

MicroRNA let-7b inhibits Melanoma cell migration

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Abstract: Objective: Melanoma, the most dangerous form of skin cancer, develops when unrepaired DNA damage to skin cells triggers mutations that lead the skin cells to multiply rapidly and form malignant tumors. The aim of this study was to evaluate the role of Micro RNA Let-7b in melanoma cell migration. **Materials and Methods:** Melanoma specimens were obtained from 30 patients with skin melanoma. The human melanoma cell line A375 was used as an in vitro model for melanoma. In the present study, the wound healing assay and real-time PCR were carried out to assess the migration ability, and to determine mRNA transcription profiles, respectively. **Result:** Results from this study showed that let-7b expression is able to significantly inhibit A375 melanoma cell migration. The wound healing assay demonstrated that A375 cell motility increases after let-7b inhibitor treatment. **Conclusion:** Let-7b displayed a higher ability to inhibit melanoma cell migration and invasion, compared to other members of the Let-7 family.

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Key words: Let-7b, Melanoma, A375 cells

Introduction:

Melanoma is a type of cancer that develops from pigment-containing cells known as melanocytes(1). Melanomas typically occur in the skin but may rarely occur in the mouth and intestines(2). The legs and back are the most commonly involved sites in women and men, respectively(2). This type of cancer occasionally develops from a mole with worrying changes, including an increase in size and color, irregular edges, itchiness or skin breakdown(3). Cancer is a multi-step process resulting from a gradual accumulation of genetic alterations caused by DNA damage(4). Thymine-thymine dimers are the most type of DNA damage caused by ultraviolet A (UVA) light(3). More importantly, UVA produces reactive oxygen species (ROS) that induces DNA damage, including single-strand breaks, oxidized pyrimidine and oxidized purine 8-oxoguanine(2). It was shown that MicroRNAs (miRNAs) play an important role in cancer etiology and are frequently deleted in solid cancers including melanoma(5). miRNAs are small noncoding RNAs

encoded in the human genome as miRNA genes, composing of either a single miRNA species (monocistronic) or several different miRNAs (polycistronic)(5). MiR155, MiR146 and Let7 are the most miRNAs involved in melanoma(6). The Let-7 miRNA precursor was first identified in a study of developmental timing in *C. elegans*; however, it was later shown to be part of a much larger class of non-coding RNAs termed microRNAs(7). Importantly, Let-7 is a highly attractive potential therapeutic option with the ability to prevent tumor genesis and angiogenesis, typically in cancers(7). Let-7 has been demonstrated to be a direct regulator of RAS expression in human cells(8). All three RAS genes in human, including K-, N-, and H-ras, have the predicted let-7 binding sequences in their 3'UTRs(8). In samples from lung cancer patients, expression of RAS and let-7 showed a reciprocal pattern, in which cancerous cells have decreased let-7 and increased RAS; however, normal cells have increased let-7 and decreased RAS(2). Another oncogene, high mobility group A2 (HMGA2),

has also been identified as a target of let-7(9). Interestingly, Let-7 directly inhibits HMGA2 by binding to its 3'UTR. Removal of the let-7 binding site by 3'UTR deletion leads to HMGA2 overexpression and tumor formation(9). In light of this, the purpose of this study was to investigate the Let-7b role in melanoma cell migration.

Material and Methods:

Patients and specimens

A total of 30 melanoma samples were collected from patients treated in the Sina Hospital, Tabriz University of Medical sciences, Tabriz, Iran. Tumors from primary sites were immediately snap-frozen and stored at -80°C .

Cell culture

The human melanoma cell line A375 was purchased from National Cell Bank of Iran (NCBI), Pasteur Institute of Iran, Tehran, Iran, and passaged in our laboratory for fewer than 6 months after resuscitation. The cells were cultured in RPMI 1640 medium (Cellgro, Manassas, VA, USA) supplemented with 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA, USA) and 1% penicillin/streptomycin (Invitrogen, Carlsbad, CA, USA). The cell cultures were maintained in a humidified air chamber at 37°C with 5% CO_2 .

