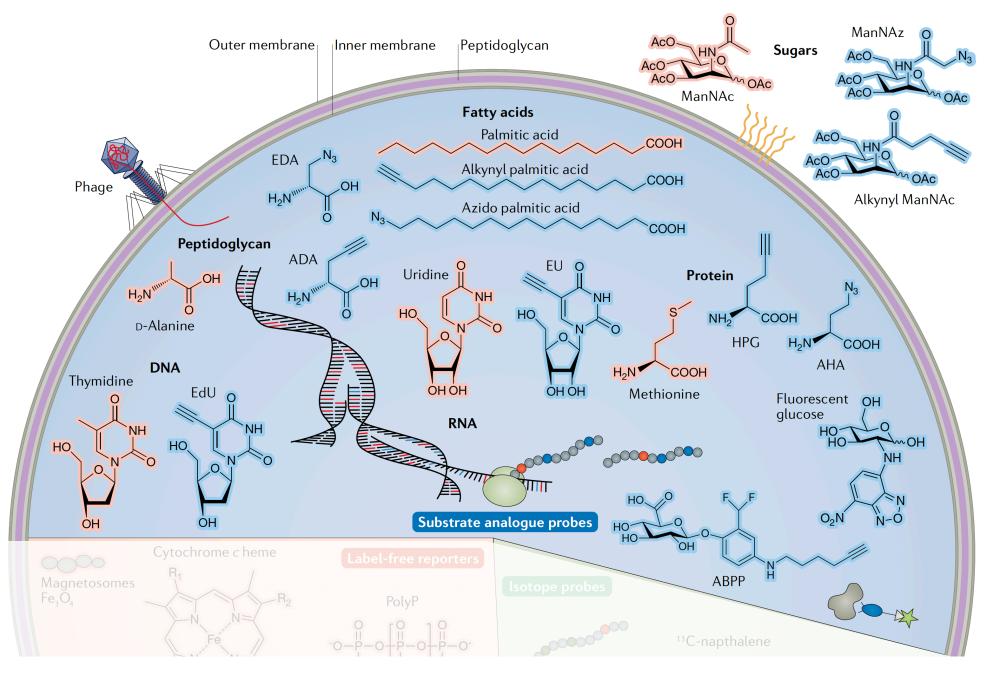
Click chemistry

complete conversion of reagents to single product

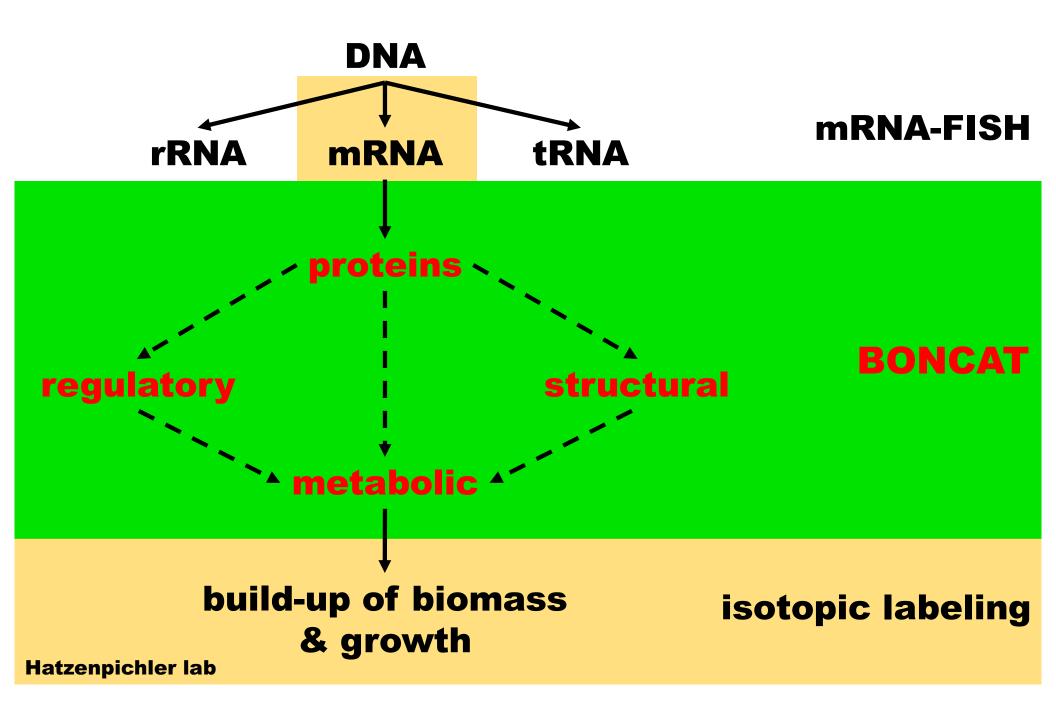
+ mild conditions

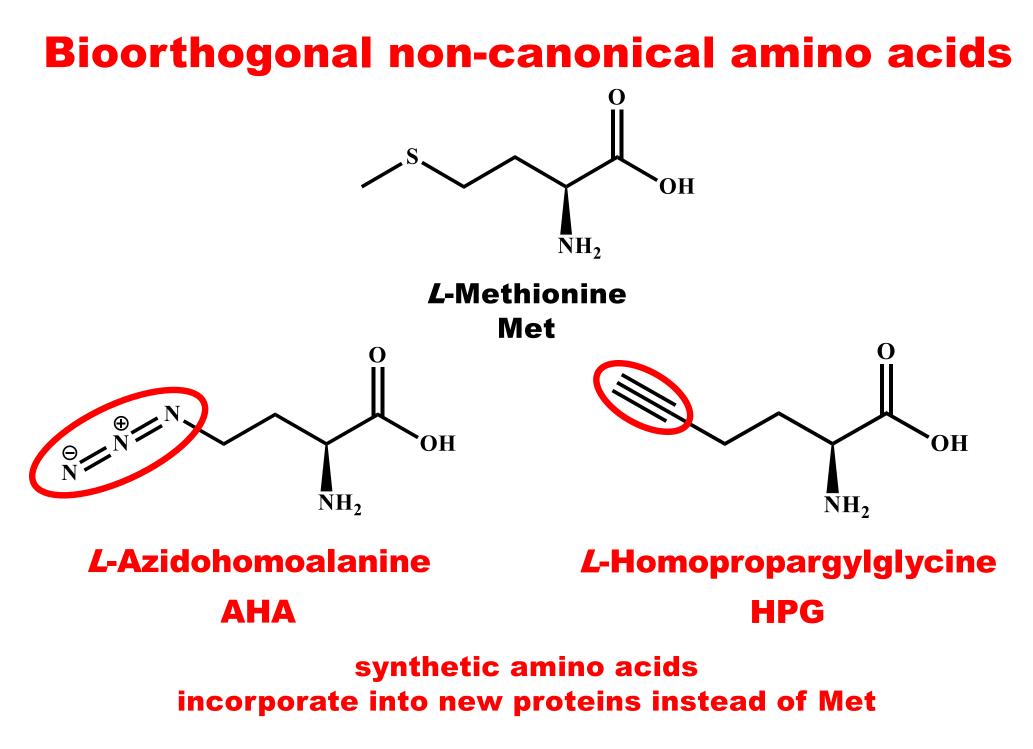
- + very fast
- + in water

Examples for clickable substrate analogs

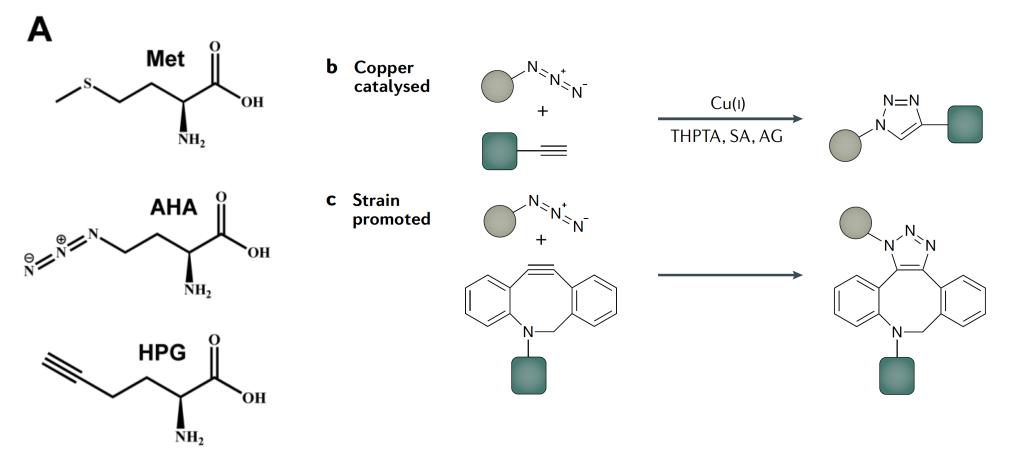


Activity assays on individual cell level





Azide-alkyne click chemistry reactions



A. Structures of Met and its surrogates AHA and HPG, which compete with Met during translation.

