

24 SAMPLES AT A TIME  
8 RUNS PER DAY

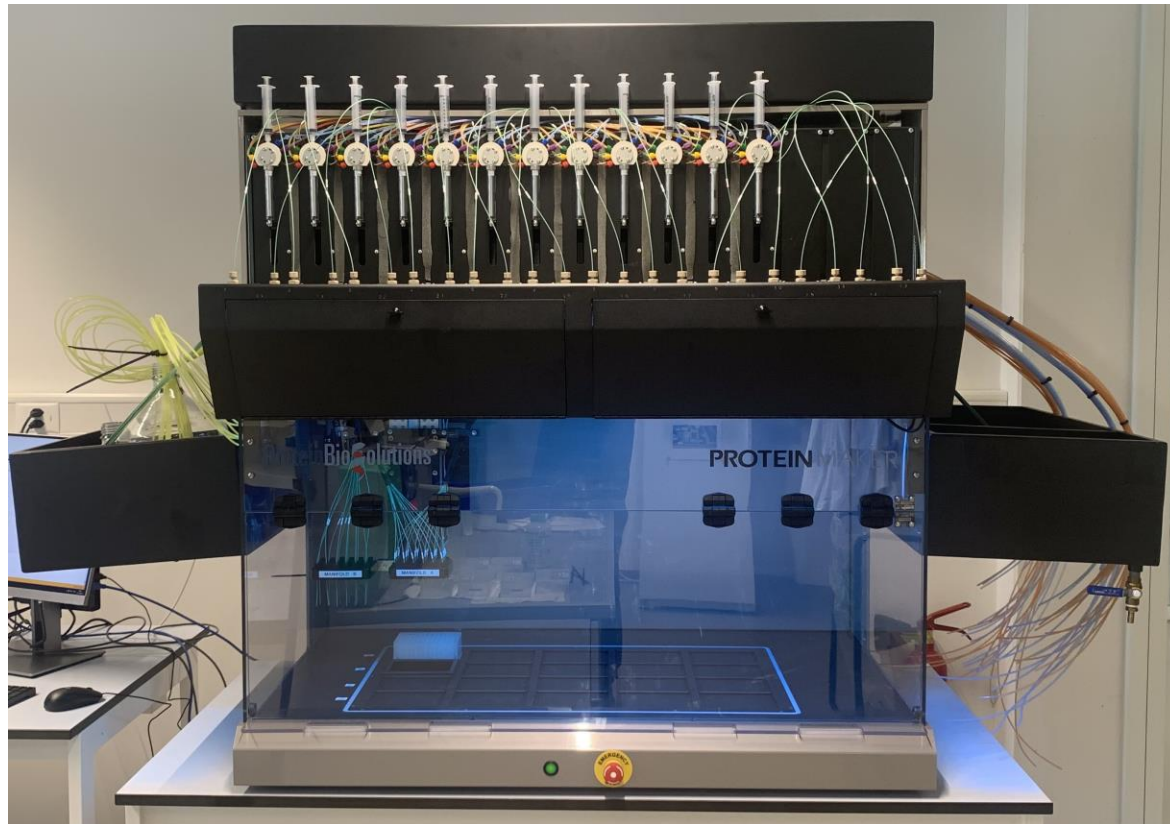
RELIEVE YOUR  
BOTTLENECKS



192 PURIFICATIONS  
PER DAY

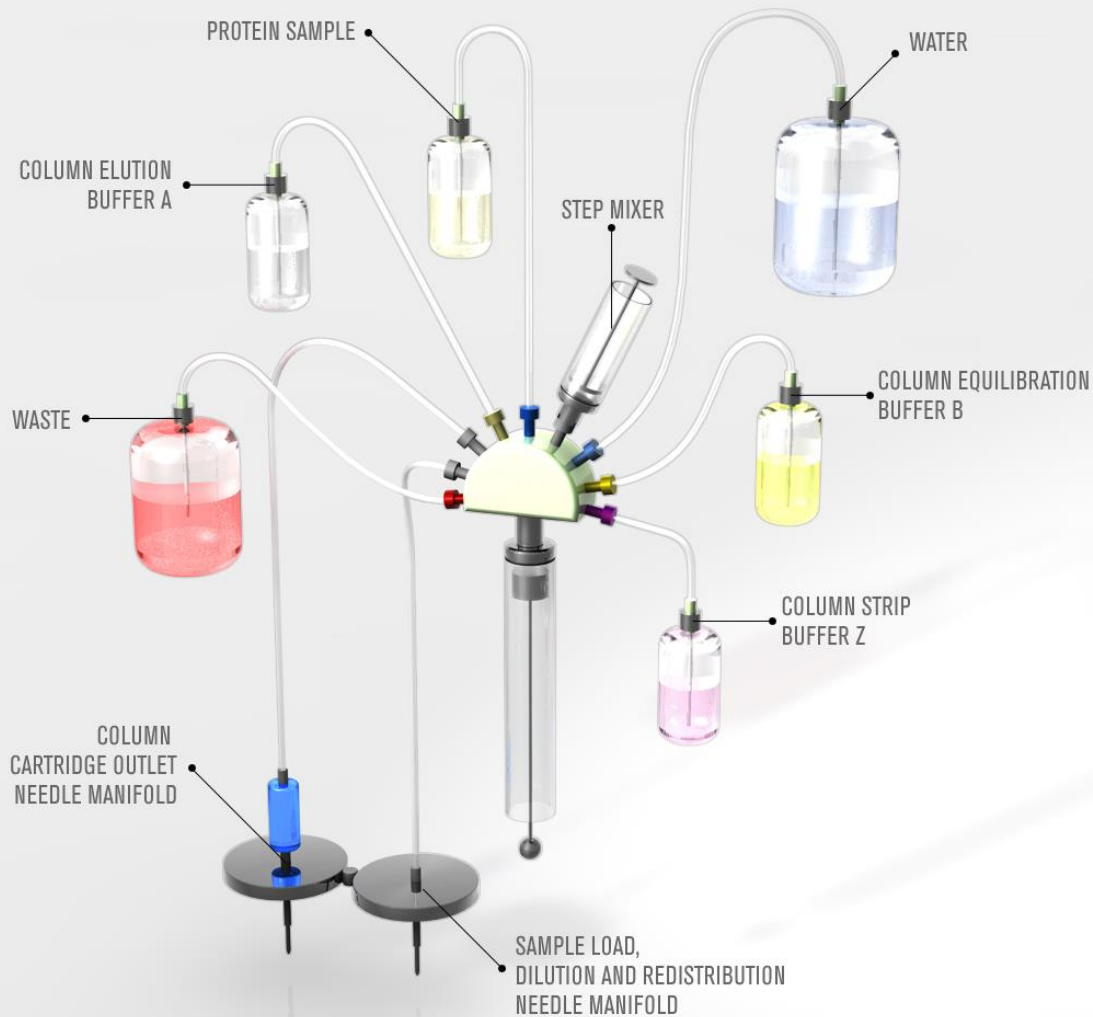
# Overview

- Purify 1 to 24 samples in parallel
- Eliminate known and unknown (!) sample degradation
