

## Genotoxic Effects Of Organophosphate Pesticide Phorate In Some Exotic Fishes Of Kashmir

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**Abstract:** Genotoxic effects of phorate, a commonly used pesticide were evaluated in two exotic sub-species of fish, *Cyprinus carpio* L. (family *Cyprinidae*) namely *Cyprinus carpio specularis* and *Cyprinus carpio communis* using micronucleus test. Genotoxicity of said pesticide was confirmed by incidence of micronucleus in peripheral erythrocytes using three sub-lethal concentrations viz 0.2ppm, 0.4ppm and 0.6ppm of phorate after 24, 48 and 72 hours. All the three concentrations were able to induce micronuclei formation in erythrocytes of both fish species. However, after 48h and 72h, a statistically significant increase was found in the frequency of micronuclei in peripheral erythrocytes of both fish species. The percentage of single micronuclei in *Cyprinus carpio specularis* ( $0.03 \pm 0.01$  in control) increased to  $1.15 \pm 0.32$  from low to high concentrations after 24h and  $2.74 \pm 0.52$  in longer exposures. In *Cyprinus carpio communis* somewhat similar results were observed with increase in percentage of single micronuclei ( $0.03 \pm 0.01$  in control) to  $1.30 \pm 0.23$  at 24h from low to high concentration and this percentage continued to increase by  $2.08 \pm 0.31$  and  $2.91 \pm 0.39$  after 48 and 72 h respectively (Mann-Whitney U test;  $p < 0.05$ ). [Maraj-ud-din Malik, Zeenat Nisar. **Genotoxic Effects Of Organophosphate Pesticide Phorate In Some Exotic Fishes Of Kashmir.** *Biomedicine and Nursing* 2015;1(1):13-17]. <http://www.nbmedicine.org>. 3

**Key words:** Micronucleus; *Cyprinus*; Phorate; Genotoxicity; Pesticide.

### Introduction

Phorate is an organophosphate pesticide effective against a wide array of insects, mites and some nematodes. It is used on a variety of field crops but 80% of its use is on corn, potatoes and cotton. Other crops include beans, peanuts, sugar beets, sorghum, wheat and soybean. In Kashmir phorate is used on a variety of crops especially it is used to control woolly apple aphid.

Phorate is very highly toxic to birds and other wild life. It has been responsible for numerous incidents of mortality in birds and fish. Fish provide a relevant model for the evaluation of aquatic genotoxicity *in situ*, as well as the action of polluted effluents, sediments or toxic compounds (Hayashi *et al.*, 1998). The advantage of fish as model organism include the fact that fish respond in a manner similar to mammalian test species to chemicals that induce peroxisome proliferation in hepatocytes and oxidative damage in hepatocytes. The advantages of using fish as model organisms include the ease with which the fish, especially aquarium species, can be held in laboratory and exposed to toxic chemicals. Since fish often respond to toxicants in a manner similar to higher vertebrates, they can be used to screen for chemicals that have the potential to cause teratogenic and carcinogenic effects in humans. With the last decade, the use of fish as appropriate models for genetic monitoring of toxic chemicals in aquatic environments has become popular (Pacheco and Santos, 1998).

A variety of *in vitro* and *in vivo* assays with fish are being used as a model system for toxicological, biochemical and developmental studies (Powers,

1989). At the cellular level the micronucleus test on various fish tissues is among the most wide spread assessments of genotoxicity in water (Al-Sabti K and Hardig J, 1990). Various investigations using fish as sentile for screening the clastogenic effects of xenobiotics indicate that these fish represent good experimental models for genotoxicity studies (Matsumoto and Colus, 2000; Porto *et al.*, 2005, Pantaleao *et al.*, 2006).

The aim of the present study is to investigate the genotoxic effects of a known organophosphate pesticide, phorate on two sub-species of *Cyprinus carpio* fish, *Cyprinus carpio communis* (scale carp) and *Cyprinus carpio specularis* (mirror carp), phenotypically differentiable according to the pattern of scales, when exposed to different concentrations of the said organophosphate pesticide, using micronucleus test.