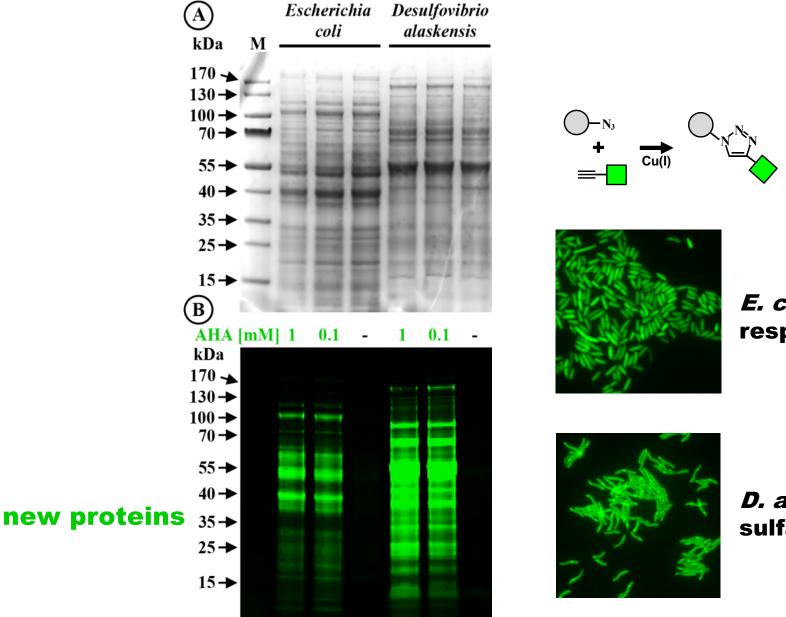
B. In Cu(I)-catalyzed click chemistry an azide group (N_3) is linked to a terminal alkyne residue, yielding a triazole conjugate.

C. Strain-promoted click chemistry allows the copper-less conjugation of an azide group (N_3) with a cyclo-octyne-carrying molecule, yielding a triazole conjugate.

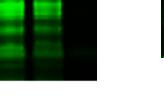
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Hatzenpichler *et al.*, 2015 Hatzenpichler *et al.*, 2020