- mg+ protein production scale
- Flexible sample volume scales from a few mL to multiple liters



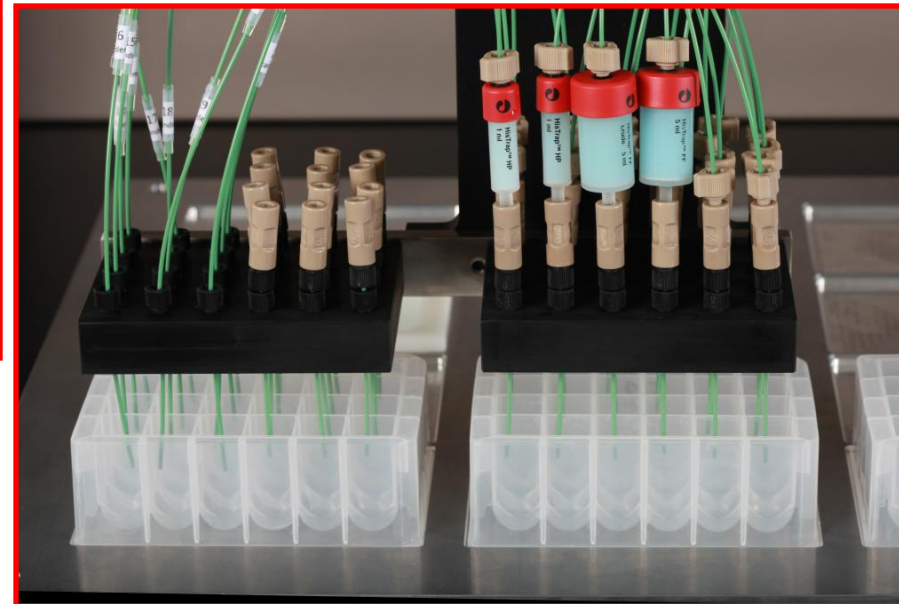
# Overview

- 24 independent flow paths
- 9-port valve configuration
- Compatible with common commercial columns