Micronucleus test can be performed on different cell types like lymphocytes, erythrocytes, fibroblasts, and exfoliated epithelial cells, without extra *in vitro* cultivation step. The micronucleus assay is generally accepted as equivalent to the cytogenetic assay in responsiveness to chemical mutagens. As scoring is less consuming, the assay is preferred for routine screening purposes. Another advantage of the micronucleus assay is that micronucleated erythrocytes persist in the peripheral blood for a relatively longer time, therefore allowing the study of chronic exposure to potential mutagens (Choy *et al.*, 1985).

Micronucleus assay originally developed with mammalian species, has been extensively used to test for genotoxic activity of chemicals (Heddle *et al.*,

1983). The micronucleus test in fish has a potential for detecting clastogenic activity. Various studies have shown that the peripheral erythrocytes of fish have a high incidence of micronuclei after exposure to different pollutants under field and laboratory conditions.

## Materials And Methods

### Experimental animals

Two sub-species of *Cyprinus carpio* L. (family *Cyprinidae*) namely *Cyprinus carpio specularis* and *Cyprinus carpio communis* were used as experimental animals as they are available throughout the year in Kashmir. Adult specimens of both sub-species were collected from Dal-lake and were identified on the basis of their scales. Fish were acclimatized for 45 days at 28°C prior to trials (Anitha *et al.*, 2000)<sup>10</sup>. Specimens were kept in polypropylene troughs each with 7-8 individuals/50 L of water. Water was kept O<sub>2</sub> saturated by aeration. The troughs were cleaned daily and the water as well as the pesticide was renewed to keep the concentration constant throughout the test period of 24, 48, 72 h. Control fish were kept in dechlorinated tap water without any treatment. Fish were fed commercial fish feed daily at least one hour prior to the replacement of water. Only healthy, active fish starved for 24hr were used for the experiment and they were allowed no food during treatment procedures.

### Insecticide chemical and dose selection

The commercial grade of phorate was obtained from G.M.Shah pesticides, Srinagar; manufactured from Cyanamid, India Ltd. (Bombay). On the basis of literature data (LC<sub>50</sub> values for phorate), three sub-lethal concentrations (0.2ppm, 0.4ppm and 0.6ppm) of phorate were then selected for the experiment.

### Experimental design

Group I (Control): One group of fish from each sub-species was selected as control, kept in chlorinated water without any treatment. They were fed once daily commercial fish feed.

Group II: The fish sample from two sub-species was subdivided into three sub groups each, based on the dose selection of the pesticide. All subgroups had equal number of fishes (five fish/group/duration) maintained in 50litre/polypropylene troughs. After treatment with pesticide, the frequencies of micronuclei in all experimental sub-groups were examined at three durations (24, 48 and 72h) and at each concentration.

### Cytogenetic studies using micronucleus test

The method of Schmid (1975) was used. The fish were injected with 0.1 ml of 0.025% colchicine and sacrificed two hours later with a slight blow on the head region. Chemical treated and control fish were

cut in the caudal. From the freshly collected blood, smears were made on grease free slides. After fixation using methanol as fixative, slides were stained with Mayer's hematoxylin, rinsed in Scott's tap water substitute, followed by another staining in eosin (Pascoe and Gatehouse, 1986). After completion of this staining process, the slides were then washed in 30%, 50%, 70% and 90% alcohol, cleaned in xylene and mounted using D.P.X. The slides were then examined using a simple light microscope. For each concentration and duration, five fish specimens were used and from each fish, six slides were studied and 1200cells were scored under 600x magnification.

### Statistical Analysis

Statistical analysis of data to verify the significant difference in the incidence of micronuclei between treated and control groups at 0.05 and 0.01 level of significance was performed using non-parametric criteria, Mann-Whitney U test to analyze the frequency of micronuclei.