Migration and invasion assays

A375 human melanoma cells were transfected with let-7b mimic (60 nM), let-7b inhibitor (90 nM), and control oligos. Forty-eight hours following transfection, culture media were removed and the cells were serum-starved for 24 hours. Seventy-two hours after transfection, cells were trypsinized, suspended in serum-free medium, and then seeded into Transwell Permeable Support inserts with 8- μm micro-porous membranes (Corning, Corning, NY, USA) at a density of 2×10^5 cells per well in a 24-well plate. Culture medium containing 10% FBS was in the lower compartment of the plate. Six hours after culture, the nonmigrating cells on the upper surface of the membrane were removed using a cotton swab. The cells that migrated to the lower surface were fixed and stained with the 3-step stain kit (Richard-Allan Scientific, Kalamazoo, MI, USA). The stained cells were photographed and counted with a hemocytometer under a microscope.

Wound healing assays

A375 human melanoma cells were seeded at a density of 1×10^6 cells per well into six-well plates and grown overnight. Cells were transfected with let-7b mimic (60 nM) and control oligos. After 48 hours, the cells were serum starved for additional 24 hours. Afterwards, a

sterile 200 μl pipette tip was applied to scratch the cells to form a wound. The cells were washed twice with PBS and then cultured in RPMI medium containing 10% FBS. Migration of the cells to the wound was observed and photographed using an inverted phase-contrast microscope equipped with digital camera system (Olympus). The healing rate was quantified by measuring the gap size after the culture. Eight different areas in each assay were chosen to evaluate the distance of the migrating cells to the origin of the wound. The healing rate of cells transfected with let-7b miRNA was normalized to that of control cells in a given experiment. The healing rate was used to represent cell migration.

Real-time PCR

Total RNA was extracted from the samples using TRIZOL reagent (Invitrogen, Carlsbad, CA, USA), and reverse-transcribed using a high capacity RNA-to-cDNA kit (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's recommendation. The cDNA was quantified by RT-PCR using an ABI Prism 7900 sequence detection system (Applied Biosystems, Foster City, CA, USA). PCR amplification was carried out using SYBR Green PCR core reagents (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's instructions. PCR amplification of the housekeeping gene B-actin was carried out as a control for each sample.

Statistical analysis

All data were represented as mean \pm SD. Statistical analysis was conducted using the Student's test. A P value less than 0.05 was considered to be statistically significant.

Results:

let-7b levels in melanoma patients, and its effect on A375 human melanoma cell migration

A decrease in let-7b levels was found in melanoma patients. let-7b was able to inhibit A375 human melanoma cell migration. To determine the clinical significance of the let-7 family in melanoma, the expression of all let-7 family members was also analyzed in a total 30 melanoma specimens using real-time RT-PCR. Results from real-time PCR summarized in Fig. 1A indicate the lower expression levels of let-7b in melanoma cells compared to other members of the family. Moreover, it should be noted that B-actin was used as a control in this study, as shown in Fig. 1A. The results strongly suggested that decreased expression of let-7 is an important step in melanoma development. To examine the effect of let-7b on A375 melanoma cell migration and invasion, the let-7b mimic or control were transfected into A375 melanoma cells. The migration and invasion abilities of

transfected cells were examined 48 hours after transfection. Our results demonstrated that let-7b expression was able to significantly inhibit A375 melanoma cell migration ($p < 0.005$; Figure 1B) and invasion ($p < 0.005$; Fig. 1C) in vitro.

The let-7b inhibitor was used to block endogenous let-7b expression. Results from the loss-of-function experiment demonstrated that the let-7b inhibitor significantly increases A375 melanoma cell migration

($p < 0.01$; Figure 1D) and invasion ($p < 0.01$; Figure 1E) in vitro.