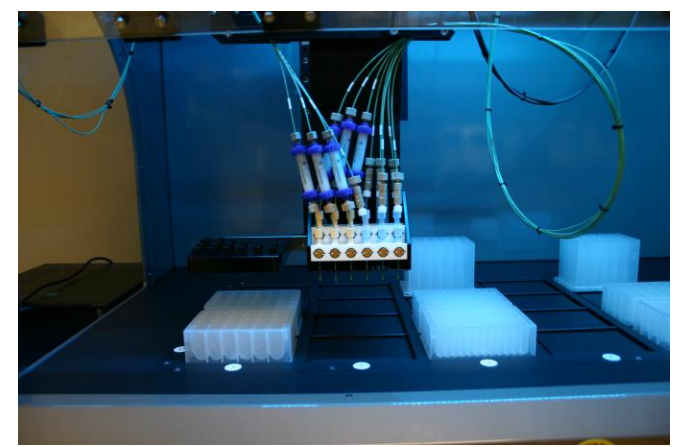


# Overview

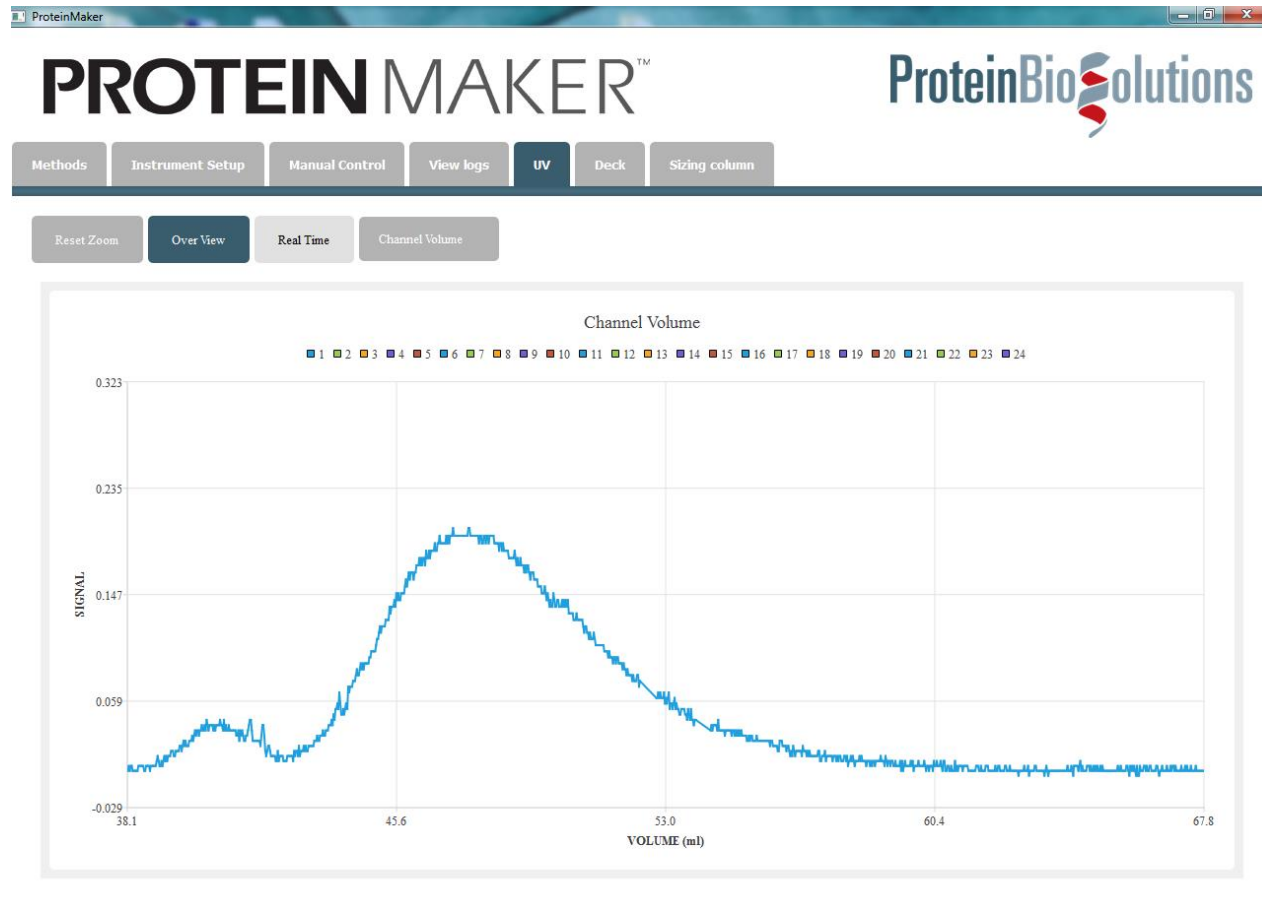
- 20 SBS deck positions (1 waste)
- Walk-away automation
- Perform parallel, 2-step purification of 12 samples