Incorporation into newly made proteins



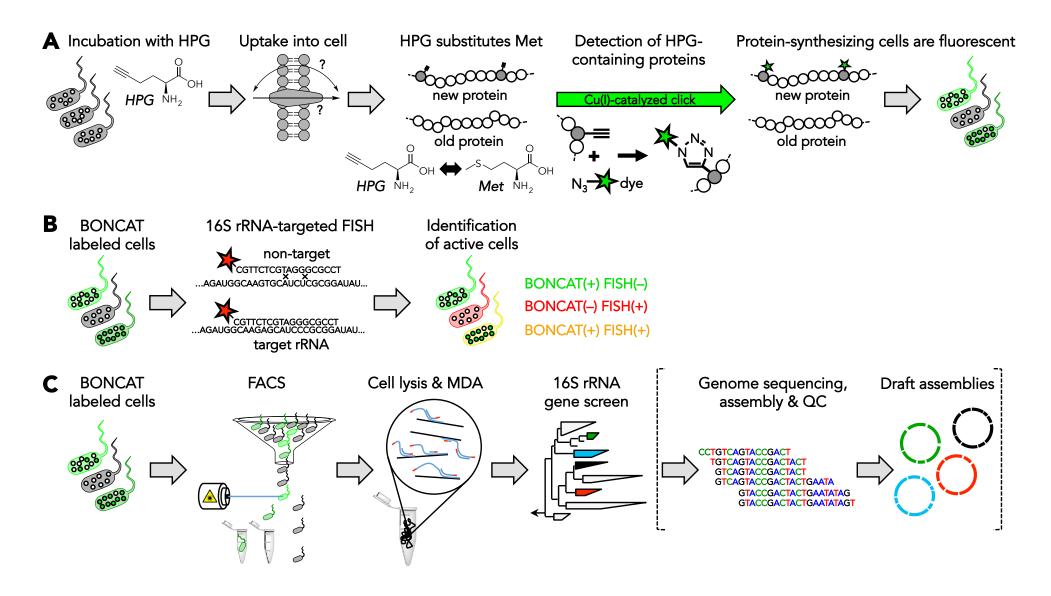
E. coli respiring glucose



D, alaskensis sulfate reducer

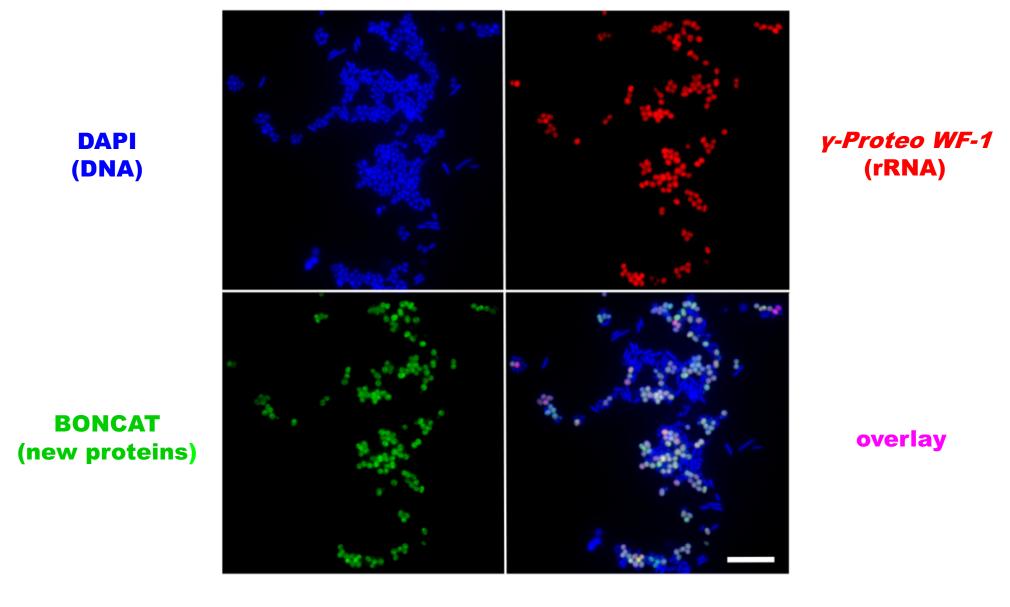
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Visualizing, identifying, and sorting translationally active microbes



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Identification of translationally active cells



Bar = 10 µm

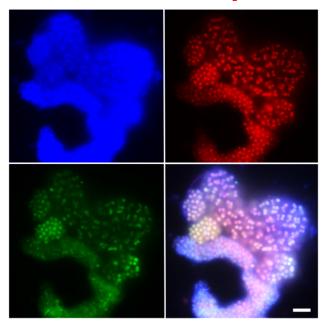
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BONCAT-FISH of uncultured microbes

Arch915

EUB338 I-III

Gam42a + competitor



Methane seep ANME-SRB consortium

Tongue biofilm and saliva

Freshwater from Lily pond on Caltech campus



Hatzenpichler et al., 2014; Hatzenpichler et al., 2015; Hatzenpichler et al., 2016

Visualizing new proteins in situ

generally applicable (works for all taxonomies and physiologies tested so far)

detectable after 2% of generation time

FISH-BONCAT links function and identity of a cell

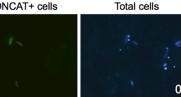
BONCAT correlates with ¹⁵NH₃ incorporation (nanoSIMS)