### Results

The genotoxicity of phorate in *Cyprinus carpio specularis* and *Cyprinus carpio communis* was confirmed by incidence of micronucleus in peripheral erythrocytes after 24, 48 and 72 hours. Three sub-lethal concentrations of phorate, 0.2 ppm, 0.4 ppm and 0.6 ppm were used, and it was observed that all these concentrations were able to induce micronucleus formation in erythrocytes of both the fish species. No increased incidence of micronucleated erythrocytes of *Cyprinus carpio specularis* and *Cyprinus carpio communis* was reported with sub lethal concentration of 0.2 ppm after 24-hour intervals. However, after 48h and 72h, a statistically significant increase was found in the frequency of micronuclei in peripheral erythrocytes of both fish species. The percentage of single micronuclei in *Cyprinus carpio specularis* (0.03±0.01 of control) increased to 1.15±0.32 from low to high concentrations after 24 h and continued to increase by 1.78±0.30 and 2.74±0.52 in longer exposures table 1 and table 2).

In *Cyprinus carpio communis* somewhat similar results were observed with increase in percentage of single micronuclei (0.03±0.01 control) to 1.30±0.23 at 24 h from low to high concentration and this percentage continued to increase by 2.08±0.31 and 2.91±0.39 after 48 and 72 h respectively.

Statistical analysis showed significant difference in the frequency of incidence of micronuclei in the erythrocytes of control and frequency of micronuclei in the erythrocytes of phorate treated groups of *Cyprinus carpio specularis* and *Cyprinus carpio communis* respectively (Mann-Whitney U test; P< 0.05). Both dose and time dependent increase in the

micronuclei frequency was observed in treated fish species and a peak value detected at higher concentrations after 72h of phorate injection, clearly showed a higher incidence of micronucleated peripheral erythrocytes.

### Discussion

Organophosphate insecticides are ubiquitous environmental contaminants because of their wide applications in agriculture. It is known that in fishes organophosphate pesticides are neurotoxic and they inhibit acetylcholinesterase activity with subsequent disruption of nervous functions, thereby interfering with some of the vital physiological functions (Rao and Rao, 1983). Since organophosphate pesticides are finding increasing use in recent years, this can result in acute and long term side effects, including sickness and death of people, useful animals, fish, birds and destruction of crops.

In the present study, positive genotoxic effects, measured as micronucleus frequency in erythrocytes from both fish species (*Cyprinus carpio specularis* and *Cyprinus carpio communis*) exposed to different doses of phorate were observed. The result of the

present study revealed a significant induction of micronuclei in peripheral erythrocytes ( $p < 0.01$  and  $p < 0.05$ ) of both fish species. The appearance of inter-specific differences observed in the present study could be attributed to the specificity of DNA repair, cell turnover time, physiological peculiarities, contaminant uptake or biotransformation in the fish species studied.

On the other hand age, sex, reproductive status, genetic constitution may affect micronucleus frequency in fish (Al-Sabti *et al.*, 1994). However, *Cyprinus carpio specularis* and *Cyprinus carpio communis* in the present study were selected from the same age groups. The exposure was performed in the laboratory under standard experimental conditions. Therefore, the inter-specific differences in micronuclei incidences should not be attributed to the intrinsic problems of the experimental system used.

In the present study a significant difference in the micronucleus incidence among treated and control groups was observed. The peak frequency of micronucleated erythrocytes was observed at 72h after exposure.

**Table 1:** Micronucleus frequencies (%) in peripheral blood erythrocytes of *Cyprinus carpio specularis* exposed to different concentrations of phorate

Treatment	Concentration	MN frequencies (%)		
		24 h	48 h	72 h
		Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD
Control	-	0.03 $\pm$ 0.01	0.05 $\pm$ 0.01	0.05 $\pm$ 0.01
Phorate	0.2 ppm	0.06 $\pm$ 0.03	0.09 $\pm$ 0.03*	0.23 $\pm$ 0.03*
	0.4 ppm	0.34 $\pm$ 0.11*	0.54 $\pm$ 0.15*	0.88 $\pm$ 0.12*
	0.6 ppm	1.15 $\pm$ 0.32*	1.78 $\pm$ 0.30*	2.74 $\pm$ 0.52*