# Flexible column sizes and locations

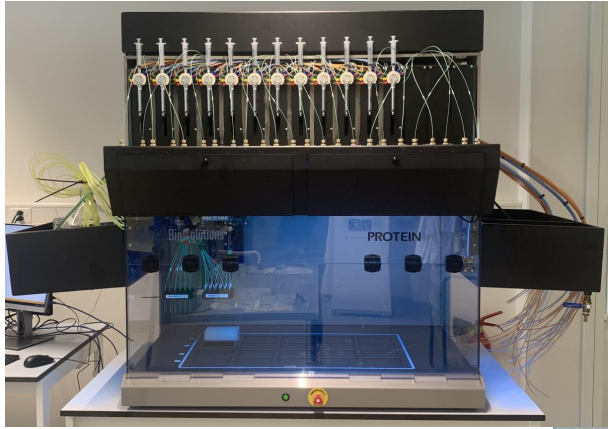


# Now with optional 24 channel UV monitoring system!



# Individual UV Flow cell block for each of 24 channels:







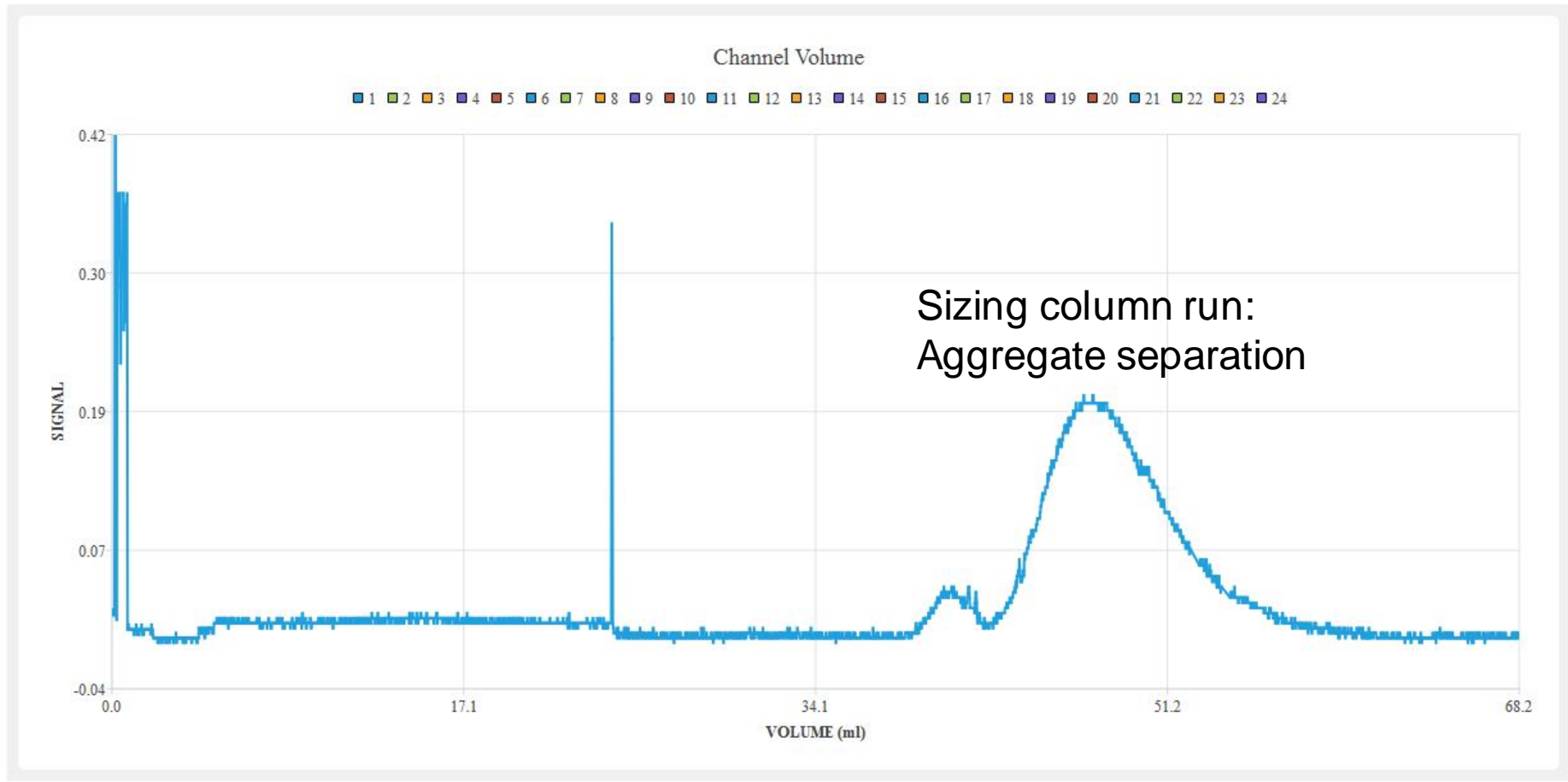


# PROTEIN MAKER™



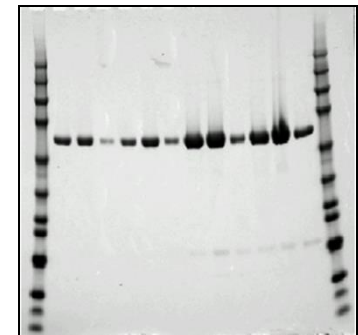
- Methods
- Instrument Setup
- Manual Control
- View logs
- UV**
- Deck
- Sizing column

- Reset Zoom
- Over View**
- Real Time
- Channel Volume



# Protein Maker™: Purification Automation

- High-throughput Protein Purification
  
- Operation Modes:
  1. Production Mode (up to 24 proteins, 1 resin)
  2. Scouting Mode (up to 24 resin types, 1 protein)
  3. Multi-column (automated 2 step purification)
  
- Common Applications / Uses:
  1. Antibody Production
  2. Crystallography Prep
  3. No Risk Scale-up



*12 protein variants purified in parallel*

# Production Mode

## Antibody production/screening:

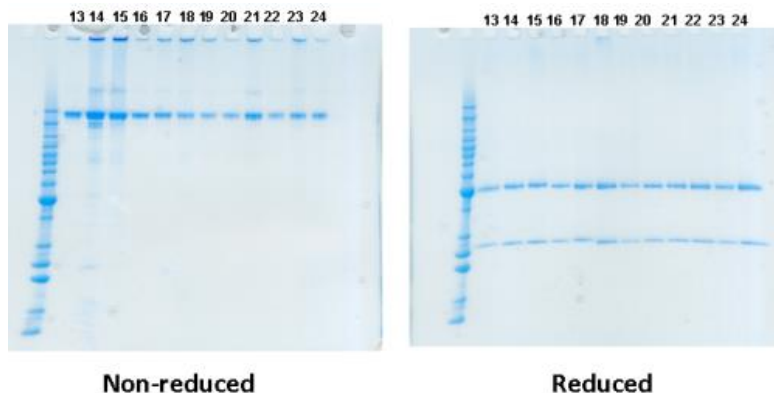
Purify mg+ amounts of up to 24 antibodies in parallel using up to or more than 1L

### Case Study: Purification of Engineered Antibodies

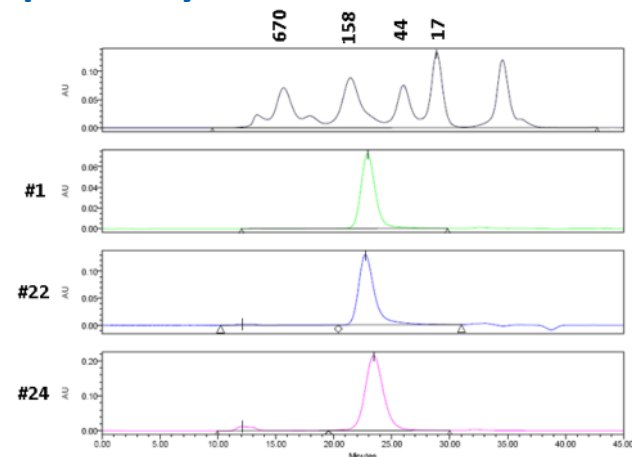
- Had: Fully humanized antibody against a viral antigen target, with high affinity for wild type target but no affinity to a prevalent naturally occurring variant
- Goal: improve affinity for variant without loss of affinity for wild type

Purified 50mL each for 24 mutations (expression in CHO cells) using 1 mL HiTrap Protein A column. Yield (2-10 mg) provided sufficient material for biophysical characterization (analytical SEC/LS, SDS-PAGE, DSC) as well as affinity measurement.