The effect of *let-7b* in melanoma cell motility

Cell motility is a factor that can impact cell migration in cancer. Results from the wound healing assay demonstrated that A375 cell motility increases after let-7b inhibitor treatment (Figure 2), emphasizing the inhibitory role of let-7b in melanoma cell motility.

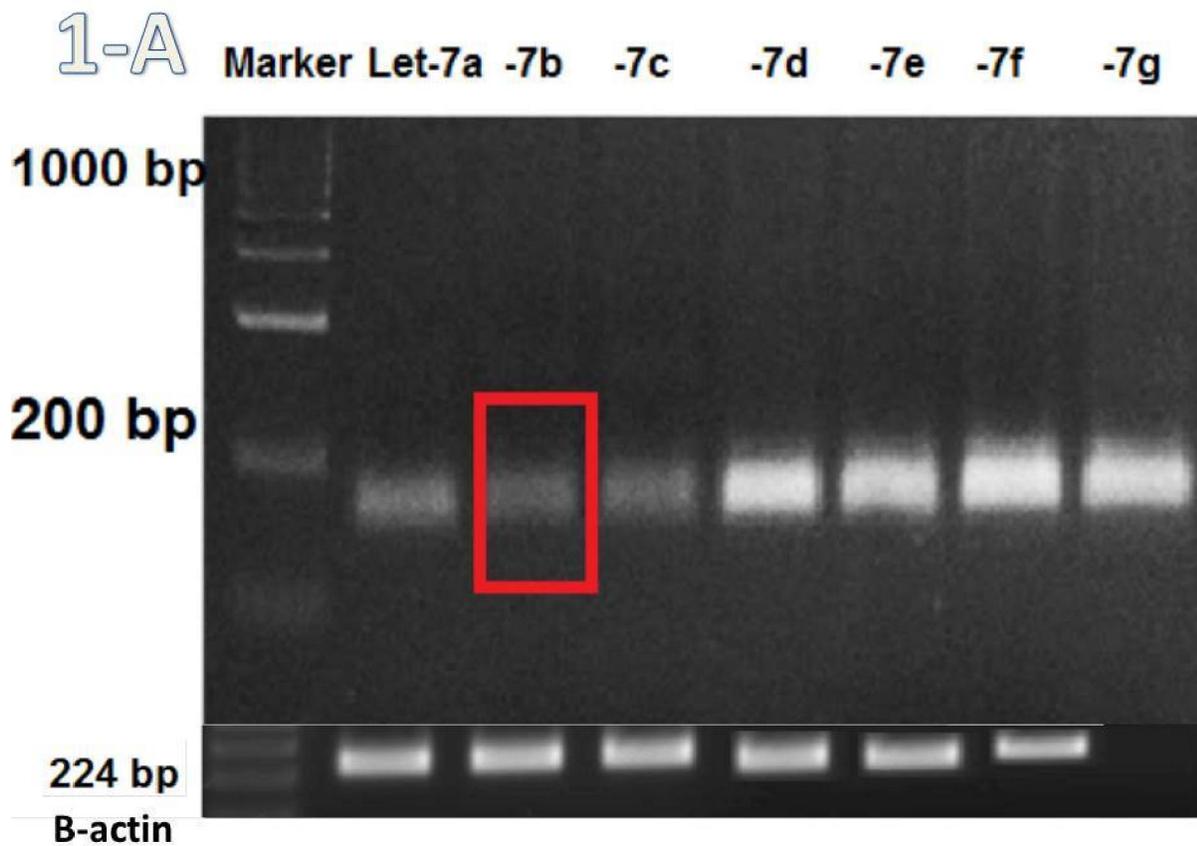


Figure 1A.