(Mann-Whitney U test) \*  $P < 0.05$

**Table 2:** Micronucleus frequencies (%) in peripheral blood erythrocytes of *Cyprinus carpio communis* exposed to different concentrations of phorate

Treatment	Concentration	MN frequencies (%)		
		24 h	48 h	72 h
		Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD
Control	-	0.03 $\pm$ 0.01	0.04 $\pm$ 0.01	0.05 $\pm$ 0.01
Phorate	0.2 ppm	0.04 $\pm$ 0.01	0.08 $\pm$ 0.03*	0.28 $\pm$ 0.01*
	0.4 ppm	0.39 $\pm$ 0.085*	0.67 $\pm$ 0.06*	1.05 $\pm$ 0.26*
	0.6 ppm	1.29 $\pm$ 0.23*	2.08 $\pm$ 0.31*	2.91 $\pm$ 0.39*

(Mann-Whitney U test) \*  $P < 0.05$

The length of cell cycle critical to micronuclei formation depends upon the time needed to replicate DNA and perform nuclear division. In man and mice the duration of the cell cycle has been well documented. There is, however, little information on the duration of the cell cycle in the tissues of teleost species since the cell cycle varies with temperature in poikilotherms (Al-Sabti and Metcalfe, 1995). The incubation times were chosen without the benefit of specific knowledge of the times required for the division of fish cells (Al-Sabti, 1994). A time dependent increase in the incidence of micronuclei in peripheral erythrocytes of *Cyprinus carpio specularis* and *Cyprinus carpio communis* was established and confirms other observations (Al-Sabti, 1986; Nepomuceno and Spano, 1995; Nepomuceno *et al.*, 1997; Gustavino *et al.*, 2001; Buschini *et al.*, 2004). Increased chromosome aberrations and micronuclei were observed in bone marrow cells of rats that received (Dhingra *et al.*, 1990). Phorate in the present study was also found to induce micronuclei in peripheral erythrocytes of fish. The carbohydrate metabolism was found to be adversely affected by phorate in the serum of fresh water fish *Clarius batrachus* (Jyothi and Narayan, 1999). Decreased total serum levels and plasma ChE activity was observed in beagle dogs treated with phorate (Piccirillo *et al.*, 1987).

The present study reveals that micronucleus assay has a great potential for detecting clastogenic substances in aqueous media. However, additional experimental evidence is needed to evaluate these hypotheses. Also further studies on aquatic organisms exposed in-vivo as well as in-vitro to various chemicals is certainly needed to clarify the mechanism of micronucleus formation.