no change in protein expression (Bagert *et al*., 2014)

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Research examples

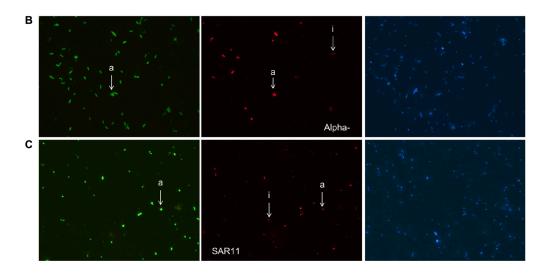
Detecting activity of environmentally relevant bacteria in ocean water

5h

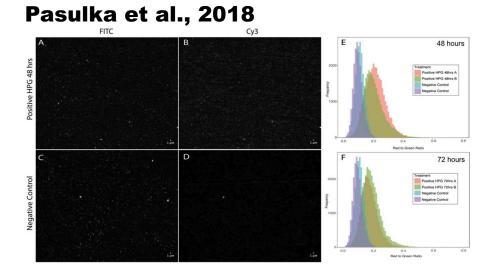
Watching cells resuscitate after long-term starvation

Sebastian et al., 2019

Leizeaga et al., 2017



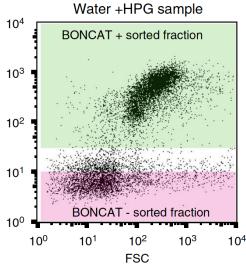
Studying virus turnover in bacterioplankton



22h

29h

10⁴ Sorting and FAM picolyl azide dye fluorescence identifying the 10³ active fraction o cells in soil with 10² **BONCAT-FACS** Couradeau et al. 10¹ 2019 10^{0}



Limitations and advantages of BONCAT-FISH

uptake and incorporation

Methionine-rich samples are tough

hard to quantitate amount of new proteins in uncultured cells

potential for cell inactivation or community shifts

links cellular identity and function

fluorescence-based in situ activity studies

metabolic screening

activity-based cell-sorting

fast + highly selective + cheap + easily available 1 h azide-alkyne ~\$500 epi-scope