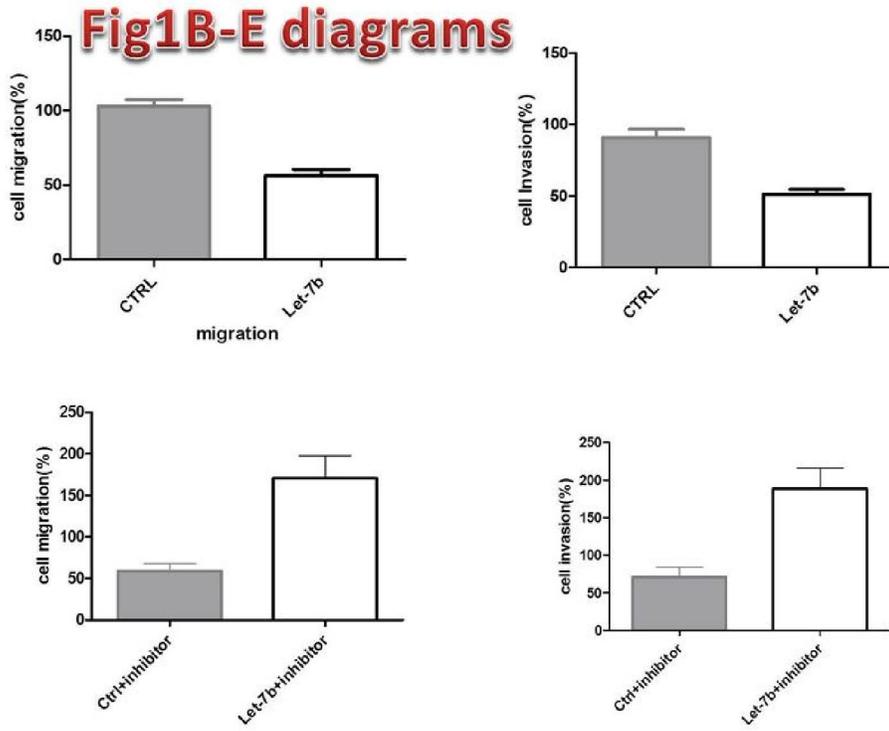


Figure 1B.