#### Example SDS-PAGE: Mutants 13-24



#### Example analytical SEC: Mutants 1, 22, and 24



# Production Mode

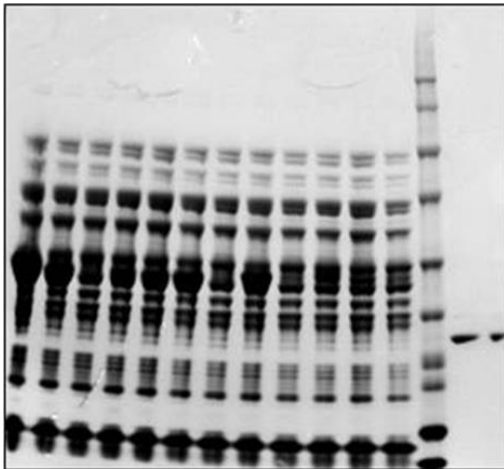
## Production of multiple constructs:

Parallel processing enables scaled-up purification of multiple constructs (internal deletion variants) to access those with high crystallizability

### Case Study: Multi-construct design and purification

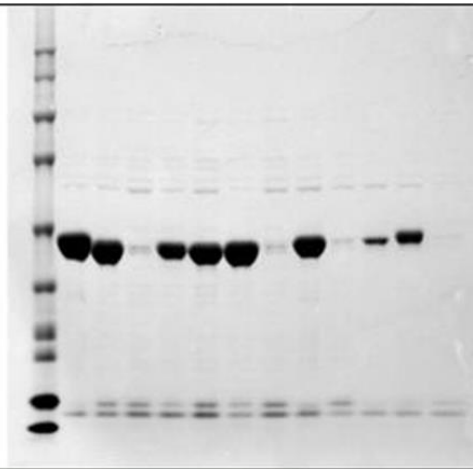
#### Lysates

1 2 3 4 5 6 7 8 9 10 11 12



#### Eluates

1 2 3 4 5 6 7 8 9 10 11 12



#### Crystals



Genes optimized for *E. coli* expression  
 Expression yields: 0.1 to 15mg per liter cell culture

# Production Mode

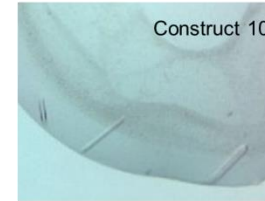
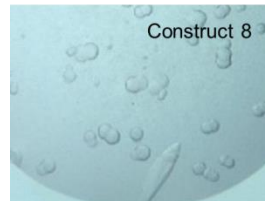
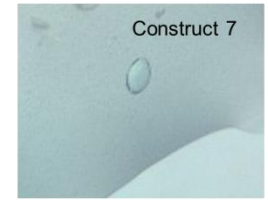
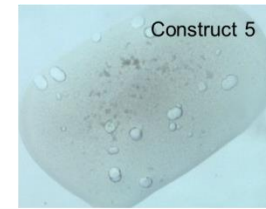
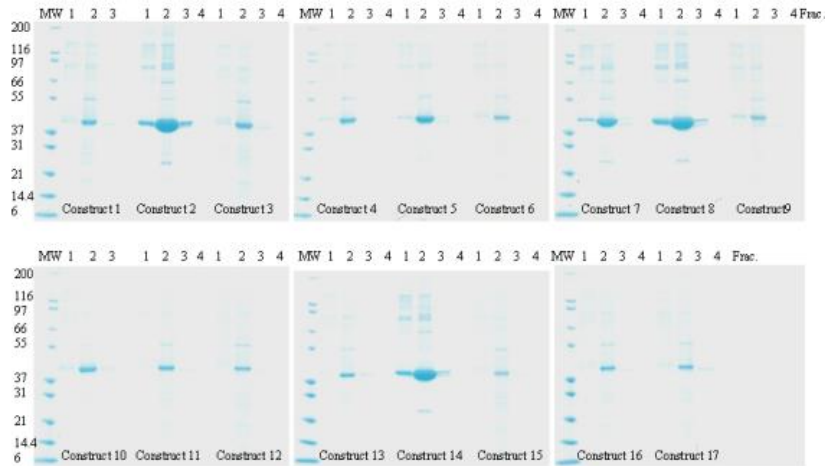
## Production of multiple constructs:

Parallel processing enables more efficient route to protein crystals

### Case Study: Production of Kinase X – 17 constructs

Parallel nickel affinity purification  
(4-fraction elution)

9 constructs selected for his-tag  
removal and crystal screening



# Production Mode

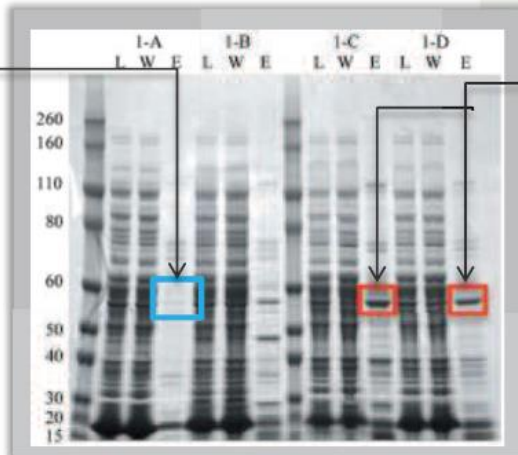
## Parallel Lysis scouting:

Test 12 different lysis buffer conditions followed by small scale IMAC purification to scale up and move forward with the strong purifiers

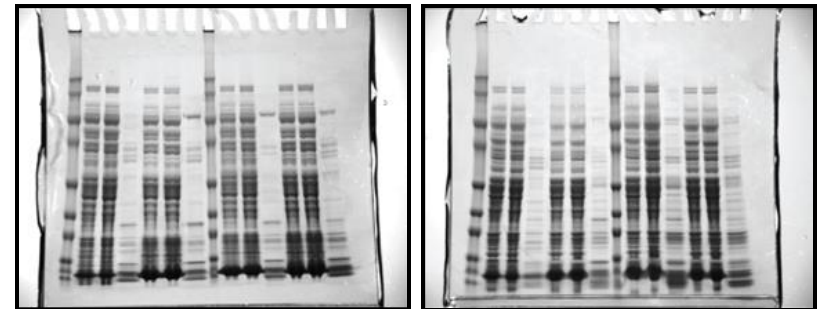
### Case Study: Parallel testing of Lysis Conditions