#### References

- Hayashi M, Veda T, Uyeno K, Wada K, Kinai, Saotome K, Tanaka N, Takai A, Sasaki YF, Asano N, Sofuni T, Ojima Y. Development of genotoxicity assay systems that use aquatic organisms. *Mutat Res.* 1998, 399: 125-133.
- Pacheco M, Santos MA (1998) Induction of liver EROD and erythrocyte nuclear abnormalities by cyclophosphamide and PAHs in *Anguilla Anguilla* L. *Ecotoxicol. Environ. Saf.* 1998, 40: 71-76.
- Powers DA. Fish as model system. *Sci.* 1989, 246: 352-358.
- Al-Sabti K, Hardig J. Micronucleus test in fish for monitoring the genotoxic effects of the industrial waste products in the Baltic Sea, Sweden. *Comp. Bioch Physiol.* 1990, 97C: 179-182.
- Matsumoto FE, Colus IMS. Micronucleus frequencies in *Astyanax bimaculatus* (characidae) treated with cyclophosphamide or vinblastine sulphate. *Gen. Mol. Biol.* 2000, 23: 489-492.
- Porto JIR, Araujo CSO, Feldberg E. Mutagenic effects of mercury pollution as revealed by micronucleus test on three Amazonian fish species. *Environ. Res.* 2005, 97: 287-292.
- Pantaleao S de M, Alcantara AV, Alves JdoPH, Spano MA. The piscine micronucleus test to assess the impact of pollution on the Japarutuba river in Brazil. *Environ Mol Mutagen.* 2006, 47.
- Choy WN, MacGregor JT, Shelby MD, Maranpot RR. Induction of micronuclei by benzene in B6C3FI mice: retrospective analysis of peripheral blood smears from NTP Carcinogenesis bioassay. *Mutat Res.* 1985, 143: 55-59.
- Heddle JA, Hite M, Kirkhart B, Mavourim K, Mac-Gregor jt, Newell GW, Salamon MF. The induction of micronuclei as a measure of genotoxicity. *Muta. Res.* 1983, 123: 61-118.
- Anitha B, Chandra N, Gopinath PM, Durairaj G. Genotoxicity evaluation of heat shock in gold fish (*Carassius auratus*). *Mutat Res.* 2000, 469: 1-8.
- Schmid W. The micronucleus test. *Mutat Res.* 1975, 31: 9-15.
- Pascoe S, Gatehouse D. The use of a simple haematoxylin and eosin staining procedure to demonstrate micronuclei within rodent bone marrow. *Mutat Res.* 1986 164: 237-234.
- Rao KSP, RKV Rao. Regulation of phosphorylases and aldolases in tissues of the teleost (*Tilapia mossambica*) under methyl parathion impact. *Bull Environ Contam Toxicol.* 1983, 31: 427-478.
- Al-Sabti K, Frank M, Andrijanic B, Knez S, and Stegnar P. Chromium induced micronuclei in fish. *J Appl Toxicol.* 1994, 14: 333-336.
- Al-Sabti K, Metcalfe CD. Fish micronuclei for assessing genotoxicity in water. *Mutat Res.* 1995, 343: 121-135.
- Al-Sabti K. Micronuclei induced by selenium, mercury, methyl mercury and their mixtures in binucleated blocked fish erythrocyte cells. *Mutat Res.* 1994, 320: 157-163.
- Al-Sabti. Comparative micronucleated erythrocyte cell induction in three cyprinids by five carcinogenic-mutagenic chemicals. *Cytobios.* 1986, 47: 147-154.
- Nepomuceno JC, Spano MA. Induction of micronuclei in peripheral erythrocytes of *Cyprinus carpio* fish by methyl parathion.

- International Contaminant Ambient*.1995, 11: 9-12.
19. Nepomuceno JC, Ferrari I, Spano MA, Centeno AJ. Detection of micronuclei in peripheral erythrocytes of *Cyprinus carpio* exposed to metallic mercury. *Environ Mol Mutagen*.1997, 30 (3): 293-297.
  20. Gustavino B, Scornajehgi KA, Minissi S, Ciccotti E. Micronuclei induced in erythrocytes of *Cyprinus carpio* (teleostei, pisces) by X-rays and colchicine. *Mutat Res*. 2001, 494: 151-159.
  21. Buschini A, Martino A, Gustavino B, Manfrinotte M, Poli P, Rossi C, Santoro M, Door AJM, Rizzoni M. Comet assay and micronucleus test in circulating erythrocytes of *Cyprinus carpio* specimens exposed in situ to lake waters treated with disinfectants for potabilization. *Mutat Res*.2004, 557: 119-129.
  22. Dhingra AK, Grover IS, Adhikari N. Chromosomal aberration and micronuclei assays of some system pesticides in bone marrow cells. *Nucleus*.1990, 33: 14-19.
  23. Jyothi B, Narayan G. Certain pesticide induced carbohydrate metabolic disorders in the serum of Fresh water fish *Clarias batrachus* (Linn.). *Fd chem Toxic*.1999, 37: 417-421.
  24. Piccirillo VJ, Schellenberger TE, Dauvin EM, 1987. 14-day range-finding oral toxicity a study in the dog with AC 35, 024. Laurel MD, USA: Tegeris laboratories, Inc, 1987; revised final rep. no. 85013; unpublished report, cited in FAO 1995.

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