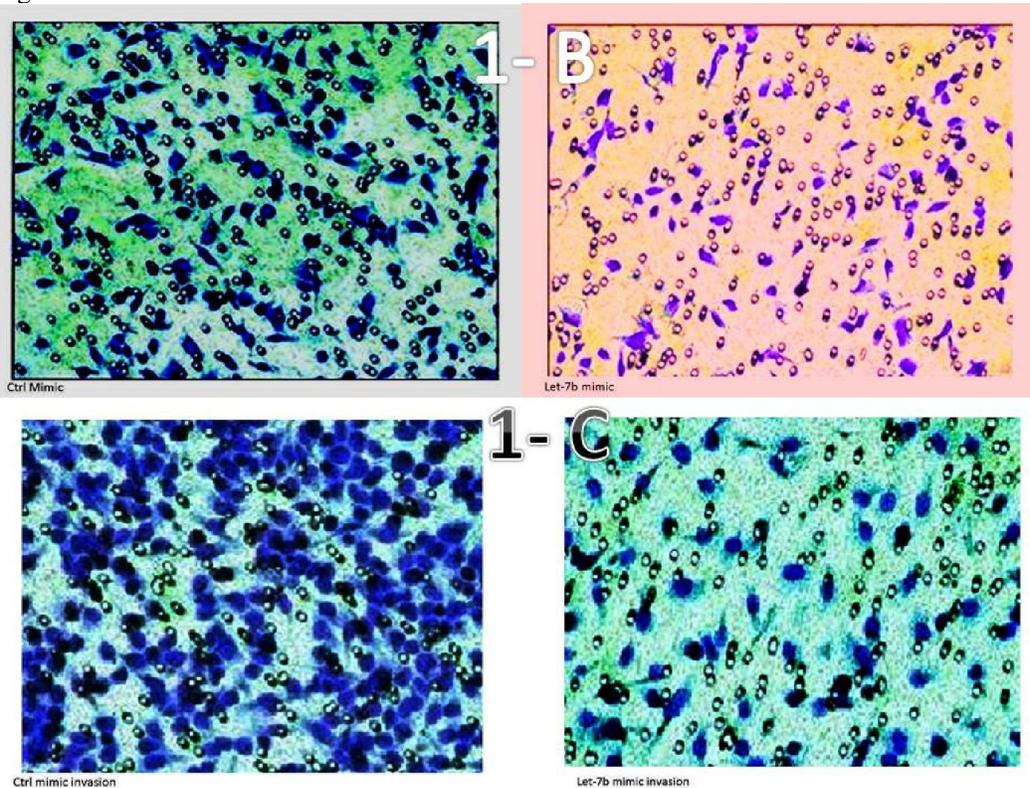


Figure 1C.

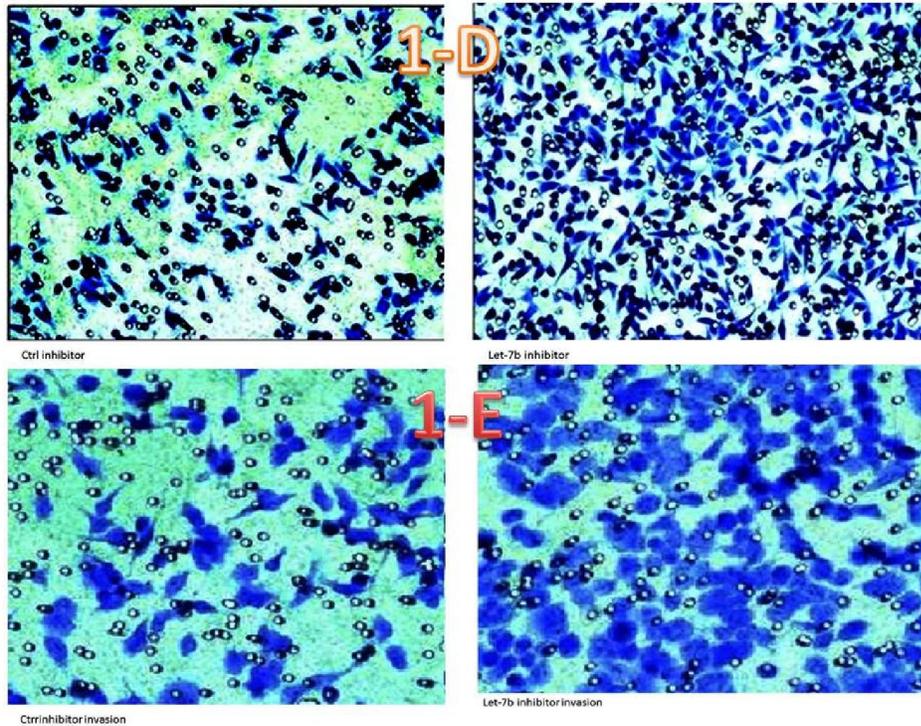


Figure 1D.

Figure 1:A) The PCR imaging of Let-7 family expression: As shown the let7-b expression is less than other family. B-C) Cell migration and invasion percentages and results: let-7b expression was able to significantly inhibit A375 melanoma cell migration ($p < 0.005$; Figure 1B) and invasion ($p < 0.005$; Fig. 1C) D-E) Cell migration and invasion percentages with using inhibitor: let-7b inhibitor significantly increases A375 melanoma cell migration ($p < 0.01$; Figure 1D) and invasion ($p < 0.01$; Figure 1E)