**Example:** Cytochrome P450 (CYP51A) insoluble in standard lysis buffer

**Result:** of 12 lysis buffers 2 yield soluble CYP51A (CHAPS/BOG + 500 mM NaCl)



Conditions:	Low Salt	High Salt	Detergent 1	Detergent 2
pH 6.0	50mM MES	50mM MES	50mM MES	50mM MES
	250mM NaCl	1M NaCl	500mM NaCl	500mM NaCl
	5% Glycerol	5% Glycerol	5% Glycerol	5% Glycerol
pH 7.5	50mM HEPES	50mM HEPES	50mM HEPES	50mM HEPES
	250mM NaCl	1M NaCl	500mM NaCl	500mM NaCl
	5% Glycerol	5% Glycerol	5% Glycerol	5% Glycerol
pH 8.0	50mM TRIS	50mM TRIS	50mM TRIS	50mM TRIS
	250mM NaCl	1M NaCl	500mM NaCl	500mM NaCl
	5% Glycerol	5% Glycerol	5% Glycerol	5% Glycerol
	0.5mM TCEP	0.5mM TCEP	0.5mM TCEP	0.5mM TCEP
			1% CHAPS	1% BOG

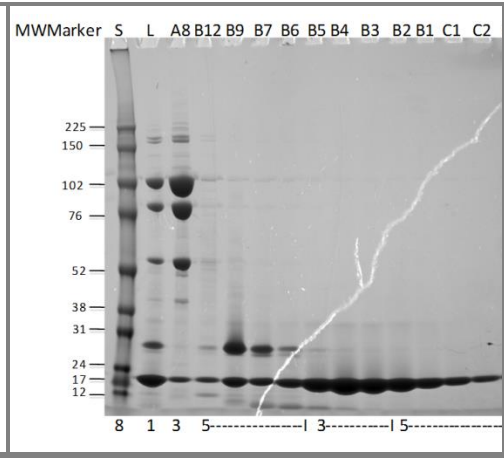
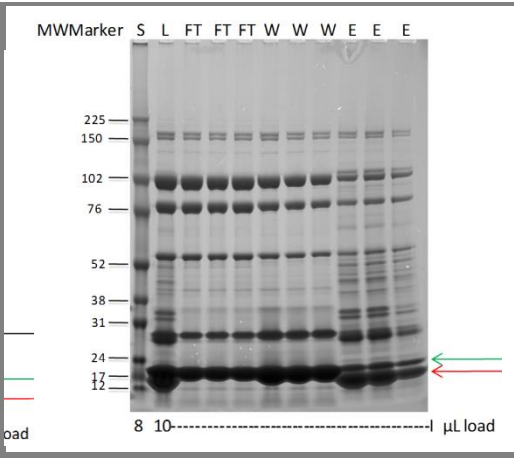
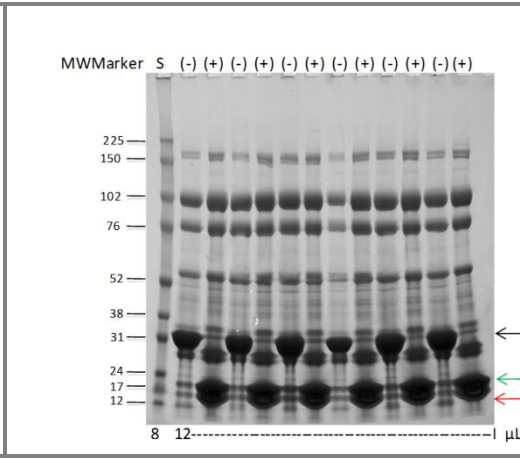
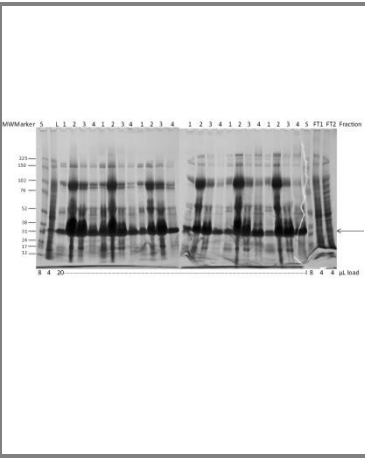


Acta Cryst. (2011). F67, 1015–1021

# Production Mode

## Risk-free Scale-up via parallelization:

### 6 x 5 mL Column Bed Volume Parallel Purification

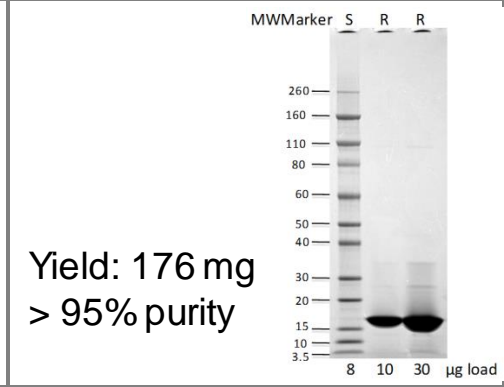
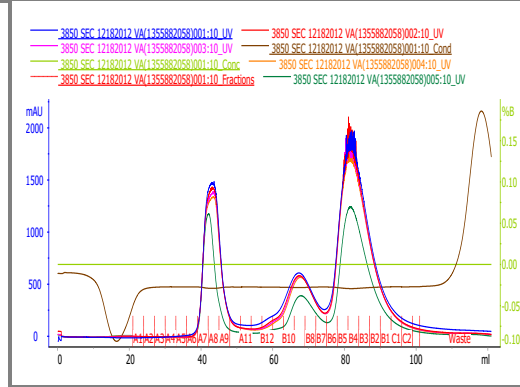


