

## *Genetic Engineering Laboratory Protocol*

### **Preparation of Competent Cells**

**Aim:** Preparation of Competent Cells using *E.coli* DH5 $\alpha$  cells

**Principle:** The ability of taking the DNA by a bacterial cell is called Competence. *E.coli* cells can be made competent chemically by adding CaCl<sub>2</sub> (or) MgCl<sub>2</sub>. These cells are able to take foreign DNA (recombinant plasmids/ amplicons). The bacterial cell surface is negatively charged and the DNA molecule is also negatively charged as a result, the bacteria cannot take the foreign DNA in to it. The addition of CaCl<sub>2</sub> the net charge (or) the charge of the cell wall and the foreign DNA is neutralized, and promotes the opening of Ca<sup>2+</sup> ion channels present on the surface of bacterial cells. Thus promotes the easy entry of foreign DNA inside the bacterial cell.

**Requirements:** *E.coli* DH5 $\alpha$  cells, Luria Bertani medium, CaCl<sub>2</sub> (0.1M, filter sterile).

#### **L.B medium composition:**

1. a. Bactotryptone: 10.0 g /L  
b. Yeast Extract: 5.0 g/l  
c. Sodium Chloride: 10.0 g/l.  
pH: 7.0  $\pm$  0.2 (Make up the pH by NaOH (or) HCl).
2. One can use readymade L.B medium for preparing L.B Broth (for preparing the media follow the instruction of manufacturer)

#### **Procedure:**

1. **Day 1:** Inoculate single colony of *E.coli* DH5 $\alpha$  cells to 3.0 ml of autoclaved L.B broth in test tube and incubate at 37°C, 180 rpm for 12-16 hours.
2. **Day 2:** 500  $\mu$ L of overnight culture inoculate to 25 ml of Autoclaved L.B broth in conical flask and incubate at 37°C, 180 rpm for 2-3 hours (or) till log phase is attained (or) the cell O.D reaches to 0.4-0.6.

3. After O.D reaches to 0.4-0.6 , take out the flask from incubator and keep in ICE for 10 min
4. Transfer the above pre cooled E.coli DH5 $\alpha$  cells in to 50 ml centrifuge tubes and do centrifugation at 3,500 rpm for 15 min at 4°C.
5. Decant the supernatant without disturbing the pellet. Now resuspend the above pellet in 3.0 ml of CaCl<sub>2</sub> (0.1M).
6. Keep the above mixture on ice for 45 min. Centrifuge the above suspension at 3,500 rpm, for 10 min at 4°C.
7. Decant the supernatant and resuspend in 1 ml of 20% Glycerol solution prepared in 0.1M CaCl<sub>2</sub> .Aliquot 200  $\mu$ L of competent cells into five sterile centrifuge tubes (sterile) and store the above competent cells at -80°C, for transformation.