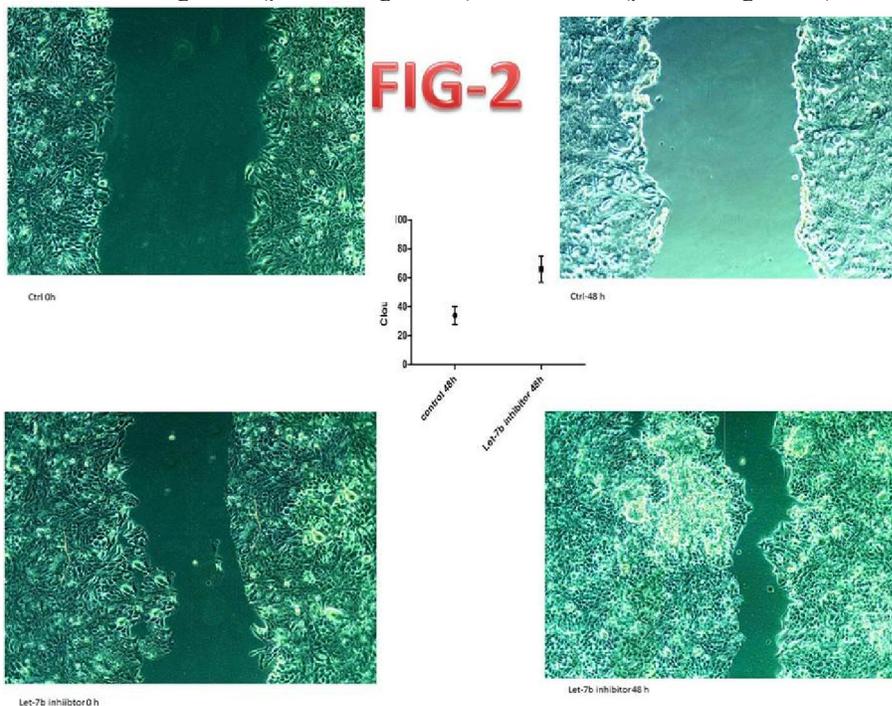


Figure 2.

Figure 2: wound healing assay result: A375 cell motility increases after let-7b inhibitor treatment

Discussion:

Melanoma is the rarest form of skin cancer, accounting for only about 4% of skin cancer cases but for as many as 74% of all skin cancer deaths(10). Melanoma is more likely to invade nearby tissues and spread to other parts of the body than other types of skin cancer(11). MiRNAs Let7 is an important tumor suppressor gene in human. The Let 7 family is a large family with thirteen members in human genome(7). Most studies demonstrated that let-7a is down-regulated in the tumor metastasis; however, few studies showed that there is a relation between let-7b and tumor growth(12). There is evidence that Let-7 can regulate various steps of the metastasis cascade in cancer cells(12). Our results reveal that let-7b could suppress melanoma cell migration and motility. Consistent with our results, a variety of studies showed the same function of let-7a in other cancers(13). The authors stated that only let-7e, among let-7 families, has no managing role in cancer metastasis. Results from the present study demonstrated that cell motility increases after let-7b inhibitor treatment. Taken together, the results from this investigation indicate that the Let-7 family, particularly Let-7b, could inhibit cell migration in malignant melanoma.

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Running title: Let-7b in Melanoma cell migration

References:

- [1]. Correa ZM. Assessing Prognosis in Uveal Melanoma. *Cancer control : journal of the Moffitt Cancer Center*. 2016 Apr;23(2):93-8. PubMed PMID: 27218785. Epub 2016/05/25. Eng.
- [2]. Pennisi A, Bloisi DD, Nardi D, Giampetruzzi AR, Mondino C, Facchiano A. Skin lesion image segmentation using Delaunay Triangulation for melanoma detection. *Computerized medical imaging and graphics : the official journal of the Computerized Medical Imaging Society*. 2016 May 7. PubMed PMID: 27215953. Epub 2016/05/25. Eng.
- [3]. Kanavy HE, Gerstenblith MR. Ultraviolet radiation and melanoma. *Seminars in cutaneous medicine and surgery*. 2011 Dec;30(4):222-8. PubMed PMID: 22123420. Epub 2011/11/30. eng.
- [4]. Pfarr N, Penzel R, Klauschen F, Heim D, Brandt R, Kazdal D, et al. Copy Number Changes of Clinically Actionable Genes in Melanoma, Non-Small Cell Lung Cancer and Colorectal Cancer - A Survey Across 822 Routine Diagnostic Cases. *Genes, chromosomes & cancer*. 2016 May 24. PubMed PMID: 27218826. Epub 2016/05/25. Eng.
- [5]. Ortiz-Quintero B. Cell-free microRNAs in blood and other body fluids, as cancer biomarkers. *Cell proliferation*. 2016 Jun;49(3):281-303. PubMed PMID: 27218664. Epub 2016/05/25. Eng.
- [6]. Sha J, Gastman BR, Morris N, Mesinkovska NA, Baron ED, Cooper KD, et al. The Response of microRNAs to Solar UVR in Skin-Resident Melanocytes Differs between Melanoma Patients and Healthy Persons. *PloS one*. 2016;11(5):e0154915. PubMed PMID: 27149382. Pubmed Central PMCID: PMC4858311. Epub 2016/05/07. eng.
- [7]. Damanakis AI, Eckhardt S, Wunderlich A, Roth S, Wissniewski TT, Bartsch DK, et al. MicroRNAs let7 expression in thyroid cancer: correlation with their deputed targets HMGA2 and SLC5A5. *Journal of cancer research and clinical oncology*. 2016 Jun;142(6):1213-20. PubMed PMID: 26960757. Epub 2016/03/11. eng.
- [8]. Fores-Martos J, Cervera-Vidal R, Chirivella E, Ramos-Jarero A, Climent J. A genomic approach to study down syndrome and cancer inverse comorbidity: untangling the chromosome 21. *Frontiers in physiology*. 2015;6:10. PubMed PMID: 25698970. Pubmed Central PMCID: PMC4316712. Epub 2015/02/24. eng.
- [9]. Zhang P, Huang C, Fu C, Tian Y, Hu Y, Wang B, et al. Cordycepin (3'-deoxyadenosine) suppressed HMGA2, Twist1 and ZEB1-dependent melanoma invasion and metastasis by targeting miR-33b. *Oncotarget*. 2015;6(12):9834-53. PubMed PMID: 25868853. Pubmed Central PMCID: PMC4496401. Epub 2015/04/15. eng.
- [10]. Yamada S, Tsukamoto S, Huang Y, Makio A, Kumazoe M, Yamashita S, et al. Epigallocatechin-3-O-gallate up-regulates microRNA-let-7b expression by activating 67-kDa laminin receptor signaling in melanoma cells. *Scientific reports*. 2016;6:19225. PubMed PMID: 26754091. Pubmed Central PMCID: PMC4709792. Epub 2016/01/13. eng.
- [11]. Murakami M, Suzuki M, Nishino Y, Funaba M. Regulatory expression of genes related to metastasis by TGF-beta and activin A in B16 murine melanoma cells. *Molecular biology reports*. 2010 Mar;37(3):1279-86. PubMed PMID: 19288218. Epub 2009/03/17. eng.

- [12]. Akbari A, Ghahremani MH, Mobini GR, Abastabar M, Akhtari J, Bolhassani M, et al. Down-regulation of miR-135b in colon adenocarcinoma induced by a TGF-beta receptor I kinase inhibitor (SD-208). Iranian journal of basic medical sciences. 2015 Sep;18(9):856-61. PubMed PMID: 26523217. Pubmed Central PMCID: PMC4620183. Epub 2015/11/03. eng.
- [13]. Albino D, Civenni G, Dallavalle C, Roos M, Jahns H, Curti L, et al. Activation of the Lin28/let7 axis by loss of ESE3/EHF promotes a tumorigenic and stem-like phenotype in prostate cancer. Cancer research. 2016 May 2. PubMed PMID: 27197175. Epub 2016/05/20. Eng.

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