## Situation

- Increased purification from 20 mg to > 100 mg

## Options

- Develop protocol for 5 x scale up
- Repeat 5-6 x of the original purifications
- Run multiple small-scale purifications in parallel – Protein Maker



No need for traditional “scale-up” if you use Protein Maker



# Scouting Mode

## Parallel resin scouting:

Scouting a variety of resins (up to 24) for optimization of 1 protein

### Case Study: Scouting Mode with a Step-Gradient Protocol

#### Glu-PGS – Antibody affinity column

- A. Equilibration Buffer: 20 mM Tris pH 8, 100mM NaCl, 0.5% NP40
- B. Elution Buffer: Equilibration buffer 1 plus 50  $\mu$ M EYMPTD peptide

#### HiTrap SP Sepharose – Cation exchange resin

- A. Equilibration Buffer: 20 mM MES pH 6.0
- B. Elution Buffer: Equilibration buffer plus 1 M NaCl

#### HiTrap Q Sepharose – Anion exchange resin

- A. Equilibration Buffer: 20 mM Tris pH 8
- B. Elution Buffer: Equilibration buffer 1 plus 1 M NaCl

#### Heparin Sepharose – Cation exchange resin

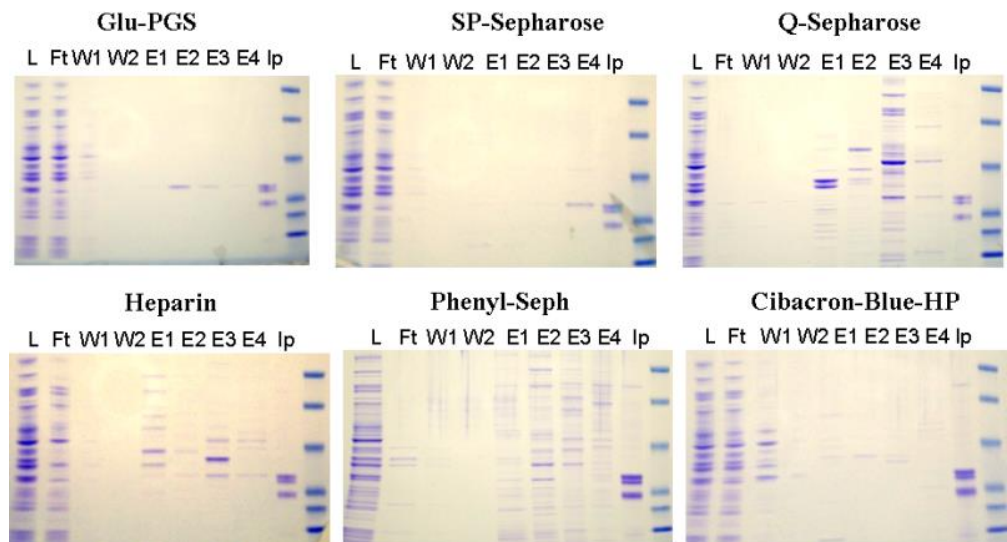
- A. Equilibration Buffer: 20 mM MES pH 6.0
- B. Elution Buffer: Equilibration buffer plus 1 M NaCl

#### HiTrap Phenyl Sepharose – hydrophobic interaction

- A. Equilibration Buffer: 20 mM Tris pH 8, 1M NH<sub>4</sub>SO<sub>4</sub>
- B. Elution Buffer: 5 mM Tris pH 8

#### HiTrap Blue Sepharose - affinity column

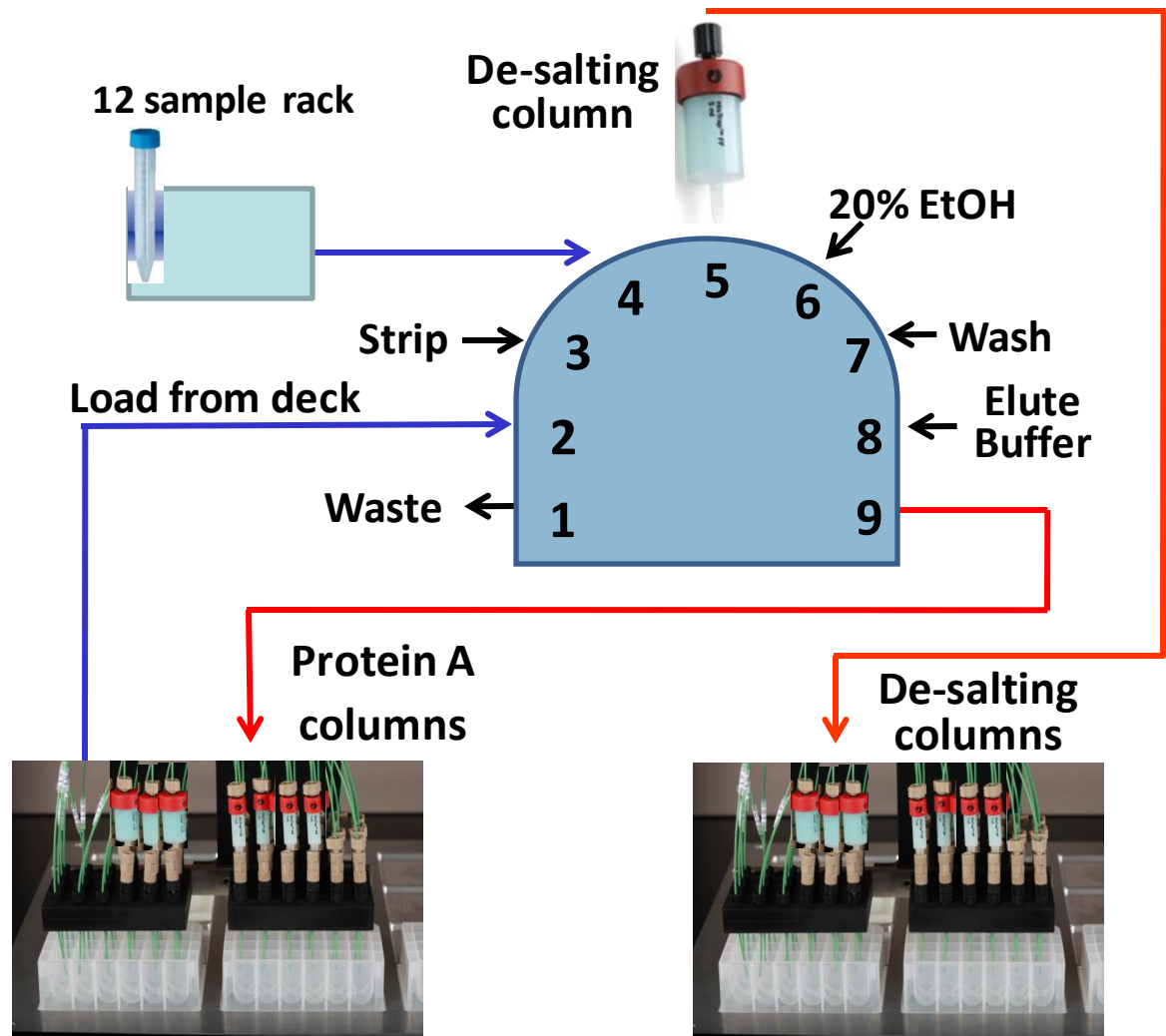
- A. Equilibration Buffer: 20 mM Tris pH 8
- B. Elution Buffer: Equilibration buffer 1 plus 1 M NaCl



# Multi-Column purification

## 2-column Purification Schematic:

- 12 Protein A columns (1 ml)
- 12 Desalting columns (5 ml)
- Load volume: 50 ml (from sample rack)
- Run time: 2.5 h unattended operation
- Run includes column regeneration



# Complex Purification Mode

## 2-step Purification:

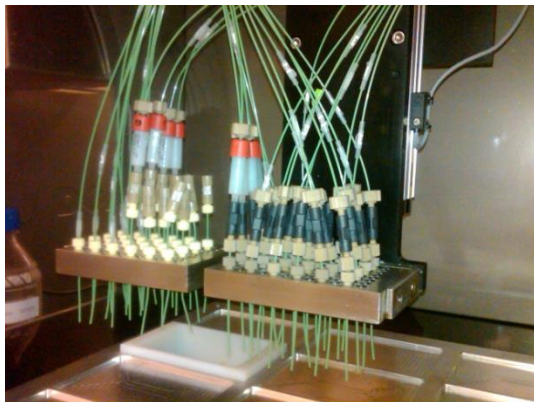
### Ni and Protein A 2-step Fab purification

#### Case Study: Need medium scale throughput of 2-step protocol

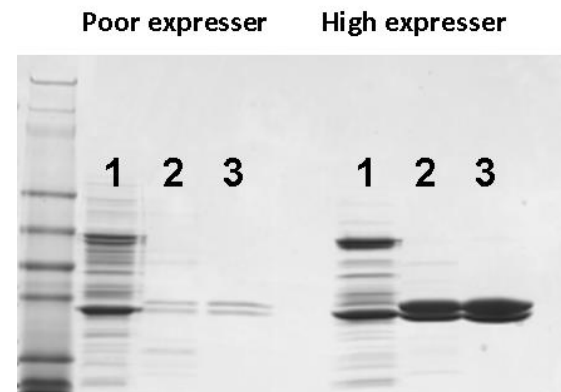
- Ni purification (GE His-Trap FF) of His-tagged Fabs alone requires additional purification, particularly for low-producing proteins, but generally results in high yield
- Ni requires buffer exchange post purification
- Protein A (GE Hi-Trap FF) is added in series to increase purity and remove requirement for buffer exchange

#### Customized Potential Protocol:

- Syringes 1-12: Load Ni, wash, and elute with 200 mM imidazole-containing buffer
- Syringes 13-24: Load Ni eluate, wash, elute with pH 2.8 buffer with immediate post-column neutralization



**Syringes 1-4 His-Trap FF (right); syringes  
13-16 Hi-Trap Protein A FF (left)**



**1: cell-free extract  
2: Ni-purified  
3: Ni and Protein A-purified**

# Customer Feedback

## OPPORTUNITY COSTS

**Make the best use of your time, money and equipment:** *“The Protein Maker™ enables us to make the best use of our time and purification equipment by saving our single channel systems for complex purification experiments while the Protein Maker™ quickly works through our routine and/or high-throughput purifications.”*

**Shrink your project timelines:** *“The parallelization of the Protein Maker™ enables one technician to purify more protein constructs in one day than could be done in a week by that same person. The practical result is that we can isolate the desired protein construct in four weeks faster on average for every project. That is extremely valuable!”*

**Enable new strategies:** *“Parallel purification of 12-24 protein samples enables new and otherwise impractical screening and scouting techniques that we wouldn’t attempt without the Protein Maker™ and have lead to key successes.”*

## INTANGIBLES

**Pleasing your colleagues, clients and project leaders:** *“Just knowing that we have this capability in house gives me the confidence to tell the end users of the proteins we produce that I can get them their pure protein in two weeks...when they would expect it to take six or more.”*